

1 **Whole genome sequencing suggests that “non-pathogenicity on banana (NPB)” is**  
2 **the ancestral state of the *Ralstonia solanacearum* IIB-4 lineage.**

3

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11

12 **Abstract**

13 The bacterial wilt pathogens in the *Ralstonia solanacearum* species complex (RSSC) have broad  
14 but finite host ranges. Population genetic surveys of RSSC pathogens show that many sequevars  
15 (subspecies groups) are predominantly recovered from wilting solanaceous plants. In contrast,  
16 strains in the IIB-4 sequevar have been isolated from plants in over a dozen families. Certain IIB-  
17 4 lineages have been classified as banana-virulent or “not pathogenic to banana (NPB)”. Prior  
18 analysis suggested that the NPB lineage has diverged from the banana-virulent IIB-4 strains. To  
19 test this model, we analyzed the phenotypes and phylogeny of a diverse collection of 19 IIB-4  
20 isolates. We used Illumina sequencing to assemble draft genomes of 12 new strains. Based on  
21 whole genome phylogenetic analysis, these IIB-4 strains clustered into five subclades. We  
22 quantified virulence of each strain on tomato, banana, melon, and impatiens plants. Overall,  
23 the virulence patterns correlated with phylogeny. Banana virulence was restricted to the 4/4  
24 IIB-4D subclade (N=4/4 strains) and IIB-4E subclade (N=1/2 strains). Subclades IIB-4D and IIB-4E  
25 are sister subclades and their closest relative, the IIB-4A-C subclade, lacked virulence on  
26 banana. Our data support a revised model in which banana virulence is an innovation within  
27 the IIB4D/E subclades.

28 **Data Summary**

29 Illumina sequencing and genome assembly data are available as NCBI BioProject PRJNA826884,  
30 and Table S1 lists the accession numbers for assemblies in GenBank and raw sequencing data in  
31 SRA. To enable future meta-analyses that identify genetic factors that drive host-range, the raw  
32 virulence data is included as Table S2.

33

## 34 **Introduction**

35 The *Ralstonia solanacearum* species complex is a group of related tropical and temperate  
36 pathogens that cause plant wilt diseases after they invade and colonize the xylem (Prior et al.  
37 2016; Lowe-Power et al. 2018; Ingel et al. 2022). Collectively, the species complex has a  
38 remarkably broad host range, capable of infecting hundreds of host species in dozens of  
39 taxonomic families (Hayward 1991; Lowe-Power et al. 2020). However, the breadth in host  
40 range is not uniformly distributed across the species complex. Strains in the *R. solanacearum*  
41 phylotype IIB sequevar 4 clade (IIB-4) exhibit a particularly broad host range (Ailloud et al. 2015;  
42 Wicker et al. 2007) (Lowe-Power et al. 2020). *R. solanacearum* IIB-4 strains have been isolated  
43 from 33 plant species in 19 botanical families (Wicker et al. 2007; Norman et al. 2009; Cellier et  
44 al. 2012; Hong et al. 2012; Cellier et al. 2015; Gutarra et al. 2017; Ramírez et al. 2020; Zulperi et  
45 al. 2014; Lowe-Power et al. 2020; Cellier and Prior 2010). They are distributed throughout  
46 Central and South America, the ancestral range of phylotype II *Ralstonia*. This lineage is also  
47 globally invasive as IIB-4 strains have become established in Malaysia (Zulperi et al. 2014) and  
48 have been imported into Florida (Norman et al. 2009).

49

50 *R. solanacearum* IIB-4 have caused several well-studied wilt epidemics. In the 1960s, *R.*  
51 *solanacearum* IIB-4 caused a high-impact Moko disease epidemic of plantains and bananas in  
52 Colombia and Peru (French and Sequeira 1970; Buddenhagen and Elsasser 1962). These Moko  
53 isolates were also virulent on solanaceous crops: tomato, pepper, and potato (French and  
54 Sequeira 1970). In the 2000s, a subgroup of *R. solanacearum* IIB-4 from Martinique were  
55 recognized as a novel “not pathogenic to banana (NPB) ecotype” with high virulence on  
56 cucurbits but low virulence to banana (Wicker et al. 2007). In 2020, a third ecotype of IIB-4 from  
57 Colombia was described (Ramírez et al. 2020); these novel Colombian strains are the only IIB-4  
58 strains that are known to be avirulent on tomato. Overall, the well-documented natural history  
59 of IIB-4 strains and the diversity of their hosts position *R. solanacearum* IIB-4 as a model clade

60 for investigating the factors that limit and enable the breadth of host range in bacterial wilt  
61 pathogens.

62

63 Our prior comparative genomic analysis of 14 phylotype II strains, including 5 IIb-4 strains, was  
64 consistent with a hypothesis that the NPB lineage diverged from the banana-virulent IIb-4  
65 strains. To test this hypothesis, we collected IIb-4 isolates from additional sources, inferred  
66 their phylogeny by whole-genome sequencing and assembly, and analyzed their virulence  
67 phenotypes on host species from four botanical families: banana, tomato, melon, and  
68 impatiens.

