

1 **MicroRNA-224 Promotes Cell Migration and Invasion by**
2 **Targeting HOXA5 expression in Hepatocellular Carcinoma**

3 Yuwu Liu¹, Chen Lin¹, Dongmei Wang², Sailan Wen³, Junpu Wang⁴, Zhihong Chen⁵,
4 Jing Du⁶, Chao Ning⁷ and Deyun Feng^{*4}.

5 1Department of Pathology, the College of Basic Medicine, Xinjiang Medical University,
6 Urumqi, 830011, China.

7 2Department of Integrated Chinese and Western Medicine, Tumor Hospital, Xinjiang
8 Medical University, Urumqi, 830000, China.

9 3Department of Pathology, the Second Xiangya Hospital, Central South University,
10 Changsha, 410008, China.

11 4Department of Pathology, Xiangya Hospital, Central South University, Changsha,
12 410008, China.

13 5Department of Pathology, the People's Hospital of Hunan Province, Changsha,
14 410002, China.

15 6Department of Physiology, the College of Basic Medicine, Xinjiang Medical
16 University, Urumqi, 830011, China.

17 7Department of Function, Houbo College, Xinjiang Medical
18 University, Karamay, 834000, China.

19

20

21

22

1 **Abstract**

2 MicroRNAs (miRNAs) are involved in the regulation of multiple cellular pathways
3 and play a key role in the development and progression of tumor. Based on the
4 cellular function of their targets, miRNAs play the role of oncogenes or tumor
5 suppressor genes. Multiple studies have shown that abnormal expression of miRNAs
6 has close relation with the incidence of HCC, but the mechanism of miRNAs in HCC
7 still needs further research. In the present study, we showed that the overexpression
8 of miR-224 can reduce the mRNA and protein expression of ADAM17 and HOXA5,
9 the silencing of miR-224 can increase the protein expression of ADAM17 and HOXA5.
10 Dual luciferase reporter assays showed that miR-224 can directly regulate the
11 expression of ADAM17 and HOXA5. Importantly, we found that miR-224 positively
12 regulates cell migration and invasion in HCC, miR-224 overexpression can promote
13 the migration and invasion of BEL-7402 cell, and miR-224 silencing can suppress the
14 migration and invasion of BEL-7402 cell. miR-224 overexpression can result in the
15 redistribution of cell cycle, the cell percentage of S phase was increased significantly,
16 the cell percentage of G1 phase was decreased significantly, and there is no
17 noticeable change for the cell percentage of G2 phase. These results demonstrated
18 that it may be exert the function of oncogenes in a particular link of cancer cell growth.
19 In conclusion, these results suggest that miR-224 will become a promising biological
20 target in the treatment strategy of hepatocellular carcinoma (HCC).

21

22

23

1 **Introduction**

2 MicroRNAs(miRNAs), a member of the non-coding RNA family, are 19 to 25
3 nt-sized transcripts cleated from 70-100 nucleotide (nt) -sized hairpin shaped
4 precursors. Among distantly related organisms, the sequence of miRNAs is
5 conserved, suggesting that these molecules are involved in the basic biological
6 processes of the organism[1-3]. To date, more than 500 miRNAs have been
7 predicted to be expressed in humans [4, 5], and these miRNAs are estimated to
8 regulate the expression of over 5,000 human genes or 30% of human proteins[6].
9 Interactions between MiRNAs and their numerous target genes may represent
10 important levels of gene regulation in cells[7]. Although the exact function of miRNAs
11 is still unknown, existing studies have shown that miRNAs have a variety of different
12 expression patterns and can modulate many different developmental and
13 physiological processes. In addition, dysregulation of miRNAs expression may lead to
14 human disease[8]. Several studies have shown that close to 50% of miRNAs genes
15 are located in cancer-related genomic regions or fragile sites, which draws attention to
16 the importance of miRNAs in cancer [9-11].

17 Abnormal expression of MiRNAs has been shown to involve a variety of solid
18 tumors, including hepatocellular carcinoma, colon cancer, prostate cancer, breast
19 cancer, ovarian cancer and lung cancer[12-15]. In addition, abnormal expression of
20 miRNAs has been found to be associated with postoperative survival of lung cancer
21 patients[16], and can be used as a biomarker for the diagnosis and treatment of lung
22 cancer[17]. According to the role of their target genes, miRNAs can play an important

1 role in the occurrence and development of tumors as both tumor suppressor genes
2 and oncogenes[18]. The use of gene chip make it possible for the analysis of
3 large-scale miRNAs expression in tumor study, we can confirm a lot of miRNAs in
4 cancer tissues and normal tissues in the differences in expression by this way, so that
5 we can better explain the imbalance in the development of tumor miRNAs. Although
6 the function of some cancer-related miRNAs has been revealed, the mechanism of
7 action of most miRNAs in tumors remains unclear[19].

8 Some miRNAs have been proved to have the function of tumor suppressor genes.
9 Johnson SM et al[20] reported that the let family can regulate the expression of RAS
10 gene, revealing that it may play an inhibitory role in the development of lung cancer. O
11 'donnell et al[21] reported that miR-17-5p and miR-20a negatively regulate the
12 expression of E2F1, thereby affecting the function of c-myc gene and ultimately
13 controlling the signals of proliferation. Some miRNAs have been shown to have the
14 function of oncogenes, and miR-21 has been found to be up-regulated in both glioma
15 and breast cancer and to play an anti-apoptotic role[22, 23]. MiR-372 and miR-373
16 play the role of oncogenes in human testicular germ cell tumor, and in the presence of
17 wild-type P53, they can inhibit the P53 signaling pathway and allow the proliferation of
18 tumor cells[24]. Multiple studies have shown that miR-224 is overexpressed in human
19 tumors, such as colon cancer [25, 26], uterine cancer[27], glioma[28], breast cancer[29]
20 and hepatocellular carcinoma[30]. This suggests that up-regulated miR-224 may play
21 an important role in tumor formation. miR-224 can affect several key physiological
22 functions of cancer cells, including apoptosis[31], proliferation, migration and

1 invasion[32, 33], which proves its potential carcinogenicity.

2 Although we have some understanding of the expression and function of miR-224

3 in hepatocellular carcinoma based on some previous reports, the exact mechanism or

4 intracellular target of miR-224 in hepatocellular carcinoma is still poorly understood.

5 Therefore, the detection of the target of miR-224 is very important for our in-depth

6 understanding of the mechanism of action of miR-224. In this study, we will further

7 investigate the mechanism of miR-224 in the development of hepatocellular

8 carcinoma. We screened the target genes of miR-224 through bioinformatics, and

9 selected ADAM17 and HOXA5 as the starting points of this study, hoping to find

10 strong evidence that miR-224 plays an important role in the development of

11 hepatocellular carcinoma.

12

13

14 **Materials and Methods**

15 **Patient samples**

16 Paired hepatocellular carcinoma (HCC) tissues and the corresponding nearby

17 noncancerous livers were obtained from 19 patients who underwent liver resection.

18 Tissue samples were immediately snap-frozen in liquid nitrogen, and long-term stored

19 at minus 80 degrees Celsius in refrigerator. These 19 samples were mainly used for

20 fluorescence quantitative PCR detection. In addition,In addition, the

21 paraffin-embedded samples of 42 cases of hepatocellular carcinoma and 8 cases of

22 hepatic hemangioma were collected from the specimen bank archived in the

1 Department of Pathology, Xiangya Hospital, Central South University. The normal
2 liver tissue incidental to a specimen of hepatic hemangioma were used as control of
3 HCC and adjacent nontumorous tissues. The 42 samples of hepatocellular carcinoma
4 and 8 samples of hepatic hemangioma were mainly used for immunohistochemical
5 detection. The written informed consents were provided for all of the patients, and all
6 aspects of this study were approved by the Ethics Committee from Xiangya Hospital,
7 Central South University, Changsha, China.

8 The clinicopathologic features of 42 patients with paraffin-embedded hepatocellular
9 carcinoma samples were described briefly (S1 file). All of 42 patients contained 37
10 men and 5 women, the sex ratio(male/female) was 7:1. The mean age was 50 yeas
11 old. According to microscopic pathological characteristics, histological grade of tumor
12 differentiation was assigned: 17% were well-differentiated, 57% were
13 moderately-differentiated, 26% were poorly-differentiated. Most of patients (36 cases)
14 were single tumor, and only 5 cases were multiple tumors among 42 patients. The
15 biggest size of tumor can be up to 20cm, the smallest was just 1cm. About 57% of the
16 patients presented microscopic capsule and/or vascular invasion. Hepatic cirrhosis
17 was recorded for 67% of the patients. Here we may draw a conclusion that cirrhotic
18 liver is closely related to the occurrence of hepatocellular carcinoma.

19 **Cell culture**

20 SMMC-7721, BEL-7402, Huh-7, HL-7702 (L-02) and HEK-293T were purchased
21 from the Cell Bank of Chinese Academy of Sciences. SMMC-7721, BEL-7402 and
22 HL-7702 (L-02) were cultured in RPMI 1640 medium (Gibco, Invitrogen)

1 supplemented with 10% (v/v) fetal bovine serum (Gibco, Invitrogen), HuH-7 and
2 HEK-293T were maintained in DMEM high glucose medium (Gibco, Invitrogen)
3 supplemented with 10%(v/v) fetal bovine serum (Gibco, Invitrogen). All media
4 contained 100 units of penicillin/ml and 100 μ g of streptomycin/ml. Cells were
5 incubated at 37°C and supplemented with 5% CO₂ in the humidified chamber.

