

# **Heterogeneous distribution of sex ratio distorters in natural populations of the isopod *Armadillidium vulgare***

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## Abstract

In the isopod *Armadillidium vulgare*, many females produce progenies with female-biased sex ratios, due to two feminizing sex ratio distorters (SRD): *Wolbachia* endosymbionts and the *f* element. We investigated the distribution and population dynamics of these SRD and mitochondrial DNA variation in 16 populations from Europe and Japan. Confirming and extending results from the 1990's, we found that the SRD are present at variable frequencies in populations, and that the *f* element is overall more frequent than *Wolbachia*. The two SRD never co-occur at high frequency in any population, suggesting an apparent mutual exclusion. We also detected *Wolbachia* or the *f* element in some males, which likely reflects insufficient titer to induce feminization or presence of masculinizing alleles. Our results are consistent with a single integration event of a *Wolbachia* genome in the *A. vulgare* genome at the origin of the *f* element, which contradicts an earlier hypothesis of frequent losses and gains. We identified strong linkage between *Wolbachia* strains and mitochondrial haplotypes, but no association between the *f* element and mitochondrial background. Our results open new perspectives on SRD evolutionary dynamics in *A. vulgare*, the evolution of genetic conflicts and their impact on the variability of sex determination systems.

## Keywords

Sex ratio distorter, endosymbiont, *Wolbachia*, *f* element, sex determination

## 1. Introduction

Sex ratio distorters (SRD) are selfish genetic elements located on sex chromosomes or transmitted by a single sex, which skew the proportion of males and females in progenies towards the sex that enhances their own vertical transmission [1]. Major SRD types include sex chromosome meiotic drivers [2,3], some B chromosomes [4] and selfish cytoplasmic genetic entities [5,6,7]. Collectively, they are found in a wide range of animal and plant species and they have had a tremendous impact on the ecology and evolution of their host species [8,9]. One of the most emblematic SRD is the bacterial endosymbiont *Wolbachia* [10,11]. *Wolbachia* is a cytoplasmic, maternally inherited alpha-proteobacterium found in a wide range of arthropods and nematodes. In arthropods, *Wolbachia* often manipulates host reproduction in favor of infected females, thereby conferring itself a transmission advantage. This is achieved through various strategies, three of which causing sex ratio distortions towards females: male killing, thelytokous parthenogenesis and feminization of genetic males [6,7,10,11].

In the terrestrial isopod *Armadillidium vulgare*, chromosomal sex determination follows female heterogamety (ZZ males and ZW females) [12–14]. However, many females produce progenies with female-biased sex ratios, due to the presence of two feminizing SRD: *Wolbachia* endosymbionts and a locus called the *f* element [6,15,16]. *Wolbachia* symbionts cause ZZ genetic males to develop as phenotypic females [17]. Three *Wolbachia* strains have been described in *A. vulgare*, for which feminization induction has been demonstrated (*wVulC* and *wVulM* strains [18,19]) or is strongly suspected (*wVulP* strain [20]). The *f* element is a nuclear insert of a large portion of a feminizing *Wolbachia* genome in the *A. vulgare* genome [21]. The *f* element induces female development, as a W chromosome does, and it shows non-Mendelian inheritance, making it an SRD [21,22]. These SRD may cause turnovers in sex determination mechanisms [6,15,23] and they could explain why sex chromosome systems are so variable in terrestrial isopods [24–27].

Testing this hypothesis requires characterizing the evolutionary dynamics of SRD such as *Wolbachia* and the *f* element in natural populations. In *A. vulgare*, this characterization is quite limited because prior studies were mostly restricted to a narrow geographic area (western France), sometimes focusing solely on *Wolbachia* [20,28–31]. The only exception is a 1993 study [32], which collated and extended results from the early 1980's [33,34]. The main observations were that *Wolbachia* and the *f* element are present at variable frequencies in field populations, and the *f* element is more frequent than *Wolbachia*. However, earlier studies were limited by the lack of molecular tests for *Wolbachia* and/or the *f* element, preventing any direct assessment of SRD presence. Instead, the authors used a

complex, indirect procedure combining a physiological test and crossings [32]. In addition to being tedious and time-consuming (generation time is one year in this species), this procedure did not allow direct and undisputable assessment of SRD presence. Moreover, it could only be run on females and therefore provided no information on SRD presence in males. Finally, it could not reveal individuals potentially carrying both SRD.

Here, we took advantage of the availability of molecular markers to directly assess SRD presence in males and females from *A. vulgare* field populations from Europe and Japan. This approach allowed us to circumvent the limitations of previous studies, and to revisit the population dynamics of *Wolbachia* and the *f* element in this species and their association mitochondrial lineages.