## 69 **Results**

70 We quantified the natural variation in virulence of 19 *Ralstonia solanacearum* IIb-4 strains  
71 against four natural host species from four botanical families. The strains were isolated from  
72 diverse plants and locations (see Table S1). Seven strains were isolated in Peru and Colombia  
73 from 1960-1963. Of these, six were from plantain during an epidemic of Moko disease, and one  
74 from *Heliconia* (Buddenhagen and Elsasser 1962; French and Sequeira 1970). Three strains were  
75 isolated on *Anthurium* and *Heliconia* during the Martinique epidemic (Wicker et al. 2007).  
76 Seven strains were isolated from ornamentals imported into Florida. Of these, six were isolated  
77 between 1996-2012 from *Pothos*, and one from *Anthurium* in 1997 (Norman et al. 2009). One  
78 strain was isolated in 1999 from cucumber in Brazil (Ailloud et al. 2015), and one in 2017 from  
79 *Pothos* in the Dominican Republic.

80

81 To determine the diversity of these strains, we used Illumina sequencing to assemble draft  
82 genomes of 12 strains. An additional four unpublished genomes were shared by Caitlyn Allen  
83 and Boris Vinatzer. The draft genome assembly of P488 is lower quality than others. The P488  
84 assembly is fragmented into 200 contigs with an  $N_{50}$  of ~56 kb while the other assemblies have  
85 40-115 contigs with an  $N_{50}$  of 111-414 kb. Full genome assembly statistics are in Table S1. We

86 collected genomes of five additional IIB-4 strains, the closely related IIB-51 strain CFBP7014,  
87 and two distantly related strains (phylotype I GMI1000 and phylotype IIC K60 (Sharma et al.  
88 2022)) from NCBI.

89

90 We constructed a phylogenetic tree using 4,317 genes comparing 369,907 intragenic SNPs (Fig  
91 1). Three of the IIB-4 genomes had longer branch lengths than expected (P488, CFBP6783, and  
92 UW179), consistent with the lower-quality, more fragmented assemblies for these strains. The  
93 tree separated the IIB-4 strains into five branches, which we refer to as subclades A to E. The  
94 *Heliconia* isolate from 1961 (UW170) was the sole genome on subclade A. Seven *Pothos* and  
95 *Anthurium* isolates from Florida, and one from the Dominican Republic all clustered in subclade  
96 B. The Brazilian cucumber isolate IBSBF1503 clustered into subclade C with the three strains  
97 from the Martinique epidemic. The four Peruvian strains from the 1960s Moko epidemic  
98 clustered into subclade D. Subclade E contained three Colombian strains isolated from plantain  
99 in the 1960s, three Colombian strains recently isolated from plantain, and one Mexican strain  
100 isolated from potato.

101

102 We quantified virulence of 19 IIB-4 strains on four diverse plant species that are documented  
103 natural hosts for *Ralstonia* IIB-4: tomato (*Solanum lycopersicum* cv. Moneymaker), melon  
104 (*Cucumis melo* cv. Sweet Granite), impatiens (*Impatiens walleriana* cv. Beacon Orange), and  
105 banana (AAA *Musa acuminata* cv. Dwarf Cavendish). Tomato, melon, and impatiens were  
106 grown from seed while banana plants were purchased as tissue cultured plantlets. Plants were  
107 grown in growth chambers at 28° C (tomato, melon, and impatiens) or 32° C (banana) with a 12  
108 hr light cycle and 40% humidity. Tomato and melon seeds were sown into Sunshine Mix #1 to a  
109 depth of 1 cm, and impatiens were sown onto the surface of Sunshine Mix and topped with 1  
110 mm vermiculite. Seedlings were transplanted into 3.5" pots with approximately 60 g Sunshine

111 Mix at 7 days (melon), 14 days (tomato), 21 days post sowing (impatiens), or at the 3-leaf stage  
112 (banana).

113

114 We inoculated plants at 21 days post sowing (tomato and melon), 35 days post sowing  
115 (impatiens), or at the 4-leaf stage (banana). We inoculated tomato and impatiens via a standard  
116 cut-petiole approach where the lowest petiole was excised, and a 2  $\mu$ l droplet of bacterial  
117 suspension (containing approximately 1000 CFU) was placed on the cut surface (Khokhani et al.  
118 2018). Due to rapid wound responses of cucurbits (Zimmermann et al. 2013), we inoculated  
119 melons by applying a 2  $\mu$ l droplet of 1000 CFU onto a razor blade and excising a petiole with the  
120 infested blade. Banana plants were inoculated by drenching the soil with a bacterial suspension  
121 (50 mL with  $5 \times 10^7$  to  $1 \times 10^8$  CFU/g soil) after the roots were lightly wounded with a 2 cm-wide  
122 metal spatula (Ailloud et al. 2016). We rated symptoms daily for 14 days (Fig. 2). We rated  
123 tomato symptoms on a standard 0-4 disease index (DI) where 0 = no wilt, 1 = 0.1-25% of leaflets  
124 wilted, 2 = 25.1-50% of leaflets wilted, 3 = 50.1-75% of leaflets wilted, and 4 = 75.1-100% of  
125 leaflets wilted (Khokhani et al. 2018). We measured symptoms on melon, impatiens, and  
126 banana as the percentage of leaves with wilt. Symptom measurements, area under the disease  
127 progress curve (AUDPC), and incidence are displayed in Table S2.