6 **RNA oligonucleotides and transfection**

7 miR-224 mimics and mimics Negative Control (mimics NC) were chemically
8 synthesized by Shanghai GenePharma Co.Ltd (Shanghai, China). miR-224 inhibitor
9 and inhibitor Negative Control(inhibitor NC) were obtained from Ambion, Inc (Austin,
10 TX, USA). Cells transfection was performed using siPORT™ NeoFX™ Transfection
11 Agent (ambition) following the manufacturer's protocol.

12 **Real-time quantitative PCR**

13 Total RNA was extracted from tissues and cultured cells using TRIzol (Invitrogen).
14 Real-time quantitative PCR (qPCR) was carried out to quantify the expression of
15 miR-224. TaqMan MicroRNA Transcription kit, TaqMan Universal Master Mix II and
16 TaqMan assays were used according to the manufacturer's protocol (Applied
17 Biosystems, USA). U6 snRNA was used as normalization control. Real-time
18 quantitative PCR (qPCR) was performed to detect the expression of ADAM17 and
19 HOXA5 . Reverse transcription was carried out using the PrimeScript® RT reagent Kit
20 with gDNA Eraser (TaKaRa), and cDNA products were amplified using SYBR ®
21 Premix ExTaqTM II Kit (TaKaRa), GAPDH was served as an internal control for total
22 cDNA content. The sequence of oligonucleotides used as PCR primers were:
23 ADAM17 (Forward): 5'- GCT GAC CCA GAT CCC ATG AAG-3', (Reverse): 5'-CTG
24 CAT TAT CCC ATG AAG TGT TCC-3'. HOXA5(Forward): 5'-GAT GCG CAA GCT
25 GCA CAT AAG-3', (Reverse) : 5'-CGG GTC AGG TAA CGG TTG AA-3'.

1 GAPDH(Forward) : 5'-GCA CCG TCA AGG CTG AGA AC-3', (Reverse) : 5'-TGG
2 TGA AGA CGC CAG TGG A-3'. The relative miRNA(miR-224) or mRNA(ADAM17
3 AND HOXA5) levels were calculated using the comparative cycle threshold [Ct, $2^{-\Delta\Delta Ct}$
4] method with their endogenous control.

5 **Immunohistochemistry (IHC)**

6 We carried out the Immunohistochemistry of Paraffin-embedded sections according to
7 a two-step protocol (Plink-2 plus@ Polymer HRP Detection System, GBI). Briefly, the
8 sections were first deparaffinized in xylene, then hydrated in a graded series of
9 ethanol (100%~50%) and tap water. The high pressure method was chosen to
10 perform antigen retrieval in citrate buffer (0.01M, pH 6.0), subsequently the sections
11 were incubated in 3% H₂O₂ at RT for 10 minutes so that endogenous peroxidase
12 activity was blocked. After the sections were washed in PBS, treated with
13 anti-ADAM17 antibody (Abcam, USA) at 4°C overnight. The sections were incubated
14 with polymer helper at 37°C for 20 minutes , subsequently incubated with poly-HRP
15 anti-Mouse IgG at the same temperature and duration. diaminobenzidine (DAB)
16 solution were used to stain the sections. The staining reactions were observed
17 carefully under a microscope and stopped with tap water in time. Finally, the sections
18 were counterstained with hematoxylin. Negative controls were obtained by omission
19 of the primary antibodies in all procedures of IHC.

20 The sections were observed by two independent pathologists. Then the staining of
21 immunohistochemistry was classified according to the percentage of cells with a
22 positive score for staining. Briefly, scoring method was shown below: The first, the
23 staining intensity was divided into four levels and scored (0, negative; 1, weak; 2,

1 moderate; 3, high). The second, the percentage of positive cells was divided into four
2 levels and scored (0, 0% positive cells; 1, <30% positive cells; 2, 30%-70% positive
3 cells; 3, >70% positive cells). The third, the sum of two scores for staining intensity
4 and percentage of cells was a final score of each sample. Samples were classified as:
5 (-) negative, the final scores were 0; (+) weak positive, the final scores were 1-2; (++)
6 moderate positive, the final scores were 3-4; (+++) strong positive, the final scores
7 were 5-6. HOX A5 demonstrated a membrane and cytoplasm staining. No signal was
8 seen with negative controls.

9 **Western blot analysis**

10 Cells were treated in lysis buffer (50 mM Tris-HCl, 0.5% sodium deoxycholate, 25 mM
11 EDTA) containing protease inhibitor cocktail (Roche) for 30 min on ice. Equal
12 amounts of protein were separated by 10% SDS-PAGE gels, and transferred onto
13 polyvinylidene difluoride membranes. After blocked with 5% skim milk solution for 2
14 hours, incubated with antibodies against ADAM17(Abcam) or HOXA5(Santa Cruz) or
15 β-actin (proteintech) at 4°C over night. The appropriate horseradish-conjugated
16 secondary antibody was incubated for 1 hour at room temperature, the signal of
17 protein was revealed using the chemiluminescence method (ECL, AURAGENE).
18 Band intensities were quantified using BIO-RAD image analysis software.

19 **Luciferase activity assay**

20 A 500bp sequence containing the predicted miR-224 binding site at the 3' UTRs of
21 ADAM17 or a 500bp sequence containing scrambled sequence cloned into the Xho
22 I/Not I site of psiCHECK-2 vector (Promega) to generate PsiCHECK-2-ADAM17-WT

1 and PsiCHECK-2 -ADAM17-mut vectors, respectively. Using the same approach, we
2 also obtained the PsiCHECK-2-HOXA5-WT and PsiCHECK-2 - HOXA5-mut vectors.
3 The whole construction process was completed by Shanghai jima pharmaceutical
4 technology co., LTD. The corresponding sequence information is as follows:
5 psiCHECKTM-2-ADAM17-WT: 5'-TGA AGA CTG GGAA GTGAC TT-3',
6 psiCHECKTM-2-ADAM17-MUT: 5'-TGA AGA CTG GGA GTGATT CA-3
7 psiCHECKTM-2-HOXA5-WT: 5'-TAC CTA TAT TCC TTG TGT AAT TAA TGC TGT
8 TGT AGAG GTGACTTG ATG AGA CAC AAC TTG TTC GAC GT-3'
9 psiCHECKTM-2-HOXA5-MUT: 5'-TAC CTA TAT TCC TTG TGT AAT TAA TGC TGT
10 TGT AGAG CACTGAAC ATG AGA CAC AAC TTG TTC GAC GT-3'
11 HEK-293T cells were cultured in 24-well plates, and PsiCHECK-2-ADAM17-WT,
12 PsiCHECK-2 -ADAM17-mut, PsiCHECK-2-HOXA5-WT, PsiCHECK-2 - HOXA5-mut
13 vectors were co-transfected respectively with miR-224 mimics or negative control
14 using Lipofectamine 2000 (Invitrogen) in 24-well plates. Luciferase assays were
15 performed using a luciferase assay kit (Promega) according to the manufacturer's
16 instructions. Firefly luciferase was used for normalization.

17 **Cell invasion and migration assay**

18 Transwell chamber (8 μ M pore diameter, Corning Costar) was used to perform the
19 assay. The filter was coated with matrigel (BD Biosciences) for examination of cell
20 invasion. However, the filter without coated matrigel was used for the migration assay.
21 The trasfected cells were resuspended in serum-free medium and added 200 μ L cell
22 resuspension solution (cell invasion: 1 \times 10⁵ cells per well, cell migration: 1 \times 10⁶ cells

1 per well) to the hydrated transwell chambers, simultaneously, 500 μ L complete
2 medium with 10% FBS was added to the bottom chambers. Cultured in the incubator
3 for 12h (cell migration) or 24h (cell invasion), removed the cells on the upper surface
4 of the membrane with cotton swab. Filters were fixed in 4% formaldehyde for 30 min
5 and stained with 0.1% crystal violet for 20 min. Cells were counted under microscope.
6 Experiments were repeated at least three times.

7 **Cell proliferation assay**

8 BEL-7402 cells (4000 cells/well) were plated in 96-well plates and incubated for 24 h.
9 MTT assay was performed to measure cell viability at 24h, 48h and 72h after
10 transfection. The absorbance at 490 nm was measured using a microplate reader
11 (Bio-Tek Instruments, Winooski, VT).

12 **Cell Cycle Analysis**

13 BEL-7402 cells were trypsinized at 48h after transfection, and washed in PBS, fixed
14 with 500 μ l of 70% ethanol at -20°C over night. Subsequently, cells were stained with
15 propidium iodide (PI) solution at 37°C for 30 min. Analysis was performed on a FACS
16 flow cytometer (BD Biosciences).

17 **Statistical analysis**

18 The two-tailed student's t-test, one-way ANOVA, Mann-Whitney U or Kruskal-Wallis
19 Test were used for data analysis. The statistical analyses were done using SPSS 19.0
20 for windows. P<0.05 was considered significant.