## 2. Materials and Methods

*A. vulgare* individuals from 16 natural populations across Europe and Japan were collected by hand. Individuals were sexed and stored in alcohol or at -20°C prior to DNA extraction. Total genomic DNA was extracted from the head and legs of each individual, as described previously [21]. We used four molecular markers to assess the presence of *Wolbachia* and the *f* element in DNA extracts: *Jtel* [21], *wsp* [35], *recR* [36] and *ftsZ* [37] (Table S1). While *Jtel* is specific to the *f* element, *wsp* and *recR* are specific to *Wolbachia*, and *ftsZ* is present in both the *f* element and *Wolbachia* [21]. We assessed the presence or absence of these markers by PCR, as described previously [21]. Different amplification patterns were expected for individuals with *Wolbachia* only (*Jtel*-, *wsp*+, *recR*+, *ftsZ*+), the *f* element only (*Jtel*+, *wsp*-, *recR*-, *ftsZ*+), both *Wolbachia* and the *f* element present (*Jtel*+, *wsp*+, *recR*+, *ftsZ*+) or both *Wolbachia* and the *f* element lacking (*Jtel*-, *wsp*-, *recR*-, *ftsZ*-). The few individuals exhibiting other amplification patterns were classified as “undetermined status”. A quantitative-PCR assay was used to measure *Wolbachia* titer in some individuals (see supplementary Methods). To characterize *Wolbachia* strain diversity, *wsp* PCR products were purified and Sanger sequenced using both forward and reverse primers by GenoScreen (Lille, France). Forward and reverse reads were assembled using Geneious® v.7.1.9 to obtain one consensus sequence per individual. To evaluate mitochondrial diversity, we amplified by PCR a ~700 bp-long portion of the Cytochrome Oxidase I (*COI*) gene in all individuals [38]. PCR products were purified and Sanger sequenced as described above. Haplotype network analysis was performed using the *pegas* package [39]. All statistical analyses were performed with R v.3.6.0 [40]. Figures were realized with *ggplot2* [41].

### 3. Results

We tested the presence of *Wolbachia* and the *f* element in 423 females and 224 males from 16 populations across Europe and Japan (Tables 1, S2). While most males lacked both SRD, 48% of females carried at least one of them. The remaining females presumably carry W chromosomes, although the existence of other feminizing elements cannot be formally excluded. As expected for feminizing elements, the SRD were mostly found in females, the *f* element being more frequent than *Wolbachia* overall. Both SRD were found in the same individuals in only 3 females from a single population (Chizé). *Wolbachia*-infected individuals carried one of the three previously known *Wolbachia* strains of *A. vulgare*: wVulC (n=62), wVulM (n=23) or wVulP (n=4).

*Wolbachia* and *f* element distribution in females was highly heterogenous among populations (Figure 1a). These SRD were found in 10 and 11 out of 16 populations, but they reached frequencies >10% in only 6 and 7 populations, respectively. The two SRD coexisted in 8 populations. A generalized linear model predicting the frequency of the *f* element as a binomial response by the proportion of individuals carrying *Wolbachia* (each statistical unit being a population) showed that the prevalence of the two SRD was significantly negatively correlated (9.8% of deviance explained, Chi-squared test,  $p < 7.9 \times 10^{-8}$ , 14 df) (Figure 1b). Hence, in Floirac, Poitiers, Saint Julien l'Ars and Pisa populations, *Wolbachia* was frequent (23-94% frequency in females) and the *f* element was rare (0-8%). By contrast, the *f* element was frequent (35-96%) and *Wolbachia* was rare (0-11%) in Prague, Beauvoir, Chizé, Coulombiers and La Crèche populations. In the other populations, both SRD were found at low to moderate frequency (0-19%), including 3 populations devoid of both SRD (Lastovo, Hyogo and Bucharest).

Males carrying *Wolbachia* or the *f* element were found in 2 and 4 out of 16 populations, respectively. In all cases, these males occurred in populations in which the corresponding SRD were the most prevalent ones in females: Beauvoir, Chizé, Coulombiers and La Crèche for the *f* element, and Floirac and Saint Julien l'Ars for *Wolbachia*. Overall, these males had much lower *Wolbachia* titer than females from their respective populations (Figure S1, Table S3).