128

129 Although all strains were pathogenic to tomato, three strains had reduced virulence: UW170  
130 (subclade A), UW162 (subclade D), and UW175 (subclade E). Because these strains have been  
131 stored as “waterstocks” (in distilled water at room temperature) for decades, we suspect that  
132 these strains acquired mutations in culture storage that reduced their virulence. The subclade A  
133 strain UW170 was moderately virulent on both tomato and impatiens but caused minor wilt on  
134 only a few banana or melon plants. The subclade D strain UW162 had moderate virulence on  
135 tomato and mild virulence on impatiens, melon, and banana. The subclade E strain UW175 had

136 moderate virulence on tomato, mild virulence on impatiens and melon, and no virulence on  
137 banana.

138

139 Virulence of strains generally correlated with their phylogenetic relationships. The subclade B  
140 and C strains lacked virulence on banana but highly virulent on tomato, impatiens, and melon.  
141 The subclade C strains displayed slightly higher virulence on melon than the subclade B strains.  
142 Three other subclade D strains were highly virulent on tomato and had low-to-moderate  
143 virulence on impatiens, melon, and banana. The subclade E strains included UW70 which was  
144 highly virulent on tomato, melon, and banana. UW70 was more virulent on impatiens than  
145 subclade D strains, but less virulent than the subclade B and subclade C strains.

146

## 147 **Discussion**

148 Pathogen populations are collections of genotypically and phenotypically diverse individuals. By  
149 inferring phylogenetic relationships of strains, we can set the groundwork for genome-wide  
150 analyses that associate genotypic differences with phenotypes. For plant pathogenic *Ralstonia*,  
151 the sequevar system has been widely used to classify strains to sub-species groups (Fegan and  
152 Prior 2005; Lowe-Power et al. 2020). Although the sequevar system is merely based on  
153 sequence alignments of a single 750 bp DNA marker, we recently validated that sequevars for  
154 *Ralstonia* phylotype II (i.e. *Ralstonia solanacearum*) accurately reflect the strains' core genome  
155 phylogenetic trees (Sharma et al. 2022). However, the sequevar system broke down for the  
156 phylotype I branch of *Ralstonia pseudosolanacearum* (Sharma et al. 2022), which is known to  
157 be highly recombinogenic (Wicker et al. 2012; Peeters et al. 2013). Moving forward, bacterial  
158 taxonomy will continue to be influenced by whole-genome comparisons (Parks et al. 2020).  
159 Taxonomic systems like the Life Identification Number (LIN) system (Tian et al. 2020; Vinatzer et  
160 al. 2017) are likely to supersede the sequevar system. LINs are a systematic taxonomy based on  
161 multiple levels of whole genome average nucleotide identity (ANI). Strains in the IIB-4 sequevar

162 have pairwise ANI of over 99.5% on their shared genes, and they are circumscribed in the LIN  
163 group 14<sub>A</sub>1<sub>B</sub>0<sub>C</sub>0<sub>D</sub>0<sub>E</sub>3<sub>F</sub>0<sub>G</sub>0<sub>H</sub>1<sub>I</sub>0<sub>J</sub>0<sub>K</sub>0<sub>L</sub>0<sub>M</sub> (Sharma et al. 2022).

164

165 Although many *Ralstonia* isolates only wilt plants efficiently at tropical temperatures, some  
166 *Ralstonia* isolates can wilt plants at temperate temperatures. Cool-virulence is famously a trait  
167 of the global pandemic lineage of *Ralstonia* (IIB-1 AKA the U.S. Select Agent-regulated “Race 3  
168 Biovar 2” group)(Chamoiseau et al. 2009), but recent studies have shown that multiple  
169 lineages of *Ralstonia* can wilt tomato plants at cool temperatures: IIB-27, III-48, and the focal  
170 clade of this study, IIB-4 (Bocsanczy et al. 2014, 2017). Although we carried out this study’s  
171 virulence assays at warm, tropical temperatures, several of our isolates, P673 (subclade B),  
172 CFBP6783 (subclade C), and UW163 (subclade D) can wilt tomatoes at cool temperatures (18°C)  
173 (Bocsanczy et al. 2014, 2017).

174

175 Early studies suggested that IIB-4 strains have binary host range phenotype with either a  
176 “Moko” phenotype with virulence on banana/ tomato/ plantain or a “NPB” phenotype with  
177 virulence on tomato/ plantain/ anthurium/ cucurbits (Ailloud et al. 2015; Wicker et al. 2007;  
178 Ailloud et al. 2016). However, our study shows that the virulence patterns are more complex.  
179 First, we show that banana virulence is rare among the strains tested. The NPB phenotype was  
180 shared by the subclade C strains previously described as “not pathogenic to banana” (Ailloud et  
181 al. 2015) and strains from subclade A and B. Our results suggest that virulence on banana  
182 might be an acquired trait within the subclade D/E lineage. Consistently, virulence on banana is  
183 a polyphyletic trait shared by distantly related lineages in phylotypes IIA, IIB, and IV (Ray et al.  
184 2021; Albuquerque et al. 2014; Obrador-Sánchez et al. 2017). Our study did not include any  
185 genomes from the recently identified IIB-4 strains from Colombia that lacks virulence on  
186 tomato (Ramírez et al. 2020). If those strains branch independently from the five subclades of

187 the 24 IIB-4 strains in this study, we propose to name the newly described strains as “subclade  
188 F”.