21 **Results**

22 **The expression of miR-224 in hepatocellular carcinoma and adjacent tissues**
23 Expression of miR-224 was detected in 19 pairs of human hepatocellular carcinoma

1 and adjacent liver tissues by real-time quantitative RT-PCR. Expression of miR-224
2 was upregulated in 2 cases of hepatocellular carcinoma tissues (>1.5 fold); It is
3 considered to be virtually unchanged in 3 cases of hepatocellular carcinoma tissues,
4 because the magnitude of increased or decreased miR-224 expression was less than
5 1.5 fold; the down-regulation of miR-224 was found in 14 cases of hepatocellular
6 carcinoma tissues (>1.5 fold), the statistical analysis showed that the expression of
7 miR-224 in hepatocellular carcinoma tissues was significantly higher than that in
8 adjacent liver tissues ($P<0.01$, Mann-Whitney U test) (Fig 1A). These results suggest
9 that miR-224 may play a role as an oncogene in hepatocellular carcinoma.
10 Fluorescence quantitative PCR was used to detect ADAM17 expression in 19
11 hepatocellular carcinoma and paracancer tissues. The results showed that ADAM17
12 expression in 6 hepatocellular carcinoma tissues was down-regulated (the reduction
13 was more than 1.5 times).ADAM17 expression in 8 hepatocellular carcinoma tissues
14 did not change significantly (the amplitude of up-regulation in 5 cases or
15 down-regulation in 2 cases was less than 1.5 times, and the ADAM17 expression in 1
16 case was not changed). ADAM17 expression was up-regulated (by more than 1.5
17 times) in 5 hepatocellular carcinoma tissues. Overall analysis showed that a total of
18 10 cases were upregulation, although the multiple of upregulation in 5 cases was
19 relatively small, statistical analysis also showed that ADAM17 expression in HCC
20 tissues was higher than that in adjacent tissues ($P<0.01$, Mann-Whitney U test) (Fig
21 1B) were statistically significant. Of course, we could also find that the number of
22 cases that were down-regulated in fact actually exceeded the number of cases that
23 were significant (the increase was more than 1.5 times). To achieve a more positive
24 result, it was necessary to increase the number of cases in the later period for
25 testing.These results suggest ADAM17 may act as an oncogene in hepatocellular
26 carcinoma.
27

28 **Expression of miR-224 in hepatocyte cancer cell lines**

29 Fluorescence quantitative PCR was used to detect the expression of miR-224 in

1 SMMC-7721, BEL-7402, Huh-7 and HL-7702 cells , and the results showed significant
2 differences in the expression of miR-224 in these cells ($P=0.001$, Kruskal-Wallis H test)
3 (Fig S1-A). Compared with HL-7702 cells, the expression of miR-224 was
4 up-regulated in SMMC-7721 cells ($P=0.025$, Mann-Whitney U test), while there was
5 no difference in BEL-7402 cells ($P=0.054$, Mann-Whitney U test), the expression of
6 miR-224 In Huh-7 cells was down-regulated ($P=0.004$, Mann-Whitney U test). The
7 expression of miR-224 in SMMC-7721 and BEL-7402 was higher than that in Huh-7
8 cells ($P=0.004$, both, Mann-Whitney U test), there was no significant difference
9 between SMMC-7721 and BEL-7402 cells ($P=0.423$, Mann-Whitney U test).

10 In order to understand the effect of mimics and inhibitors of miR-224 on hepatocyte
11 cancer cells, miR-224 mimics, mimics NC, miR-224 inhibitor and inhibitor NC were
12 transfected into SMMC-7721, BEL-7402 and Huh-7, and the expression of miR-224
13 was detected by fluorescence quantitative PCR 48h later. The results showed that the
14 expression of miR-224 in SMMC-7721, BEL-7402 and Huh-7 was approximately 50,
15 77 and 7545 times higher than that in the control group, respectively, after transfection
16 with miR-224 mimics. ($P<0.05$, all, Mann-Whitney U test)(Fig S1 B, C and D). While
17 compared with the control group, the expression of miR-224 in SMMC-7721,
18 BEL-7402 and Huh-7 decreased by approximately 19, 32 and 64 times, respectively,
19 after transfection with miR-224 inhibitor. ($P<0.05$, all, Mann-Whitney U test)(Fig S1 B,
20 C, D). The results suggested that the mimics and inhibitors of miR-224 could
21 effectively promote or inhibit the expression of miR-224 in hepatocyte cancer cells.

22 **Identification and validation of target genes of miR-224**

1 In order to better understand the mechanism of miR-224 in the development of HCC,
2 TargetScan[34], miRanda[35], PITA[36] and other computational methods were used
3 to predict the target genes of miR-224. Among the many predicted target genes,
4 ADAM17 and HOXA5 were selected as the targets for further study. The 3'UTR region
5 of the mRNA encoded by ADAM17 and HOXA5 genes contains sequences that are
6 partially complementary to miR-224 (Fig 2).

7 miR-224 mimics, mimics NC, miR-224 inhibitor and inhibitor NC were transfected
8 into SMMC-7721, BEL-7402 and Huh-7 cells, respectively, and the mRNA expression
9 levels of ADAM17 and HOXA5 were detected by fluorescence quantitative PCR and
10 Western blotting 48h later. The results showed that the overexpression of miR-224 in
11 SMMC-7721 and BEL-7402 could significantly reduce the mRNA expression of
12 ADAM17 and HOXA5 ($P<0.05$, all, the two-tailed Student's t-test)(Fig 3), while the
13 overexpression of miR-224 in Huh-7 had no significant effect on the mRNA
14 expression of ADAM17 and HOXA5 ($P>0.05$, all,The two-tailed Student's t-test (Fig 3).

15 Inhibition of miR-224 expression in three hepatocyte cancer cells could not effectively
16 increase the mRNA expression levels of ADAM17 and HOXA5 ($P>0.05$, all, the
17 two-tailed Student's t-test)(Fig 3).

18 The overexpression of miR-224 could cause the protein expression level of
19 ADAM17 in SMMC-7721, BEL-7402 and Huh-7 cells to decline, the protein
20 expression level of HOXA5 in SMMC-7721 and BEL-7402 cells decreased, but it had
21 no significant effect on the protein expression level of HOXA5 in Huh-7 cells. The
22 inhibition of miR-224 could cause the expression level of ADAM17 in SMMC-7721,

1 BEL-7402 and Huh-7 cells to rise, the expression level of HOXA5 in SMMC-7721 and
2 BEL-7402 cells to rise, but had no significant effect on the expression level of HOXA5
3 in Huh-7 cells (Fig 4).

4 In order to detect the direct interaction between miR-224 and its target genes
5 ADAM17 and HOXA5, we co-transfected miR-224 mimics or mimics NC with wild-type
6 or mutant vectors of ADAM17 and HOXA5 into 293T cells, and then detected the
7 changes of double luciferase of the vector. We found that the fluorescence activity of
8 the wild-type reporting genomes of ADAM17 and HOXA5 was significantly lower than
9 that of the control group ($P<0.05$, one-way ANOVA) (Fig 5). This result suggests that
10 miR-224 can directly regulate ADAM17 and HOXA5.

11 **Regulation of miR-224 to Cell Migration and Invasion Viability**

12 Combining the effects of miR-224 mimics and inhibitor on three strains of HCC,
13 the regulatory effects of two target genes, and the biological characteristics of cancer
14 cells, BEL-7402 cells were selected to complete the biological function test of
15 miR-224 overexpression or inhibition.

16 We evaluated the effect of miR-224 on cell migration and invasion ability in vitro
17 by enhancing or silencing the expression of miR-224. 48h after transfection with
18 miR-224 mimics or mimic NC into BEL-7402 cells, the effect of miR-224
19 overexpression on the migration and invasion ability of BEL-7402 cells was detected
20 by Transwell assay. The results showed that compared with the mimics NC group, the
21 migration ($P<0.01$, the two-tailed Student's t-test) (Fig 6A) and invasion ($P<0.01$, the
22 two-tailed Student's t-test) (Fig 7A) capabilities of the miR-224 mimics group were
23 significantly enhanced. 48h after transfection of BEL-7402 cells with miR-224 inhibitor

1 or inhibitor NC, the effect of miR-224 silencing on the migration and invasion ability of
2 bel-7402 cells was determined by Transwell assay. The results showed that
3 compared with the inhibitor NC group, the ability of miR-224 inhibitor group to migrate
4 ($P < 0.01$, the two-tailed Student's t-test) (Fig 6B) and to invade ($P < 0.01$, the
5 two-tailed Student's t-test) (Fig 7B) was significantly reduced.

6 **Effect of miR-224 on proliferation of BEL-7402 cells**

7 In order to detect whether the abnormal expression of miR-224 had an impact on
8 the growth of cancer cells during the development of HCC, we transfected miR-224
9 mimics, mimic NC, miR-224 inhibitor and inhibitor NC into BEL-7402 cells, and
10 detected the cell activity by MTT method at 24h, 48h and 72h after transfection,
11 respectively. The results showed that overexpression or silencing of miR-224 had no
12 significant effect on the proliferation of BEL-7402 cells ($P > 0.05$, the two-tailed
13 Student's t-test)(Fig 8).