The 642 individuals sequenced at the *COI* gene presented a total of 92 segregating sites defining 23 haplotypes (named I to XXIII; GenBank accession numbers in Table S4), with 1 to 7 haplotypes per population (Table S2, Figure 2). The most frequent and widespread haplotype (I) was found in 188 individuals from 10 populations. The second most frequent and widespread haplotype (V) was found

in 106 individuals from 7 populations. We found 21 out of the 23 haplotypes among individuals lacking both *Wolbachia* and the *f* element (Table 2, Figure 2). Among individuals carrying the *f* element, 6 haplotypes were found, all but one (I, II, III, V and VI) being shared with individuals lacking both *Wolbachia* and the *f* element, and one (IV) being carried by a single individual in the entire dataset. Among *Wolbachia*-infected individuals, all those carrying wVulC were associated with either haplotype V or its close relatives (XI and XII). All individuals carrying wVulM were associated with haplotype II and those carrying wVulP with haplotype VII. Of the 5 haplotypes found in *Wolbachia*-infected individuals, 4 were shared with individuals lacking both *Wolbachia* and the *f* element (II, V, VII and XII), 2 of which were also shared with individuals carrying the *f* element (II and V), and one (XI) was present in a single individual in the entire dataset.

#### 4. Discussion

Our results provide direct evidence that the *f* element is overall more frequent than *Wolbachia* in the sampled *A. vulgare* populations. We detected the *f* element in 11 *A. vulgare* natural populations from 4 European countries (Czech Republic, France, Germany and The Netherlands) and Japan. Together with its previous detection in Denmark [21], our results indicate that the *f* element has spread to a wide geographical range. The relative frequencies of the *f* element and *Wolbachia* are highly variable among populations and, in general, when one SRD is frequent, the other SRD is rare. Overall, these results are consistent with earlier results from the 1990's [32], although no molecular assay allowing direct testing was available at that time and SRD presence or absence was inferred indirectly.

As the *Jtel* marker is located across the site of integration of the *f* element in the *A. vulgare* chromosome [21], *f* element presence in various populations can be explained by a single event of integration of a *Wolbachia* genome in the *A. vulgare* genome. A less parsimonious scenario would require independent insertions at the same chromosomal site, which is highly unlikely. The former scenario contradicts an earlier hypothesis on the evolutionary dynamics of the *f* element, which suggested that the *f* element was unstably integrated in the *A. vulgare* genome, experiencing frequent loss from oocytes and recurrent gain from *Wolbachia* endosymbionts [22,23,42–44]. Under this scenario, multiple independent *f*-like elements would be expected to segregate at low frequencies in populations, they should be integrated in different genomic locations and they should all be able to induce feminization [16]. While our results do not formally invalidate the possibility of additional *f*-like integrations in *A. vulgare* populations, which the *Jtel* marker would not detect, it

does not appear to be the most parsimonious hypothesis. Examination of sex ratios from progenies of wild-caught females lacking both SRD may offer further insight into this issue.

Using molecular assays allowed us to circumvent two limitations of the previously used physiological test: the impossibility to detect *Wolbachia* and the *f* element in males, and the impossibility to detect individuals carrying both SRD. Regarding *Wolbachia* presence in males, the historic protocol was only applicable to females per design [29,30,32] and subsequent PCR screens for *Wolbachia* infection have mostly focused on testing females [20,30,31]. In fact, males have seldom been tested and found to carry *Wolbachia* [45]. Here, we detected *Wolbachia* in 7 males from 2 populations (Florac and Saint Julien l'Ars), carrying either wVulC or wVulM strains. The failure of feminization by *Wolbachia* most certainly reflects insufficient bacterial titer to induce feminization (Figure S1). These field observations hence support the view that titer is an important factor for successful feminization, as low titer is linked to incomplete feminization and intersexual phenotypes [42,46].

We also detected the presence of the *f* element in 11 males from 4 populations. Historically, the presence of the *f* element in males has been indirectly inferred from crossings and the resulting sex-ratios biases of progenies [22,43,47]. Our results constitute the first direct evidence for the presence of the *f* element in *A. vulgare* males. In all 4 populations in which *f*-carrying males were found, the *f* element was also frequent in females. Altogether, these observations suggest that the 11 males carrying the *f* element also carry the masculinizing dominant *M* allele [16,43,47]. Indeed, the *M* allele is able to restore a male phenotype in individuals carrying the *f* element [16,43,47]. Because of sex ratio selection, the *M* allele is thought to have been selected to restore males in response to female-biased sex ratios caused by the *f* element [47]. Thus, the *M* allele is expected to rise in frequency when the *f* element is frequent in a population [47], which is consistent with our observations. Unfortunately, no molecular marker of the *M* allele is currently available, which prevents any direct assessment of its actual presence in these populations. Thus, we cannot exclude that males carrying the *f* element simply carry non-feminizing variants of this SRD.