189

190 One subclade E strain (UW70 also known as “CIP417”) was highly virulent on all host species  
191 tested. UW70 was isolated from plantain in Colombia in the 1960s and is closely related to  
192 strains isolated from plantain in Colombia in 2012 (UA-1591 and UA-1617)(Ramírez et al. 2020).  
193 Interestingly, UW70 was highly virulent on melons, but UA-1591 is reported to be avirulent on  
194 cucumber (Ramírez et al. 2020). Cucurbit virulence has also been documented in phylotype I  
195 strains isolated in Taiwan, Thailand, China, and Japan (He et al. 2021; Yahiaoui et al. 2017;  
196 Horita et al. 2014; Lin et al. 2014).

197

198 Traits of several of this study’s isolates were studied: subclade C strain CFBP6783, subclade D  
199 strain UW163 and subclade E strain UW70. Lebeau et al. (2011) measured virulence of  
200 CFBP6783 and 11 other diverse *Ralstonia* strains against 30 accessions of tomato, eggplant, or  
201 pepper accessions in the “Core-TEP” panel. In this study, CFBP6783 exhibited a unique pattern  
202 of high virulence on 10/10 tomato, 10/10 pepper, and 6/10 eggplant accessions. CFBP6783 is  
203 virulent on plantain (AAB *Musa* sp. cv. Dominico-Harton)(Wicker et al. 2007) and potato (cv.  
204 Bintje Munterschen x Franschen)(Cellier and Prior 2010), but is not virulent on Cavendish  
205 banana (Wicker et al. 2007 and this study). UW70 and UW163 were highly virulent on *Musa*  
206 *balbisiana* Colla, tomato (*Solanum lycopersicum* L. ‘Bonny Best’), pepper (*Capsicum annuum* L.  
207 ‘Sweet Yellow’), and potato (*Solanum tuberosum* L. ‘Russet Burbank’)(French and Sequeira  
208 1970). In contrast, they exhibited low virulence on eggplant (*Solanum melongena* L. ‘Black  
209 Beauty’) and tobacco (*Nicotiana tabacum* L. ‘Bottom Special’). In a separate study, UW163 (D  
210 subclade) and UW179 (E subclade) lacked virulence on Anthurium while subclade C strains  
211 CFBP6783 and IBSBF1503 were virulent on Anthurium (Ailloud et al. 2015).

212

213 Our findings reinforce the long-recognized phenotypic plasticity within the *Ralstonia*  
214 *solanacearum* species complex (Buddenhagen 1985). In addition to host range variation within  
215 sequevar IIB-4, banana-virulent strains in IIA-24 and IIA-41 clades vary in their virulence on  
216 tomato (Albuquerque et al. 2014). A study of phylotype I *Ralstonia* from Guanxi, China showed  
217 that closely related strains shared virulence on tomato, eggplant, tobacco, and potato but  
218 varied in their virulence on pepper, mulberry, ginger, and cucurbits (He et al. 2021).

219  
220 In attempts to classify strains by their host range, scientists have attempted to sort *Ralstonia*  
221 into “Races”, “Pathoprototypes”, “Pathotypes”, and “Ecotypes” (Lebeau et al. 2011; Cellier et al.  
222 2012; Buddenhagen 1985; Peeters et al. 2013). Having clear phenotypic classifications would be  
223 both intrinsically satisfying and immensely helpful for clarifying regulations. In the future, this  
224 may be possible through sophisticated statistical analysis. However, these data science  
225 approaches will require a concerted and sustained effort to generate robust data on the  
226 genomic and phenotypic diversity of the global *Ralstonia* population. In parallel, a systematic  
227 database should be developed to consolidate and communicate knowledge on host range of  
228 different *Ralstonia* lineages. Continued study is needed to gain deeper insight into the genetic  
229 and evolutionary drivers of host range in the *Ralstonia solanacearum* species complex

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237

238 **Conflicts of Interest**

239

240 The authors declare that there are no conflicts of interest.

241

242 **References**

243 Ailloud, F., Lowe, T., Cellier, G., Roche, D., Allen, C., and Prior, P. 2015. Comparative genomic  
244 analysis of *Ralstonia solanacearum* reveals candidate genes for host specificity. *BMC Genomics*.  
245 16:270.

246 Ailloud, F., Lowe, T. M., Robène, I., Cruveiller, S., Allen, C., and Prior, P. 2016. *In planta*  
247 comparative transcriptomics of host-adapted strains of *Ralstonia solanacearum*. *PeerJ*.  
248 4:e1549.

249 Albuquerque, G. M. R., Santos, L. A., Felix, K. C. S., Rollemburg, C. L., Silva, A. M. F., Souza, E. B.,  
250 et al. 2014. Moko disease-causing strains of *Ralstonia solanacearum* from Brazil extend known  
251 diversity in paraphyletic Phylotype II. *Phytopathology*. 104:1175–1182.

252 Bayliss, S. C., Thorpe, H. A., Coyle, N. M., Sheppard, S. K., and Feil, E. J. 2019. PIRATE: A fast and  
253 scalable pangenomics toolbox for clustering diverged orthologues in bacteria. *Gigascience*. 8  
254 Available at: <https://academic.oup.com/gigascience/article/8/10/giz119/5584409> [Accessed  
255 May 5, 2021].