14 **Effect of miR-224 on the cell cycle of BEL-7402**

15 In order to detect whether the abnormal expression of miR-224 had an impact on
16 the distribution of cell cycle during the development of HCC, we transfected miR-224
17 mimics, mimic NC, miR-224 inhibitor and inhibitor NC into BEL-7402 cells, and
18 detected the cell cycle changes by flow cytometry 48h after transfection, we found that
19 miR-224 mimics could reduce the cell percentage in G1 phase of the cell cycle (P
20 < 0.05 , the two-tailed Student's t-test) (Fig 9A) and increase the cell percentage in S
21 phase ($P < 0.01$, the two-tailed Student's t-test) (Fig 9A), but had no significant effect
22 on the cell percentage composition in G2 phase ($P = 0.682$, The two-tailed Student's

1 t-test (Fig 9A).In addition, after miR-224 was silenced, flow results showed that the
2 cell percentage of miR-224 inhibitor group in G1, S and G2 phases did not change
3 significantly compared with that of inhibitor NC group ($P>0.05$, all, the two-tailed
4 Student's t-test)(Fig 9B).These results suggest that miR-224 overexpression can
5 promote the proliferation of bel-7402 cells.

6 **Discussion**

7 Hepatocellular carcinoma (HCC) is one of the most common malignant tumors,Its
8 morbidity and mortality rates have been on the rise in recent years[37], the high
9 recurrence or metastasis rate after surgery may hinder the improvement of the
10 survival time of hepatocellular carcinoma patients[37]。 In order to get a better method
11 to detect hepatocellular carcinoma, a biological target is needed to effectively screen
12 high-risk patients for early diagnosis so that interventions can be made earlier,
13 increasing the likelihood of a cure.MiRNAs appear as important regulators of genes,
14 and ongoing studies on their functions and target genes reveal that MiRNAs play a
15 key role in several important cellular functions. MiRNAs possess several
16 characteristics that make them possible targets for early diagnosis of tumors or
17 powerful tools for gene therapy, which will bring new hope for the diagnosis and
18 treatment of tumors[38] ,this is very attractive to researchers.

19 miR-224 is present in mammals, including humans, and is located on the X
20 chromosome[39]. It has been reported that miR-224 is abnormally expressed in a
21 variety of tumors, suggesting that it may play an important role in the biological
22 process of tumors.According to the different malignant phenotypes of tumor cells, the

1 biological role of miR-224 is also different, and it also shows contradictory properties
2 in different studies. Several reports have confirmed that the expression of miR-224 in
3 hepatocellular carcinoma is up-regulated, and overexpression of miR-224 can
4 promote proliferation and invasion of hepatocellular carcinoma cells[30, 40] .However,
5 in prostate cancer, the expression of miR-224 is down-regulated and inhibits the
6 migration and invasion of prostate cancer cells by regulating TPD52[41]. In addition,
7 overexpression of miR-224 in adult neurohemangiomas can inhibit cell
8 proliferation[42]. These results suggest that the mechanism of miR-224 in tumors is
9 complex, requiring further study.

10 In this study, we used a variety of methods to identify and confirm the target genes of
11 miR-224 and their mechanism of action in hepatocarcinoma. The up-regulation of
12 miR-224 can reduce the mRNA and protein expression levels of ADAM17 and
13 HOXA5, and the double luciferase experiment further proved that miR-224 has a
14 direct targeting regulation effect on its target genes ADAM17 and HOXA5. Importantly,
15 in the functional test of miR-224, we found that overexpression of miR-224
16 significantly enhanced the migration and invasion ability of Bel-7402 cells.In contrast,
17 silencing of miR-224 inhibited the migration and invasion of Bel-7402 cells. In addition,
18 overexpression of miR-224 can change the cell cycle of Bel-7402 cells, and the cell
19 proportion of S phase increases significantly, while the cell proportion of G1 phase
20 decreases significantly, while the cell proportion of G2 phase remains unchanged,
21 suggesting that overexpression of miR-224 can promote the proliferation of Bel-7402
22 cells.The reason for this result may be that overexpression of miR-224 causes

1 changes in genes related to G1/S transformation and DNA replication, but further
2 studies are needed to prove this. Paradoxically, the silencing of miR-224 had no
3 meaningful effect on the cell cycle. In addition, in the analysis of MTT experimental
4 results, we failed to find evidence of the effect of overexpression or silencing of
5 miR-224 on the proliferation of Bel-7402 cells. This may be due to the lack of an
6 experimental strategy for stable overexpression or silencing of genes that hinders the
7 analysis of these experiments, although individual differences between cell models
8 should also be fully considered.

9 Using bioinformatics to predict miRNAs target genes is a convenient method for
10 preliminary screening of target genes, which can provide us with many candidate
11 target genes. Among these predicted target genes, many target genes have been
12 experimentally proved to play very important roles in the occurrence and development
13 of tumors. These target genes of miR-224 with rich functional categories further
14 confirmed that miR-224 may be directly or indirectly involved in the biological process
15 of tumors. It remains a difficult task to fully understand the mechanisms by which
16 miRNAs dysfunction may promote tumors, and although bioinformatics tools can
17 help reveal the putative mRNA target genes, experimental data confirmation remains
18 a necessary tool. Some target genes of miR-224 have been reported, including
19 RKIP[43], p21[44], Cdc42[45], TPD52[41], TRIB1[46]、HOXD10[47], SMAD4[48],
20 API-5[31] and CXCR4[49]. Some of these target genes promote tumor development,
21 while others do not. In this study, the data showed that miR-224 was involved in the
22 migration and invasion of liver cancer cells, and its direct regulatory relationship with

1 the two target genes ADAM17 and HOXA5 has also been proved. We need to further
2 analyze which pathway actually plays a regulatory role in the migration and invasion
3 of liver cancer cells. Our previous studies have found that ADAM17 is overexpressed
4 in HCC tissues, and this abnormal expression is associated with microvascular
5 invasion of HCC[50]. The ANG II -EGFR signaling pathway mediated by ADAMs
6 (ADAM9 and ADAM17) can accelerate the proliferation and infiltration of hepatocyte
7 cancer cells huh-7 and PLC/PRF/5, and ADAMs inhibitors can inhibit the role of this
8 pathway[51].In this study, upregulation of miR-224 inhibited the expression of
9 ADAM17, while the decreased expression of ADAM17 inhibited the migration and
10 invasion of cancer cells. Therefore, the enhancement of migration and invasion of
11 Bel-7402 cells by miR-224 may not be realized by directly regulating the target gene
12 ADAM17.

13 HOXA5 is down-regulated in hepatocellular carcinoma, suggesting that it may be a
14 tumor suppressor gene in hepatocellular carcinoma.In addition, it has also been
15 reported that HOXA5 is also low-expressed in non-small cell lung cancer [52]and oral
16 squamous cell carcinoma[53]. As the upstream regulatory network molecule of
17 AMELY (Amelogenin y-linked), HOXA5 is involved in the regulation of cell proliferation
18 and angiogenesis and other biological functions of hepatocellular carcinoma[54].

19 Inhibition of HOXA5 can lead to increased proliferation, migration and invasion of lung
20 cancer A549 cells[55]. In this study, overexpression of miR-224 reduced the
21 expression levels of HOXA5 mRNA and protein, while silencing of miR-224 increased
22 the expression levels of HOXA5 mRNA and protein. This is consistent with the trend

1 that miR-224 regulates the migration and invasion of hepatocyte cancer cells
2 BEL-7402. Therefore, miR-224 may enhance the migration and invasion of cancer
3 cells by directly regulating the expression of target gene HOXA5.
4 Tumor invasion involves several different steps[56], miR-224 may also regulate other
5 target genes associated with tumor invasion, which is certainly a very complex
6 information network. Our findings indicate that miR-224 plays a very important role in
7 the occurrence and development of hepatocellular carcinoma. Further understanding
8 of the mechanism of action of miR-224 and other target genes in hepatocellular
9 carcinoma is conducive to the exploration of its exact role in the initiation and
10 progression of hepatocellular carcinoma, which provides us with new research
11 directions and challenges.

12 **Acknowledgements**

13 We thank Teacher Xiaojing Yang and Dr Zhaojun Duan for her excellent technical
14 assistance.

15 **Funding**

16 This work was supported by Xinjiang Uygur Autonomous Region Natural Science
17 Fund (No.2016D01C165).

18 **Figs legends**

19 **Fig 1. Expression of miR-224 in hepatocellular carcinoma.** The relative levels of
20 miR-224 expression in hepatocellular carcinoma tissues and adjacent liver tissues
21 were analyzed by real time PCR, and normalized to small nuclear RNA U6. (P<0.01)
22 (T/N-Tumor/Normal)

1 **Fig 2. The predicted binding sites of miR-224 and target genes ADAM17 and**
2 **HOXA5.** The binding site of ADAM17 to miR-224 was predicted by miRanda (V1.9),
3 and the binding site of HOXA5 to miR-224 was predicted by TargetScan (v4.1).

4 **Fig 3. mRNA expression changes of target genes ADAM17 and HOXA5 after**
5 **overexpression or inhibition of miR-224.** A,B,C. mRNA expression changes of
6 ADAM17 in SMMC-7721, BEL-7402 and Huh-7 cells after overexpression or inhibition
7 of miR-224. D,E,F. mRNA expression changes of HOXA5 in SMMC-7721, BEL-7402
8 and Huh-7 cells after overexpression or inhibition of miR-224. (*,P< 0.05, **,P< 0.01)

9 **Fig 4. Effect of overexpression or inhibition of miR-224 on protein expression**
10 **levels of its target genes ADAM17 and HOXA5.** SMMC-7721, BEL-7402 and
11 HuH-7 cells were transfected with miR-224 mimics, mimics negative control(NC),
12 miR-224 inhibitor and inhibitor negative control(NC), at 48 hours after trasfectedon,
13 protein expression of ADAM17 and HOXA5 were analyzed by Western blotting, as
14 compared to negative control (NC). ADAM17 protein has two subunits, the molecular
15 weight of 93 kDa and 79 kDa, respectively. HOXA5 protein has also two subunits,
16 the molecular weight of 55 kDa and 43 kDa, respectively.