Our results show that *Wolbachia* and the *f* element never co-occur at high frequency in any population. This apparent mutual exclusion can be explained considering that co-occurrence of multiple feminizing factors in a population should favor the most transmitted one [16,48]. Hence, *Wolbachia* is expected to lead to the loss of nuclear feminizing elements in *A. vulgare* populations. This situation does not result from an interference between chromosomes and *Wolbachia* within individuals, but from counter selection of nuclear feminizing alleles in a population that becomes increasingly biased towards females. Hence, the rise of *Wolbachia* would associate with the decline

of the *f* element in a population. Why, under these circumstances, *Wolbachia* has not invaded all *A. vulgare* populations is still unclear and may reflect fitness effects or possible resistance genes.

As a result, only very few individuals were found to carry both *Wolbachia* and the *f* element. They represent only 3 females, all from the Chizé population (Figure 1a). These were likely born from mothers carrying *Wolbachia* and fathers carrying the *f* element, which are frequent at Chizé. The apparent absence of carriers of both SRD in other populations where these SRD are present could simply be explained by the paucity of males carrying the *f* element.

Mapping SRD distribution onto mitochondrial genealogy showed excellent congruence between *Wolbachia* strains and mitochondrial haplotypes (*wVulC-V*, *wVulM-II* and *wVulP-VII*). Such strong association has previously been noted in *A. vulgare-Wolbachia* interactions at a smaller geographic scale [30,31] and, more generally, in many arthropod-*Wolbachia* interactions [49]. This result corroborates the rarity of non-maternal transmission of *Wolbachia* in *A. vulgare*. By contrast, the *f* element was found in 6 different mitochondrial backgrounds (I-VI) scattered across the mitochondrial phylogeny, indicating no particular association between the *f* element and mitochondria. This result confirms and extends earlier data focused on western France and in which *f* element presence in females was indirectly inferred based on sex ratios of their progenies [30]. This observation can be explained by the occasional paternal transmission of the *f* element, which breaks its association with mitochondrial background [16,22,30].



217 **Data accessibility**

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219 All data are provided in the electronic supplementary material.

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## Figure legends

**Figure 1. (A)** Prevalence of *Wolbachia* and the *f* element in males (m) and females (f) from 16 *Armadillidium vulgare* populations. **(B)** Relative proportions of *Wolbachia* and the *f* element in 16 *A. vulgare* populations (represented by open circles).

**Figure 2.** Haplotype network of 23 mitochondrial variants (I-XXIII) from 16 *Armadillidium vulgare* populations. Each circle represents one haplotype and circle diameter is proportional to the number of individuals carrying the haplotype. Branch lengths connecting circles are proportional to divergence between haplotypes. Sex ratio distorter frequencies are color-coded for each haplotype.

245 **Table 1. Prevalence of *Wolbachia* and *f* element sex ratio distorters in 16 populations of *Armadillidium vulgare*.**

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Population (Country)	Sampling year	Sample size	Sex	Number of individuals	No <i>f</i> element, no <i>Wolbachia</i>	Only <i>f</i> element	Only <i>Wolbachia</i>				Both wVulM and <i>f</i> element	Undetermined status
							wVulC	wVulM	wVulP	Undetermined		
Lastovo	2017	54	Males	30	30							
(Croatia)			Females	24	24							
Prague	2018	36	Males	9	9							
(Czech Republic)			Females	27	1	26						
Beauvoir	2017	31	Males	6	5	1						
(France)			Females	25	9	14		1				1
Chizé	2017	52	Males	8	2	6						
(France)			Females	44	3	36		2			3	
Coulombiers	2017	24	Males	4	2	2						
(France)			Females	20	6	13	1					
Floirac	2016	114	Males	38	34		2					2
(France)			Females	76	21	6	40	9				
Gript	2017	45	Males	15	15							
(France)			Females	30	26	2	2					
La Crèche	2017	58	Males	21	19	2						
(France)			Females	37	23	13	1					
Poitiers	2015	23	Males	4	4							
(France)			Females	19	10	1			4	4		
Saint Julien l'Ars	2016	31	Males	14	9		1	3		1		
(France)			Females	17	1		12	3				1
Göttingen	2017	24	Males	7	3							4
(Germany)			Females	17	11	3		2				1
Pisa	2017	28	Males	15	15							
(Italy)			Females	13	10		3					
Hyogo	2018	50	Males	21	18							3

(Japan)			Females	29	26					3
Tottori	2018	49	Males	21	21					
(Japan)			Females	28	26	2				
Bucharest	2017	17	Males	9	9					
(Romania)			Females	8	8					
Wageningen	2018	11	Males	2	2					
(The Netherlands)			Females	9	7	1				1
Total males				224	197	11	3	3	1	9
Total females				423	212	117	59	17	4	7
Total				647	409	128	62	20	4	16