256 Bocsanczy, A. M., Achenbach, U. C. M., Mangravita-Novo, A., Chow, M., and Norman, D. J. 2014.  
257 Proteomic comparison of *Ralstonia solanacearum* strains reveals temperature dependent  
258 virulence factors. *BMC Genomics*. 15:280.

259 Bocsanczy, A. M., Huguet-Tapia, J. C., and Norman, D. J. 2017. Comparative Genomics of  
260 *Ralstonia solanacearum* Identifies Candidate Genes Associated with Cool Virulence. *Front. Plant  
261 Sci.* 8:1565.

262 Buddenhagen. 1985. Bacterial wilt revisited. Bacterial wilt disease in Asia and the. Available at:  
263 <https://ageconsearch.umn.edu/record/134643/files/PR013.pdf#page=124>.

264 Buddenhagen, I. W., and Elsasser, T. A. 1962. An insect-spread bacterial wilt epiphytotic of  
265 bluggoe banana. *Nature*. 194:164–165.

266 Cellier, G., Moreau, A., Chabirand, A., Hostachy, B., Ailloud, F., and Prior, P. 2015. A duplex PCR  
267 assay for the detection of *Ralstonia solanacearum* phylotype II strains in *Musa* spp. *PLoS One*.  
268 10:e0122182.

269 Cellier, G., and Prior, P. 2010. Deciphering Phenotypic Diversity of *Ralstonia solanacearum*  
270 Strains Pathogenic to Potato. *Phytopathology*®. 100:1250–1261.

271 Cellier, G., Remenant, B., Chiroleu, F., Lefevre, P., and Prior, P. 2012. Phylogeny and  
272 population structure of brown rot- and Moko disease-causing strains of *Ralstonia solanacearum*  
273 phylotype II. *Appl. Environ. Microbiol.* 78:2367–2375.

274 Champoiseau, P. G., Jones, J. B., and Allen, C. 2009. *Ralstonia solanacearum* Race 3 Biovar 2  
275 causes tropical losses and temperate anxieties. *Plant Health Prog*. Available at:  
276 <https://apsjournals.apsnet.org/doi/abs/10.1094/PHP-2009-0313-01-RV>.

277 Fegan, M., and Prior, P. 2005. How complex is the *Ralstonia solanacearum* species complex. In  
278 *Bacterial wilt disease and the Ralstonia solanacearum species complex*, APS press St. Paul, p.  
279 449–461.

280 French, E. R., and Sequeira, L. 1970. Strains of *Pseudomonas solanacearum* from Central and  
281 South America: a comparative study. *Phytopathology*. 60:506–512.

282 Gutarra, L., Herrera, J., Fernandez, E., Kreuze, J., and Lindqvist-Kreuze, H. 2017. Diversity,  
283 pathogenicity, and current occurrence of bacterial wilt bacterium *Ralstonia solanacearum* in  
284 Peru. *Front. Plant Sci.* 8:1221.

285 Hayward, A. C. 1991. Biology and epidemiology of bacterial wilt caused by *Pseudomonas*  
286 *solanacearum*. *Annu. Rev. Phytopathol.* 29:65–87.

287 He, Y., Chen, Y., Zhang, Y., Qin, X., Wei, X., Zheng, D., et al. 2021. Genetic diversity of *Ralstonia*  
288 *solanacearum* species complex strains obtained from Guangxi, China and their pathogenicity on  
289 plants in the Cucurbitaceae family and other botanical families. *Plant Pathol.* Available at:  
290 <https://onlinelibrary.wiley.com/doi/10.1111/ppa.13389>.

291 Hong, J. C., Norman, D. J., Reed, D. L., Momol, M. T., and Jones, J. B. 2012. Diversity among  
292 *Ralstonia solanacearum* strains isolated from the Southeastern United States. *Phytopathology*.  
293 102:924–936.

294 Horita, M., Tsuchiya, K., Suga, Y., Yano, K., Waki, T., Kurose, D., et al. 2014. Current classification  
295 of *Ralstonia solanacearum* and genetic diversity of the strains in Japan. *J. Gen. Plant Pathol.*  
296 80:455–465.

297 Ingel, B., Caldwell, D., Duong, F., Parkinson, D. Y., McCulloh, K. A., Iyer-Pascuzzi, A. S., et al.  
298 2022. Revisiting the source of wilt symptoms: X-ray microcomputed tomography provides  
299 direct evidence that *Ralstonia* biomass clogs xylem vessels. *Phytofrontiers*. Available at:  
300 <http://dx.doi.org/10.1094/PHYTOFR-06-21-0041-R> [Accessed April 9, 2021].

301 Khokhani, D., Tran, T. M., Lowe-Power, T. M., and Allen, C. 2018. Plant assays for quantifying  
302 *Ralstonia solanacearum* virulence. *Bio-protocol*. Available at: <https://en.bio-protocol.org/CN/e3028>.

303 Lebeau, A., Daunay, M.-C., Frary, A., Palloix, A., Wang, J.-F., Dintinger, J., et al. 2011. Bacterial  
304 wilt resistance in tomato, pepper, and eggplant: genetic resources respond to diverse strains in  
305 the *Ralstonia solanacearum* species complex. *Phytopathology*. 101:154–165.