17 **Fig 5. Double luciferase activity of target genes ADAM17 and HOXA5 of miR-224**
18 **was detected.** A. Luciferase activity of wild type or mutant ADAM17 3'UTR reporter
19 gene in HEK-293T cells transfected with miR-224 mimics or mimics negative
20 control(NC) was detemined. The results showed that luciferase activity of wild type
21 ADAM17 3'UTR transfected with miR-224 mimics was decreased compare to others
22 group in HEK-293T cells. B. Luciferase activity of wild type or mutant HOXA5 3'UTR

1 reporter gene in HEK-293T cells transfected with miR-224 mimics or mimics negative
2 control(NC) was determined. The results showed that luciferase activity of wild type
3 HOXA5 3'UTR transfected with miR-224 mimics was decreased compare to others
4 group in HEK-293T cells. (*, P<0.05, **, P<0.01)

5 **Fig 6. miR-224 affects the migration ability of BEL -7402 cells.** A. The
6 overexpression of miR-224 promoted the migration ability of BEL-7402 cells. B.
7 miR-224 silencing inhibited the migration of BEL-7402 cells. (*,P< 0.05, **,P< 0.01)

8 **Fig 7. miR-224 affects the invasion ability of BEL -7402 cells.** A. The
9 overexpression of miR-224 promoted the invasion ability of BEL-7402 cells. B.
10 miR-224 silencing inhibited the invasion of BEL-7402 cells. (*,P< 0.05, **,P< 0.01)

11 **Fig 8. Effect of miR-224 on proliferation of bel-7402 cells.** A. The overexpression
12 of miR-224 had no effect on the proliferation of BEL-7402 cells. B. The silencing of
13 miR-224 had no effect on the proliferation of BEL-7402 cells.

14 **Fig 9. The effect of miR-224 on the cell cycle of bel-7402.** A. The effect of
15 overexpression of miR-224 on the cell cycle of BEL-7402, Cell cycle analysis was
16 performed by FCM (flow cytometry), results showed that miR-224 mimics could
17 reduce the cell percentage in G1 phase of the cell cycle and increase the cell
18 percentage in S phase , but had no significant effect on the cell percentage

19 composition in G2 phase B. The effect of miR-224 silencing on the cell cycle of
20 BEL-7402, Cell cycle analysis was performed by FCM (flow cytometry), results
21 showed that the cell percentage of miR-224 inhibitor group in G1, S and G2 phases
22 did not change significantly compared with that of inhibitor NC group. (*.P< 0.05, **P<

1 0.01).

2

3 **Supporting information**

4 **S1 Fig Expression of miR-224 in hepatocyte cancer cells.** A. Expression of
5 miR-224 in hepatocyte cancer cells SMMC-7721, BEL-7402, Huh-7 and normal
6 human hepatocytes HL-7702. B,C,D. after transfection of miR-224 mimics and
7 inhibitor into SMMC-7721, BEL-7402 and Huh-7, the expression level of miR-224 was
8 significantly increased or decreased compared with the negative control group. (*,P<
9 0.05, **,P< 0.01)

10 **References:**

11 [1]. Pasquinelli, A.E., et al., Conservation of the sequence and temporal expression of let-7
12 heterochronic regulatory RNA. *Nature*, 2000. 408(6808): p. 86-9.
13 [2]. Ke, X.S., et al., MicroRNAs: key participants in gene regulatory networks. *Curr Opin Chem Biol*,
14 2003. 7(4): p. 516-23.
15 [3]. Moss, E.G., Silencing unhealthy alleles naturally. *Trends Biotechnol*, 2003. 21(5): p. 185-7.
16 [4]. Griffiths-Jones, S., The microRNA Registry. *Nucleic Acids Res*, 2004. 32(Database issue): p.
17 D109-11.
18 [5]. Griffiths-Jones, S., et al., miRBase: microRNA sequences, targets and gene nomenclature. *Nucleic
19 Acids Res*, 2006. 34(Database issue): p. D140-4.
20 [6]. Lewis, B.P., C.B. Burge and D.P. Bartel, Conserved seed pairing, often flanked by adenosines,
21 indicates that thousands of human genes are microRNA targets. *Cell*, 2005. 120(1): p. 15-20.
22 [7]. Ambros, V., The functions of animal microRNAs. *Nature*, 2004. 431(7006): p. 350-5.
23 [8]. He, L. and G.J. Hannon, MicroRNAs: small RNAs with a big role in gene regulation. *Nat Rev
24 Genet*, 2004. 5(7): p. 522-31.
25 [9]. Calin, G.A., et al., MicroRNA profiling reveals distinct signatures in B cell chronic lymphocytic
26 leukemias. *Proc Natl Acad Sci U S A*, 2004. 101(32): p. 11755-60.
27 [10]. Calin, G.A., et al., Human microRNA genes are frequently located at fragile sites and genomic
28 regions involved in cancers. *Proc Natl Acad Sci U S A*, 2004. 101(9): p. 2999-3004.
29 [11]. Sevignani, C., et al., Mammalian microRNAs: a small world for fine-tuning gene expression.
30 *Mamm Genome*, 2006. 17(3): p. 189-202.
31 [12]. Chen, P.J., et al., Androgen pathway stimulates microRNA-216a transcription to suppress the
32 tumor suppressor in lung cancer-1 gene in early hepatocarcinogenesis. *Hepatology*, 2012. 56(2): p.
33 632-43.
34 [13]. Fu, J., et al., Identifying microRNA-mRNA regulatory network in colorectal cancer by a

1 combination of expression profile and bioinformatics analysis. *BMC Syst Biol*, 2012. 6: p. 68.

2 [14]. Lichner, Z., et al., Pleiotropic action of renal cell carcinoma-dysregulated miRNAs on
3 hypoxia-related signaling pathways. *Am J Pathol*, 2012. 180(4): p. 1675-87.

4 [15]. Fellenberg, J., et al., A microRNA signature differentiates between giant cell tumor derived
5 neoplastic stromal cells and mesenchymal stem cells. *Cancer Lett*, 2012. 321(2): p. 162-8.

6 [16]. Takamizawa, J., et al., Reduced expression of the let-7 microRNAs in human lung cancers in
7 association with shortened postoperative survival. *Cancer Res*, 2004. 64(11): p. 3753-6.

8 [17]. Yanaihara, N., et al., Unique microRNA molecular profiles in lung cancer diagnosis and prognosis.
9 *Cancer Cell*, 2006. 9(3): p. 189-98.

10 [18]. Esquela-Kerscher, A., et al., Post-embryonic expression of *C. elegans* microRNAs belonging to the
11 lin-4 and let-7 families in the hypodermis and the reproductive system. *Dev Dyn*, 2005. 234(4): p.
12 868-77.

13 [19]. Calin, G.A. and C.M. Croce, MicroRNA-cancer connection: the beginning of a new tale. *Cancer Res*,
14 2006. 66(15): p. 7390-4.

15 [20]. Johnson, S.M., et al., RAS is regulated by the let-7 microRNA family. *Cell*, 2005. 120(5): p.
16 635-47.

17 [21]. O'Donnell, K.A., et al., c-Myc-regulated microRNAs modulate E2F1 expression. *Nature*, 2005.
18 435(7043): p. 839-43.

19 [22]. Chan, J.A., A.M. Krichevsky and K.S. Kosik, MicroRNA-21 is an antiapoptotic factor in human
20 glioblastoma cells. *Cancer Res*, 2005. 65(14): p. 6029-33.

21 [23]. Iorio, M.V., et al., MicroRNA gene expression deregulation in human breast cancer. *Cancer Res*,
22 2005. 65(16): p. 7065-70.

23 [24]. Voorhoeve, P.M., et al., A genetic screen implicates miRNA-372 and miRNA-373 as oncogenes in
24 testicular germ cell tumors. *Cell*, 2006. 124(6): p. 1169-81.

25 [25]. Arndt, G.M., et al., Characterization of global microRNA expression reveals oncogenic potential of
26 miR-145 in metastatic colorectal cancer. *BMC Cancer*, 2009. 9: p. 374.

27 [26]. Wang, Y.X., et al., Initial study of microRNA expression profiles of colonic cancer without lymph
28 node metastasis. *J Dig Dis*, 2010. 11(1): p. 50-4.

29 [27]. Shen, S.N., et al., Upregulation of microRNA-224 is associated with aggressive progression and
30 poor prognosis in human cervical cancer. *Diagn Pathol*, 2013. 8: p. 69.

31 [28]. Lu, S., et al., Upregulation of microRNA-224 confers a poor prognosis in glioma patients. *Clin
32 Transl Oncol*, 2013. 15(7): p. 569-74.

33 [29]. Zhou, F., et al., MicroRNA and histopathological characterization of pure mucinous breast
34 carcinoma. *Cancer Biol Med*, 2013. 10(1): p. 22-7.