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249 **Table 2. Distribution of mitochondrial haplotypes in 642 *Armadillidium vulgare* individuals from 16 populations.**

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Sex ratio distorter status	Number of individuals	Haplotype number	Haplotype list
No <i>f</i> element, no <i>Wolbachia</i>	404	21	I, II, III, V, VI, VII, VIII, IX, X, XII, XIII, XIV, XV, XVI, XVII, XVIII, XIX, XX, XXI, XXII, XXIII
<i>f</i> element only	128	6	I, II, III, IV, V, VI
<i>Wolbachia</i> (wVulC strain) only	62	3	V, XI, XII
<i>Wolbachia</i> (wVulM strain) only	20	1	II
<i>Wolbachia</i> (wVulP strain) only	4	1	VII
<i>Wolbachia</i> (undetermined strain) only	5	2	II, VII
Both wVulM and <i>f</i> element	3	1	II
Undetermined status	16	4	I, V, VI, XIX

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## References

1. Beukeboom LW, Perrin N. 2014 *The evolution of sex determination*. Oxford: Oxford University Press.
2. Jaenike J. 2001 Sex chromosome meiotic drive. *Annu. Rev. Ecol. Syst.* **32**, 25–49. (doi:10.1146/annurev.ecolsys.32.081501.113958)
3. Helleu Q, Gérard PR, Montchamp-Moreau C. 2014 Sex chromosome drive. *Cold Spring Harb. Perspect. Biol.* **7**, a017616. (doi:10.1101/cshperspect.a017616)
4. Camacho JPM, Schmid M, Cabrero J. 2011 B chromosomes and sex in animals. *Sex. Dev. Genet. Mol. Biol. Evol. Endocrinol. Embryol. Pathol. Sex Determin. Differ.* **5**, 155–166. (doi:10.1159/000324930)
5. Chase CD. 2007 Cytoplasmic male sterility: a window to the world of plant mitochondrial-nuclear interactions. *Trends Genet.* **23**, 81–90. (doi:10.1016/j.tig.2006.12.004)
6. Cordaux R, Bouchon D, Greve P. 2011 The impact of endosymbionts on the evolution of host sex-determination mechanisms. *Trends Genet* **27**, 332–41.
7. Hurst GDD, Frost CL. 2015 Reproductive parasitism: maternally inherited symbionts in a biparental world. *Cold Spring Harb. Perspect. Biol.* **7**, a017699. (doi:10.1101/cshperspect.a017699)
8. Burt A, Trivers R. 2006 *Genes in conflict*. Cambridge, Massachusetts: The Belknap Press of Harvard University Press.
9. Werren JH. 2011 Selfish genetic elements, genetic conflict, and evolutionary innovation. *Proc. Natl. Acad. Sci. U. S. A.* **108 Suppl 2**, 10863–10870. (doi:10.1073/pnas.1102343108)
10. Werren JH, Baldo L, Clark ME. 2008 Wolbachia: master manipulators of invertebrate biology. *Nat Rev Microbiol* **6**, 741–51. (doi:10.1038/nrmicro1969)
11. Kaur R, Shropshire JD, Cross KL, Leigh B, Mansueto AJ, Stewart V, Bordenstein SR, Bordenstein SR. 2021 Living in the endosymbiotic world of Wolbachia: A centennial review. *Cell Host Microbe* **29**, 879–893. (doi:10.1016/j.chom.2021.03.006)
12. Juchault P, Legrand JJ. 1972 Croisement de néo-mâles expérimentaux chez *Armadillidium vulgare* Latr. (Crustace, Isopode, Oniscoïde). Mise en évidence d'une hétérogamétie femelle. *C R Acad Sci Paris* **274**, 1387–1389.
13. Chebbi MA, Becking T, Moumen B, Giraud I, Gilbert C, Peccoud J, Cordaux R. 2019 The Genome of *Armadillidium vulgare* (Crustacea, Isopoda) Provides Insights into Sex Chromosome Evolution in the Context of Cytoplasmic Sex Determination. *Mol. Biol. Evol.* **36**, 727–741. (doi:10.1093/molbev/msz010)
14. Cordaux R, Chebbi MA, Giraud I, Pleydell DRJ, Peccoud J. 2021 Characterization of a Sex-Determining Region and Its Genomic Context via Statistical Estimates of Haplotype Frequencies in Daughters and Sons Sequenced in Pools. *Genome Biol. Evol.* **13**, evab121. (doi:10.1093/gbe/evab121)