306 Letunic, I., and Bork, P. 2021. Interactive Tree Of Life (iTOL) v5: an online tool for phylogenetic  
307 tree display and annotation. *Nucleic Acids Res.* 49:W293–W296.

308 Lin, C.-H., Tsai, K.-C., Prior, P., and Wang, J.-F. 2014. Phylogenetic relationships and population  
309 structure of *Ralstonia solanacearum* isolated from diverse origins in Taiwan. *Plant Pathol.*  
310 63:1395–1403.

311 Lowe-Power, T., Avalos, J., Charco Munoz, M., and Chipman, K. 2020. A meta-analysis of the  
312 known global distribution and host range of the *Ralstonia* species complex. *bioRxiv*. Available  
313 at: <https://www.biorxiv.org/content/10.1101/2020.07.13.189936v1.abstract>.

314 Lowe-Power, T. M., Khokhani, D., and Allen, C. 2018. How *Ralstonia solanacearum* exploits and  
315 thrives in the flowing plant xylem environment. *Trends Microbiol.* 26:929–942.

316 Norman, D. J., Zapata, M., Gabriel, D. W., Duan, Y. P., Yuen, J. M. F., Mangravita-Novo, A., et al.  
317 2009. Genetic diversity and host range variation of *Ralstonia solanacearum* strains entering  
318 North America. *Phytopathology*. 99:1070–1077.

319 Obrador-Sánchez, J. A., Tzec-Simá, M., Higuera-Ciapara, I., and Canto-Canché, B. 2017. Genetic  
320 diversity of *Ralstonia solanacearum* strains from Mexico associated with Moko disease. *Eur. J.  
321 Plant Pathol.* 149:817–830.

323 Parks, D. H., Chuvochina, M., Chaumeil, P.-A., Rinke, C., Mussig, A. J., and Hugenholz, P. 2020. A  
324 complete domain-to-species taxonomy for Bacteria and Archaea. *Nat. Biotechnol.* 38:1079–  
325 1086.

326 Peeters, N., Guidot, A., Vailleau, F., and Valls, M. 2013. *Ralstonia solanacearum*, a widespread  
327 bacterial plant pathogen in the post-genomic era. *Mol. Plant Pathol.* 14:651–662.

328 Prior, P., Ailloud, F., Dalsing, B. L., Remenant, B., Sanchez, B., and Allen, C. 2016. Genomic and  
329 proteomic evidence supporting the division of the plant pathogen *Ralstonia solanacearum* into  
330 three species. *BMC Genomics.* 17:90.

331 Ramírez, M., Moncada, R. N., Villegas-Escobar, V., Jackson, R. W., and Ramírez, C. A. 2020.  
332 Phylogenetic and pathogenic variability of strains of *Ralstonia solanacearum* causing moko  
333 disease in Colombia. *Plant Pathol.* 69:360–369.

334 Ray, J. D., Subandiyah, S., Rincon-Florez, V. A., Prakoso, A. B., Mudita, I. W., Carvalhais, L. C., et  
335 al. 2021. Geographic Expansion of Banana Blood Disease in Southeast Asia. *Plant Dis.* 105:2792–  
336 2800.

337 Sharma, P., Johnson, M. A., Mazloom, R., Allen, C., Heath, L. S., Lowe-Power, T. M., et al. 2022.  
338 Meta-analysis of the *Ralstonia solanacearum* species complex (RSSC) based on comparative  
339 evolutionary genomics and reverse ecology. *Microb Genom.* 8 Available at:  
340 <http://dx.doi.org/10.1099/mgen.0.000791>.

341 Stamatakis, A. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large  
342 phylogenies. *Bioinformatics.* 30:1312–1313.

343 Tian, L., Huang, C., Mazloom, R., Heath, L. S., and Vinatzer, B. A. 2020. LINbase: a web server for  
344 genome-based identification of prokaryotes as members of crowdsourced taxa. *Nucleic Acids  
345 Res.* 48:W529–W537.

346 Vinatzer, B. A., Weisberg, A. J., Monteil, C. L., Elmarakeby, H. A., Sheppard, S. K., and Heath, L. S.  
347 2017. A Proposal for a Genome Similarity-Based Taxonomy for Plant-Pathogenic Bacteria that Is  
348 Sufficiently Precise to Reflect Phylogeny, Host Range, and Outbreak Affiliation Applied to  
349 *Pseudomonas syringae* sensu lato as a Proof of Concept. *Phytopathology.* 107:18–28.

350 Wicker, E., Grassart, L., Coranson-Beaudu, R., Mian, D., Guilbaud, C., Fegan, M., et al. 2007.  
351 *Ralstonia solanacearum* strains from Martinique (French West Indies) exhibiting a new  
352 pathogenic potential. *Appl. Environ. Microbiol.* 73:6790–6801.

353 Wicker, E., Lefevre, P., de Cambiaire, J.-C., Lemaire, C., Poussier, S., and Prior, P. 2012.  
354 Contrasting recombination patterns and demographic histories of the plant pathogen *Ralstonia  
355 solanacearum* inferred from MLSA. *ISME J.* 6:961–974.