35 [30]. Zhang, Y., et al., Involvement of microRNA-224 in cell proliferation, migration, invasion, and
36 anti-apoptosis in hepatocellular carcinoma. *J Gastroenterol Hepatol*, 2013. 28(3): p. 565-75.

37 [31]. Wang, Y., et al., Profiling microRNA expression in hepatocellular carcinoma reveals
38 microRNA-224 up-regulation and apoptosis inhibitor-5 as a microRNA-224-specific target. *J Biol Chem*,
39 2008. 283(19): p. 13205-15.

40 [32]. Li, Q., et al., MicroRNA-224 is upregulated in HepG2 cells and involved in cellular migration and
41 invasion. *J Gastroenterol Hepatol*, 2010. 25(1): p. 164-71.

42 [33]. Li, X., et al., Regulation of miRNA expression by Src and contact normalization: effects on
43 nonanchored cell growth and migration. *Oncogene*, 2009. 28(48): p. 4272-83.

44 [34]. Friedman, R.C., et al., Most mammalian mRNAs are conserved targets of microRNAs. *Genome*

1 Res, 2009. 19(1): p. 92-105.

2 [35]. John, B., et al., Human MicroRNA targets. *PLoS Biol*, 2004. 2(11): p. e363.

3 [36]. Kertesz, M., et al., The role of site accessibility in microRNA target recognition. *Nat Genet*, 2007. 39(10): p. 1278-84.

4 [37]. Siegel, R., et al., Cancer statistics, 2014. *CA Cancer J Clin*, 2014. 64(1): p. 9-29.

5 [38]. Schaefer, A., et al., Diagnostic and prognostic implications of microRNA profiling in prostate
6 carcinoma. *Int J Cancer*, 2010. 126(5): p. 1166-76.

7 [39]. Landgraf, P., et al., A mammalian microRNA expression atlas based on small RNA library
8 sequencing. *Cell*, 2007. 129(7): p. 1401-14.

9 [40]. Wang, Y., et al., MicroRNA-224 is up-regulated in hepatocellular carcinoma through epigenetic
10 mechanisms. *FASEB J*, 2012. 26(7): p. 3032-41.

11 [41]. Goto, Y., et al., Tumour-suppressive microRNA-224 inhibits cancer cell migration and invasion via
12 targeting oncogenic TPD52 in prostate cancer. *FEBS Lett*, 2014. 588(10): p. 1973-82.

13 [42]. Gokhale, A., et al., Distinctive microRNA signature of medulloblastomas associated with the WNT
14 signaling pathway. *J Cancer Res Ther*, 2010. 6(4): p. 521-9.

15 [43]. Liu, H., et al., RKIP suppresses gastric cancer cell proliferation and invasion and enhances
16 apoptosis regulated by microRNA-224. *Tumour Biol*, 2014.

17 [44]. Wang, H., et al., MiR-224 promotes the chemoresistance of human lung adenocarcinoma cells to
18 cisplatin via regulating G(1)/S transition and apoptosis by targeting p21(WAF1/CIP1). *Br J Cancer*, 2014.
19 111(2): p. 339-54.

20 [45]. Ke, T.W., et al., MicroRNA-224 suppresses colorectal cancer cell migration by targeting Cdc42.
21 *Dis Markers*, 2014. 2014: p. 617150.

22 [46]. Lin, Z.Y., et al., MicroRNA-224 inhibits progression of human prostate cancer by downregulating
23 TRIB1. *Int J Cancer*, 2014. 135(3): p. 541-50.

24 [47]. Li, Q., et al., miR-224 promotion of cell migration and invasion by targeting Homeobox D 10 gene
25 in human hepatocellular carcinoma. *J Gastroenterol Hepatol*, 2014. 29(4): p. 835-42.

26 [48]. Wang, Y., et al., MicroRNA-224 targets SMAD family member 4 to promote cell proliferation and
27 negatively influence patient survival. *PLoS One*, 2013. 8(7): p. e68744.

28 [49]. Zhu, S., et al., Ubc9 promotes breast cell invasion and metastasis in a sumoylation-independent
29 manner. *Oncogene*, 2010. 29(12): p. 1763-72.

30 [50]. Liu, Y., et al., Expression of miR-224, miR-145, and their putative target ADAM17 in
31 hepatocellular carcinoma. *Acta Biochim Biophys Sin (Shanghai)*, 2014. 46(8): p. 720-2.

32 [51]. Itabashi, H., et al., Angiotensin II and epidermal growth factor receptor cross-talk mediated by a
33 disintegrin and metalloprotease accelerates tumor cell proliferation of hepatocellular carcinoma cell lines.
34 *Hepatol Res*, 2008. 38(6): p. 601-13.

35 [52]. Kim, D.S., et al., Epigenetic inactivation of Homeobox A5 gene in nonsmall cell lung cancer and its
36 relationship with clinicopathological features. *Mol Carcinog*, 2009. 48(12): p. 1109-15.

37 [53]. Rodini, C.O., et al., Homeobox gene expression profile indicates HOXA5 as a candidate prognostic
38 marker in oral squamous cell carcinoma. *Int J Oncol*, 2012. 40(4): p. 1180-8.

39 [54]. Qi, L., et al., Activated amelogenin Y-linked (AMELY) regulation and angiogenesis in human
40 hepatocellular carcinoma by biocomputation. *Oncol Lett*, 2013. 5(3): p. 1075-1079.

41 [55]. Liu, X.H., et al., MicroRNA-196a promotes non-small cell lung cancer cell proliferation and
42 invasion through targeting HOXA5. *BMC Cancer*, 2012. 12: p. 348.