15. Rigaud T, Juchault P, Mocquard JP. 1997 The evolution of sex determination in isopods crustaceans. *Bioessays* **19**, 409–416.
16. Cordaux R, Gilbert C. 2017 Evolutionary Significance of Wolbachia-to-Animal Horizontal Gene Transfer: Female Sex Determination and the f Element in the Isopod *Armadillidium vulgare*. *Genes* **8**, 186. (doi:10.3390/genes8070186)
17. Martin G, Juchault P, Legrand JJ. 1973 Mise en évidence d'un micro-organisme intracytoplasmique symbiote de l'Oniscoïde *Armadillidium vulgare* L. dont la présence accompagne l'intersexualité ou la féminisation totale des mâles génétiques de la lignée thélygène. *Comptes Rendus Académie Sci. Paris* **276**, 2313–2316.
18. Rigaud T, Souty Grosset C, Raimond R, Mocquard JP, Juchault P. 1991 Feminizing endocytobiosis in the terrestrial crustacean *Armadillidium vulgare* Latr. (Isopoda): Recent acquisitions. *Endocytobiosis Cell Res* **7**, 259–273.
19. Cordaux R, Michel-Salzat A, Frelon-Raimond M, Rigaud T, Bouchon D. 2004 Evidence for a new feminizing Wolbachia strain in the isopod *Armadillidium vulgare*: evolutionary implications. *Heredity* **93**, 78–84. (doi:10.1038/sj.hdy.6800482)
20. Verne S, Johnson M, Bouchon D, Grandjean F. 2007 Evidence for recombination between feminizing Wolbachia in the isopod genus *Armadillidium*. *Gene* **397**, 58–66. (doi:10.1016/j.gene.2007.04.006)
21. Leclercq Sb, Thézé J, Chebbi MA, Giraud I, Moumen B, Ernenwein L, Greve P, Gilbert C, Cordaux R. 2016 Birth of a W sex chromosome by horizontal transfer of *Wolbachia* bacterial symbiont genome. *Proc. Natl. Acad. Sci. U. S. A.* **113**, 15036–15041. (doi:10.1073/pnas.1608979113)
22. Legrand JJ, Juchault P. 1984 Nouvelles données sur le déterminisme génétique et épigénétique de la monogénie chez le crustacés isopodes terrestres *Armadillidium vulgare* Latr. *Génét Sél Evol* **16**, 57–84.
23. Juchault P, Mocquard JP. 1993 Transfer of a parasitic sex factor to the nuclear genome of the host: A hypothesis on the evolution of sex-determining mechanisms in the terrestrial isopod *Armadillidium vulgare* Latr. *J Evol Biol* **6**, 511–528.
24. Juchault P, Rigaud T. 1995 Evidence for female heterogamety in two terrestrial crustaceans and the problem of sex chromosome evolution in isopods. *Heredity* **75**, 466–471.
25. Becking T, Giraud I, Raimond M, Moumen B, Chandler C, Cordaux R, Gilbert C. 2017 Diversity and evolution of sex determination systems in terrestrial isopods. *Sci. Rep.* **7**, 1–14. (doi:10.1038/s41598-017-01195-4)
26. Becking T, Chebbi MA, Giraud I, Moumen B, Laverré T, Caubet Y, Peccoud J, Gilbert C, Cordaux R. 2019 Sex chromosomes control vertical transmission of feminizing Wolbachia symbionts in an isopod. *PLOS Biol.* **17**, e3000438. (doi:10.1371/journal.pbio.3000438)
27. Russell A, Borrelli S, Fontana R, Laricchiuta J, Pascar J, Becking T, Giraud I, Cordaux R, Chandler CH. 2021 Evolutionary transition to XY sex chromosomes associated with Y-linked duplication of a male hormone gene in a terrestrial isopod. *Heredity* **127**, 266–277. (doi:10.1038/s41437-021-00457-2)