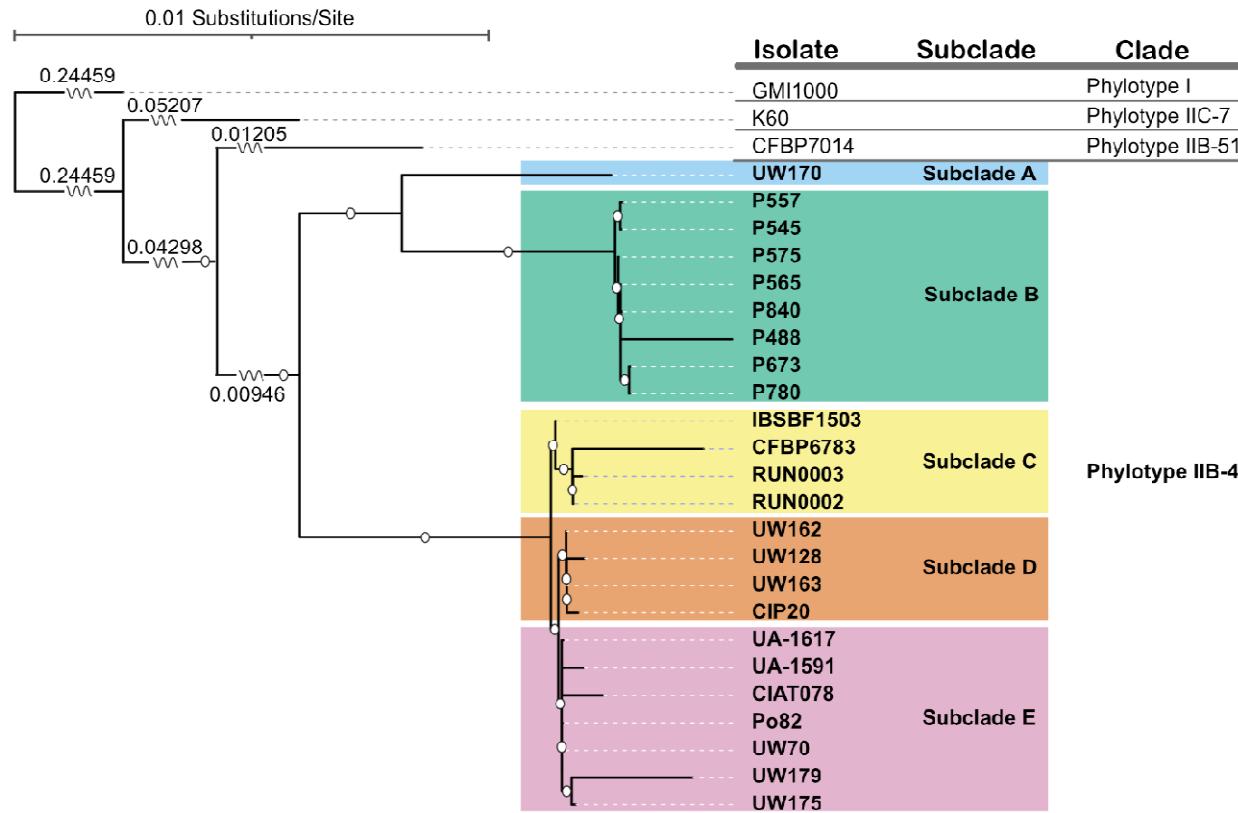
356 Yahiaoui, N., Chéron, J.-J., Ravelomanantsoa, S., Hamza, A. A., Petrousse, B., Jeetah, R., et al.  
357 2017. Genetic diversity of the *Ralstonia solanacearum* species complex in the Southwest Indian  
358 Ocean Islands. *Front. Plant Sci.* 8:2139.

359 Zimmermann, M. R., Hafke, J. B., van Bel, A. J. E., and Furch, A. C. U. 2013. Interaction of xylem  
360 and phloem during exudation and wound occlusion in *Cucurbita maxima*. *Plant Cell Environ.*  
361 36:237–247.

362 Zulperi, D., Sijam, K., Ahmad, Z. A. M., Awang, Y., and Mohd Hata, E. 2014. Phylotype  
363 classification of *Ralstonia solanacearum* biovar 1 strains isolated from banana (*Musa* spp.) in  
364 Malaysia. *Archives of Phytopathology and Plant Protection.* 47:2352–2364.

365

366 **Figures**



367

368 **Fig 1. Maximum likelihood tree describing 27 strains of the *Ralstonia solanacearum* species**

369 **complex, based on 369,907 intragenic SNPs.** The tree includes 24 IIB-4 strains and three

370 reference strains from phylotype IIB-51 (CFBP7014), IIC-7 (K60) (Sharma et al. 2022), and I

371 (GMI1000). Strains are described as belonging to one of five IIB-4 subclades (A-E). 95%

372 bootstrap support is indicated by a white circle. Some branch lengths were condensed for

373 clarity. SNPs were identified by grouping orthologous genes across all isolates using PIRATE

374 v1.0.4 with default parameters and nucleotide-based clustering (Bayliss et al. 2019).