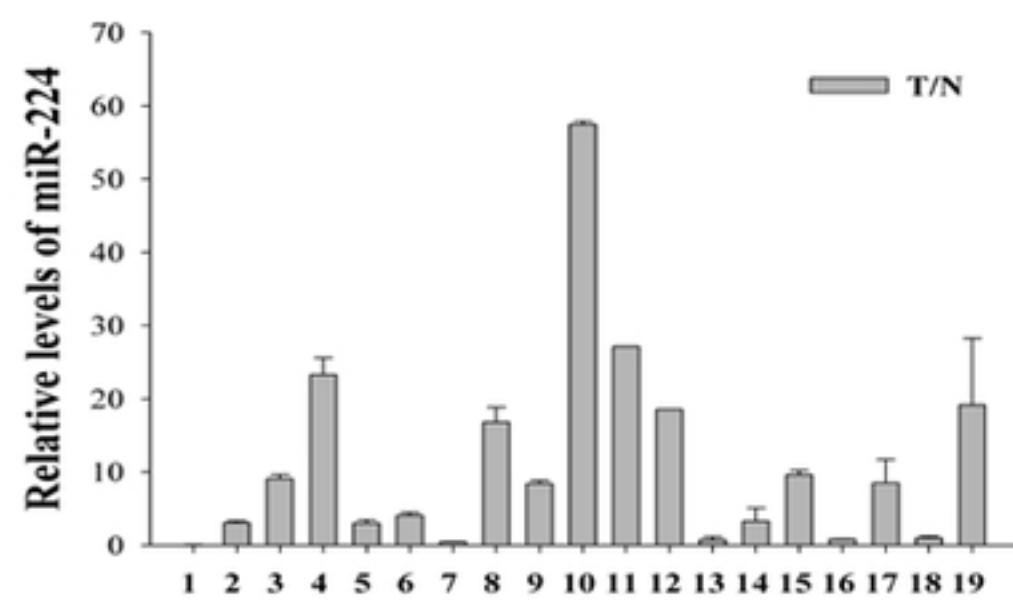
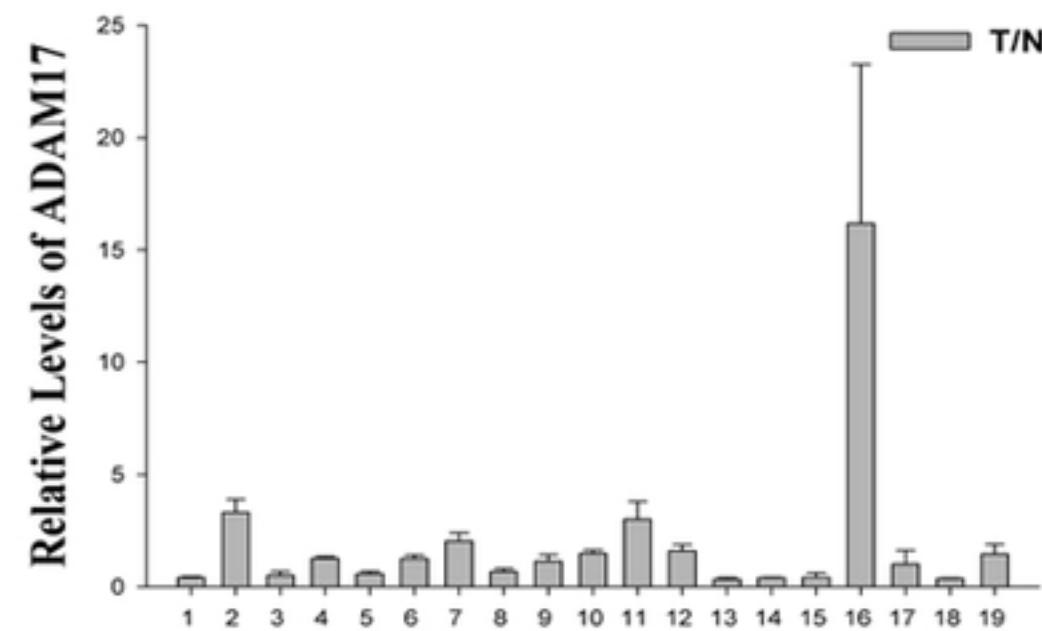
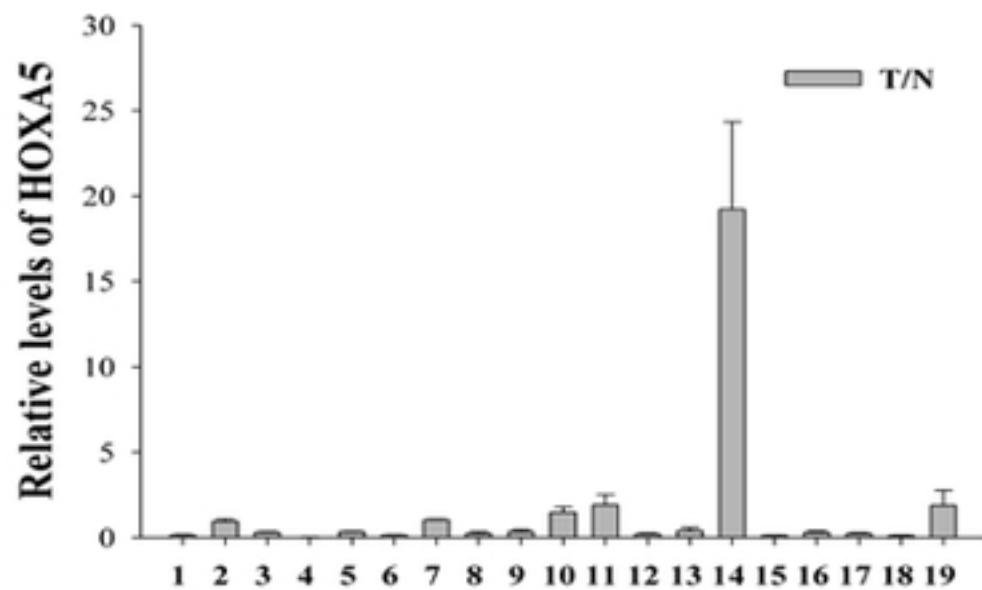
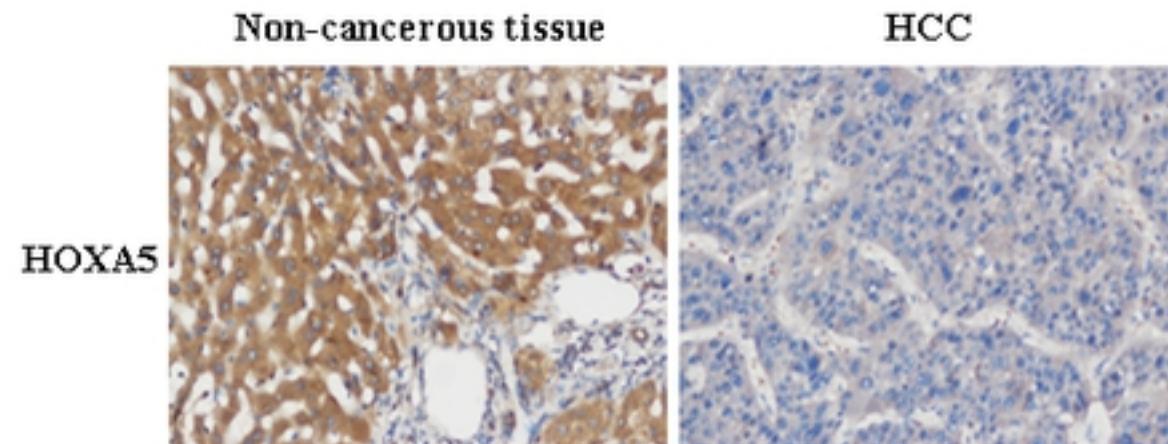
43 [56]. Fidler, I.J., The pathogenesis of cancer metastasis: the 'seed and soil' hypothesis revisited. *Nat Rev*