- 330 28. Juchault P, Legrand JJ, Mocquard JP. 1980 Contribution à l'étude qualitative et quantitative  
331 des facteurs contrôlant le sexe dans les populations du crustacé isopode terrestre *Armadillidium*  
332 *vulgare* Latreille. I. La population de Niort (Deux Sèvres). *Arch Zool Exp Gen* **121**, 3–27.
- 333 29. Grandjean F, Rigaud T, Raimond R, Juchault P, Souty-Grosset C. 1993 Mitochondrial DNA  
334 polymorphism and feminizing sex factor dynamics in a natural population of *Armadillidium*  
335 *vulgare* (Crustacea, Isopoda). *Genetica* **92**, 55–60.
- 336 30. Rigaud T, Bouchon D, Souty-Grosset C, Raimond R. 1999 Mitochondrial DNA polymorphism,  
337 sex ratio distorters and population genetics in the isopod *Armadillidium vulgare*. *Genetics* **152**,  
338 1669–1677.
- 339 31. Verne S, Johnson M, Bouchon D, Grandjean F. 2012 Effects of parasitic sex-ratio distorters on  
340 host genetic structure in the *Armadillidium vulgare*-*Wolbachia* association. *J. Evol. Biol.* **25**, 264–  
341 76. (doi:10.1111/j.1420-9101.2011.02413.x)
- 342 32. Juchault P, Rigaud T, Mocquard JP. 1993 EVOLUTION OF SEX DETERMINATION AND SEX-  
343 RATIO VARIABILITY IN WILD POPULATIONS OF ARMADILLIDIUM-VULGARE (LATR) (CRUSTACEA,  
344 ISOPODA) - A CASE-STUDY IN CONFLICT-RESOLUTION. *Acta Oecologica-Int. J. Ecol.* **14**, 547–562.
- 345 33. Juchault P, Legrand JJ. 1981 Contribution à l'étude qualitative et quantitative des facteurs  
346 contrôlant le sexe dans les populations du Crustacé Isopode terrestre *Armadillidium vulgare* Latr.  
347 II - Populations hébergeant le facteur féminisant F (bactérie intracytoplasmique). *Arch Zool Exp*  
348 *Gén* **122**, 65–74.
- 349 34. Juchault P, Legrand JJ. 1981 Contribution à l'étude qualitative et quantitative des facteurs  
350 contrôlant le sexe dans les populations du Crustacé isopode terrestre *Armadillidium vulgare*  
351 Latreille. III. Populations n'hébergeant pas le facteur féminisant F (bactéroïde intracytoplasmique).  
352 *Arch Zool Exp Gén* **122**, 117–131.
- 353 35. Braig HR, Zhou WG, Dobson SL, O'Neill SL. 1998 Cloning and characterization of a gene  
354 encoding the major surface protein of the bacterial endosymbiont *Wolbachia pipientis*. *J.*  
355 *Bacteriol.* **180**, 2373–2378.
- 356 36. Badawi M, Giraud I, Vavre F, Grève P, Cordaux R. 2014 Signs of Neutralization in a Redundant  
357 Gene Involved in Homologous Recombination in *Wolbachia* Endosymbionts. *Genome Biol. Evol.* **6**,  
358 2654–2664. (doi:10.1093/gbe/evu207)
- 359 37. Werren JH, Zhang W, Guo LR. 1995 Evolution and phylogeny of *Wolbachia*: reproductive  
360 parasites of arthropods. *Proc. Biol. Sci.* **261**, 55–63. (doi:10.1098/rspb.1995.0117)
- 361 38. Folmer O, Black M, Hoeh W, Lutz R, Vrijenhoek R. 1994 DNA primers for amplification of  
362 mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Mol. Mar.*  
363 *Biol. Biotechnol.* **3**, 294–299.
- 364 39. Paradis E. 2010 pegas: an R package for population genetics with an integrated–modular  
365 approach. *Bioinformatics* **26**, 419–420.
- 366 40. R Development Core Team. 2013 *R: A language and environment for statistical computing*.  
367 See <http://www.R-project.org/>.
- 368 41. Wickham H *et al.* 2020 ggplot2: Create Elegant Data Visualisations Using the Grammar of  
369 Graphics.



42. Juchault P, Legrand JJ. 1989 Sex determination and monogeny in terrestrial isopods *Armadillidium vulgare* (Latreille, 1804) and *Armadillidium nasatum* bundde-lund, 1885. *Monit. Zool Ital NS Monogr* **4**, 359–375.
43. Juchault P, Rigaud T, Mocquard J-P. 1992 Evolution of sex-determining mechanisms in a wild population of *Armadillidium vulgare* Latr. (Crustacea, Isopod) : competition between two feminizing parasitic sex factors. *Heredity* **69**, 382–390.
44. Rigaud T, Mocquard J-P, Juchault P. 1992 The spread of parasitic sex factors in populations of *Armadillidium vulgare* Latr. (Crustacea, Oniscidae): effects on sex ratio. *Génét Sél Evol* **24**, 3–18.
45. Dittmer J, Lesobre J, Moumen B, Bouchon D. 2016 Host origin and tissue microhabitat shaping the microbiota of the terrestrial isopod *Armadillidium vulgare*. *FEMS Microbiol. Ecol.* **92**, fiw063. (doi:10.1093/femsec/fiw063)
46. Legrand JJ, Juchault P. 1986 Rôle des bactéries symbiotiques dans l’intersexualité, la monogénie et la spéciation chez les crustacés oniscoïdes. *Boll Zool* **53**, 161–172.
47. Rigaud T, Juchault P. 1993 Conflict between feminizing sex ratio distorters and an autosomal masculinizing gene in the terrestrial isopod *Armadillidium vulgare* Latr. *Genetics* **133**, 247–252.
48. Taylor DR. 1990 Evolutionary consequences of cytoplasmic sex ratio distorters. *Evol. Ecol.* **4**, 235–248.
49. Galtier N, Nabholz B, Glémin S, Hurst GDD. 2009 Mitochondrial DNA as a marker of molecular diversity: a reappraisal. *Mol. Ecol.* **18**, 4541–4550. (doi:10.1111/j.1365-294X.2009.04380.x)