375 Orthologous gene groups identified by PIRATE were concatenated and reduced to positions

376 present in at least 20 accessions, with at least one variable nucleotide, by a python script

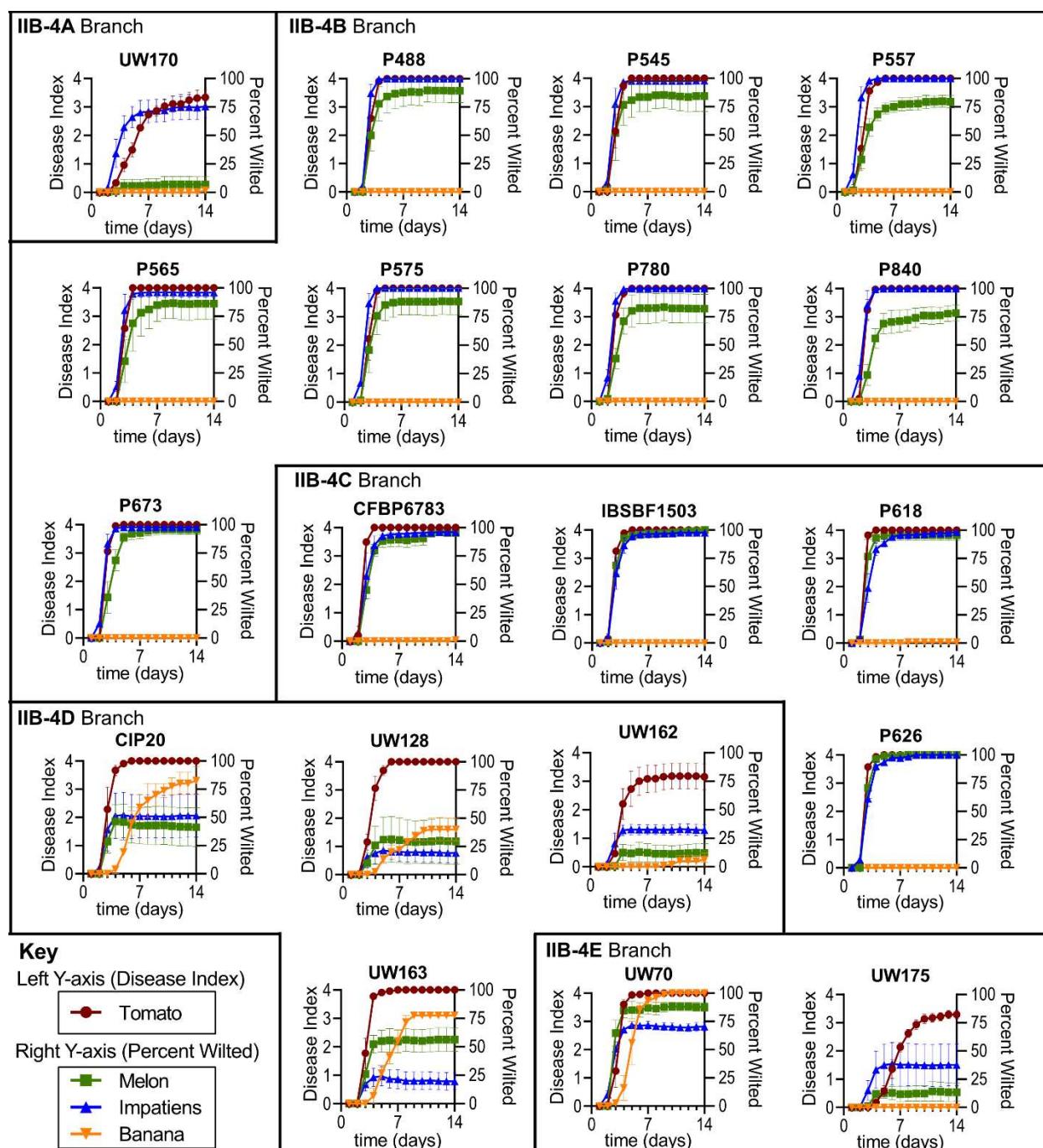
377 `Phylip_reducer.py` ([github.com/Erysiphe-graminis/Utilities](https://github.com/Erysiphe-graminis/Utilities)). These positions were used to

378 generate the tree using RAxML v8.2.12 with the GTRGAMMA model, Lewis ascertainment bias

379 correction, and 1000 bootstraps (Stamatakis 2014). GMI1000 was the outgroup. The tree was

380 visualized using iTOL v6.5.2 (Letunic and Bork 2021), and the figure prepared using Adobe  
381 Illustrator. To evaluate the impact of excluding infrequently occurring genes, two additional  
382 trees were generated using the same parameters except that SNP positions could be present in  
383 (1) at least 10 accessions, or (2) in any number of accessions. These trees were topologically  
384 identical with subtle variations in lengths of some branches.

385



386

387 **Fig 2.** Disease progress of 19 *R. solanacearum* IIB-4 strains on four host species: tomato cv.  
 388 Moneymaker ( $n \geq 44$  plants), melon cv. Sweet Granite ( $n \geq 43$ ), impatiens cv. Beacon Orange  
 389 ( $n \geq 44$ ), and Banana cv. Dwarf Cavendish ( $n \geq 19$ ). Each graph represents one strain. Virulence on  
 390 tomato was quantified on 0-4 disease index (left Y-axes), and virulence on other hosts was  
 391 quantified as a percentage of leaves with wilt (right Y-axes). Symbols represent the mean of 2-3

392 trials and bars show standard error of the mean (SEM). Strains are grouped based on the  
393 phylogeny from Fig 1. Raw virulence data is available in Table S2.

394

0.01 Substitutions/Site