Figure 1

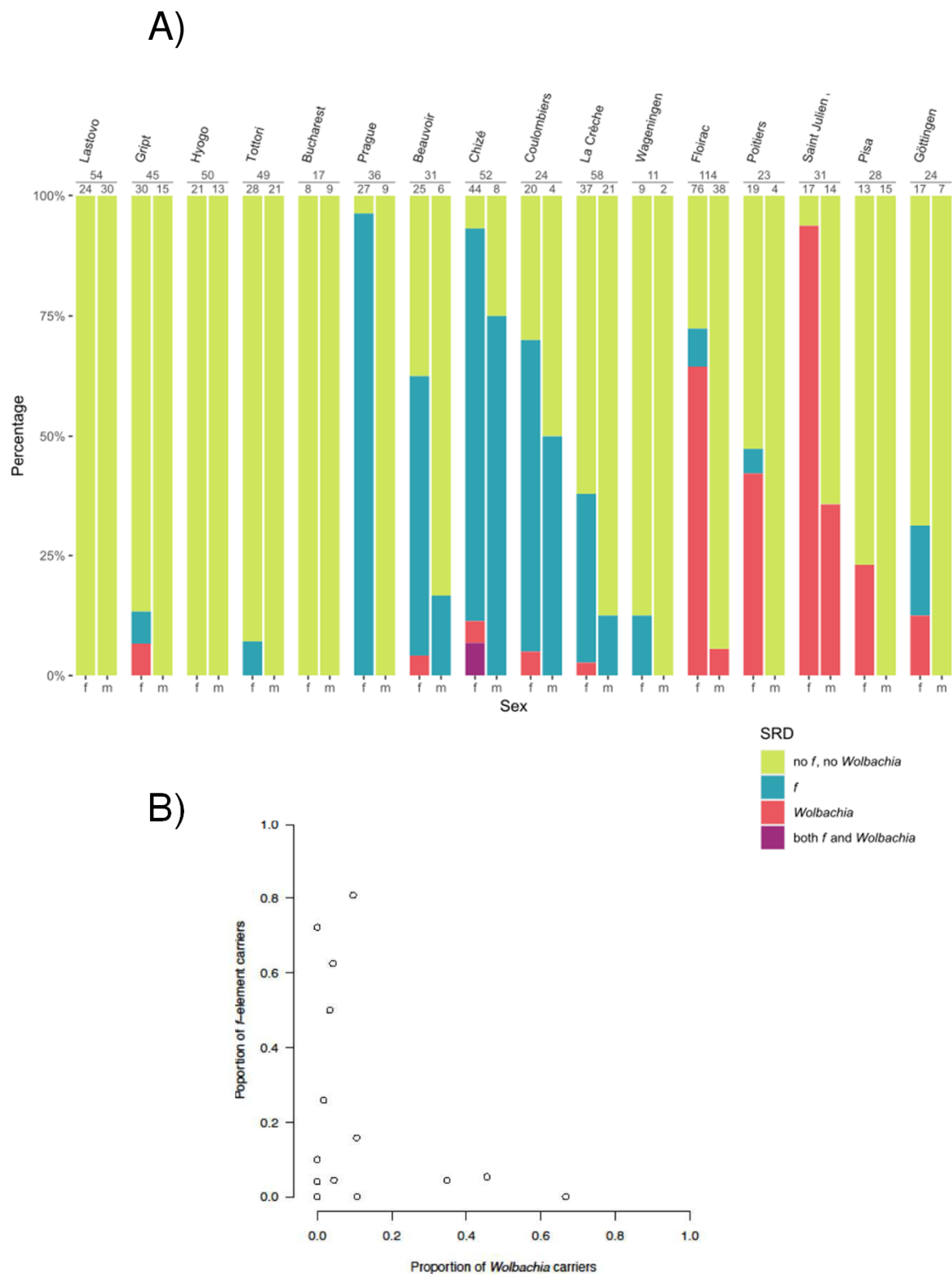


Figure 2