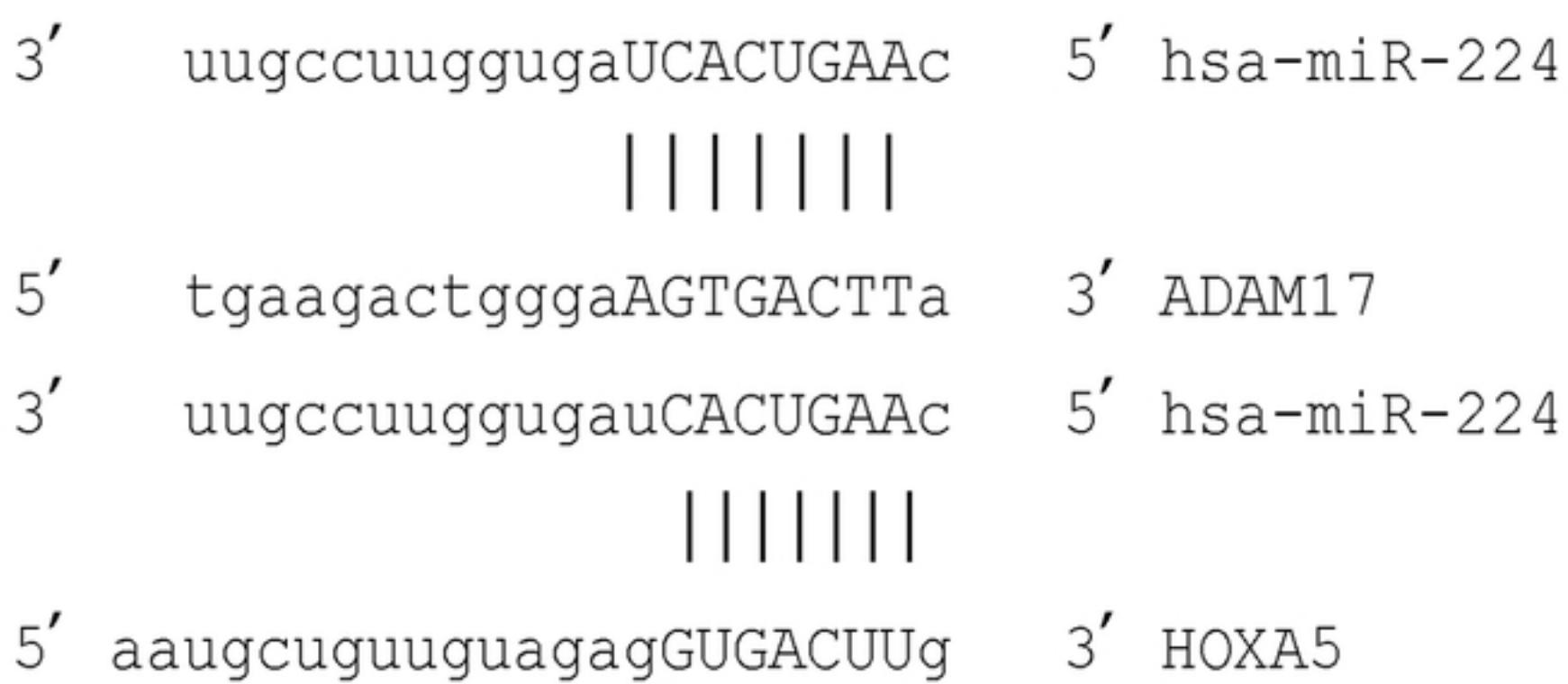
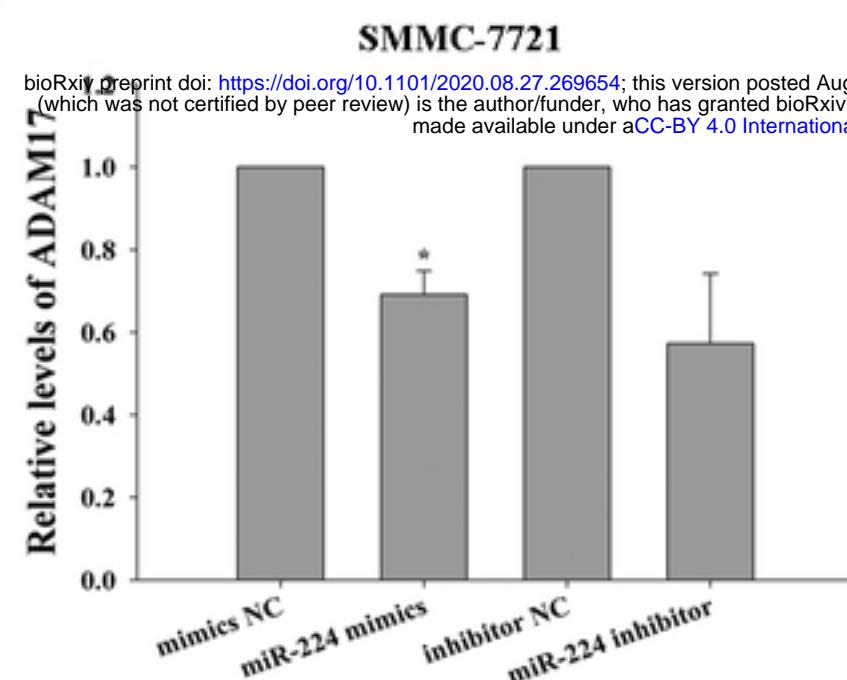
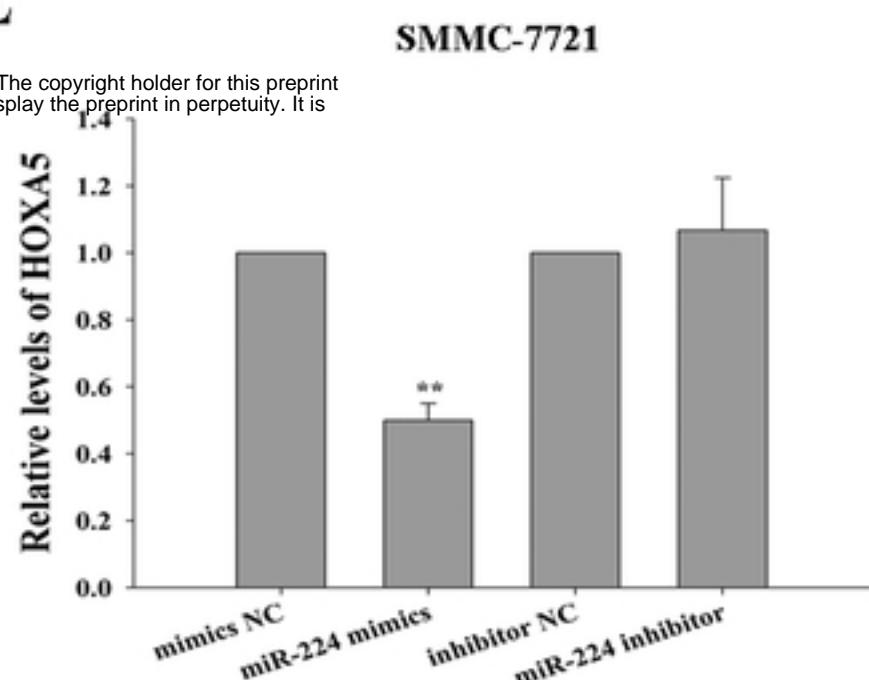
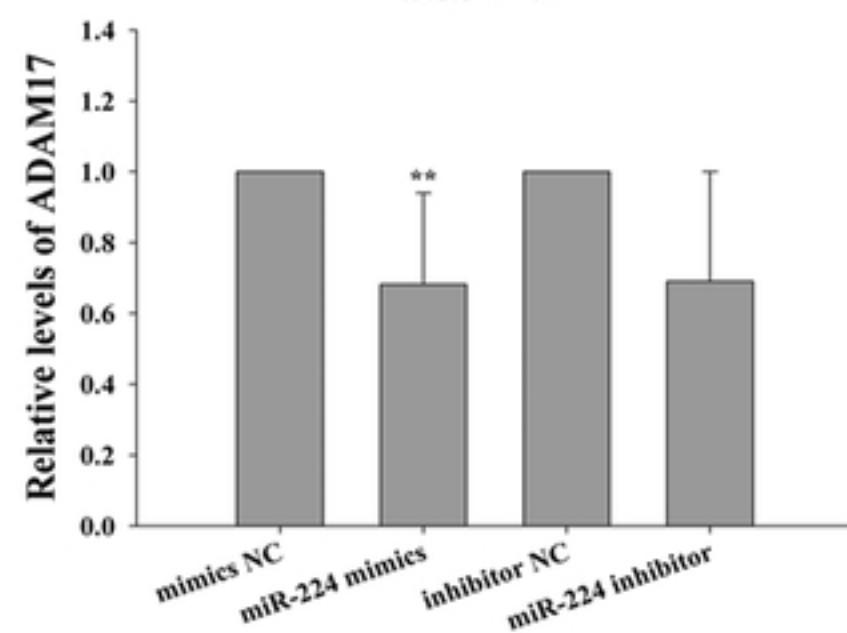
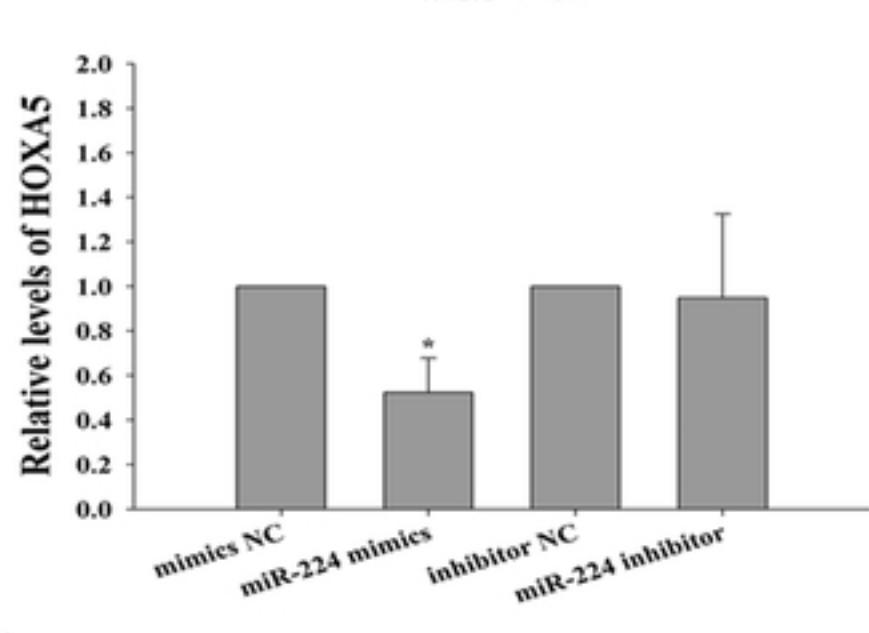
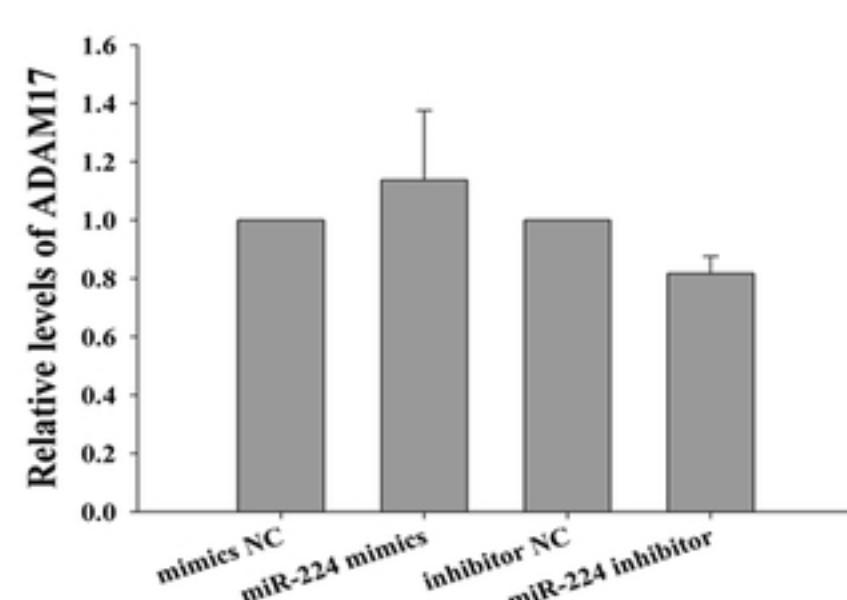
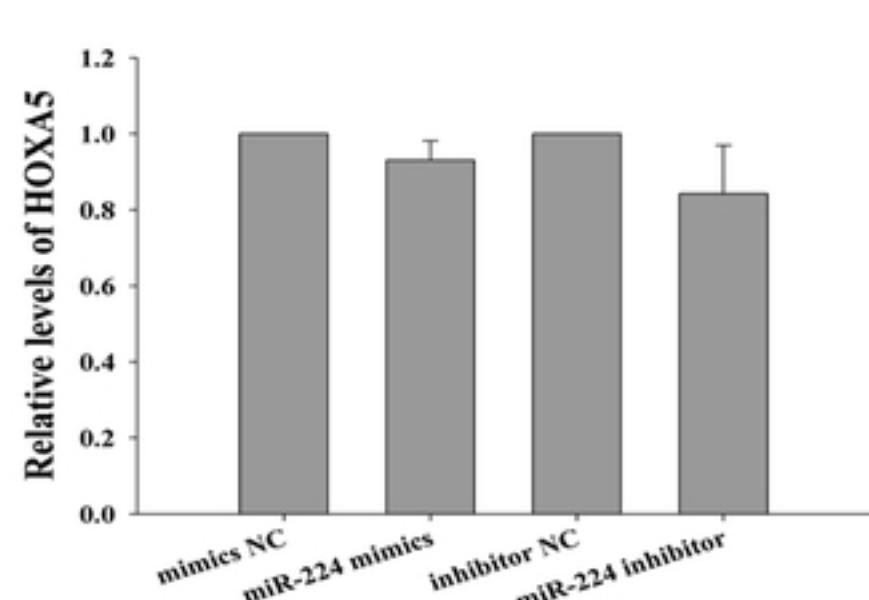
1 Cancer, 2003. 3(6): p. 453-8.

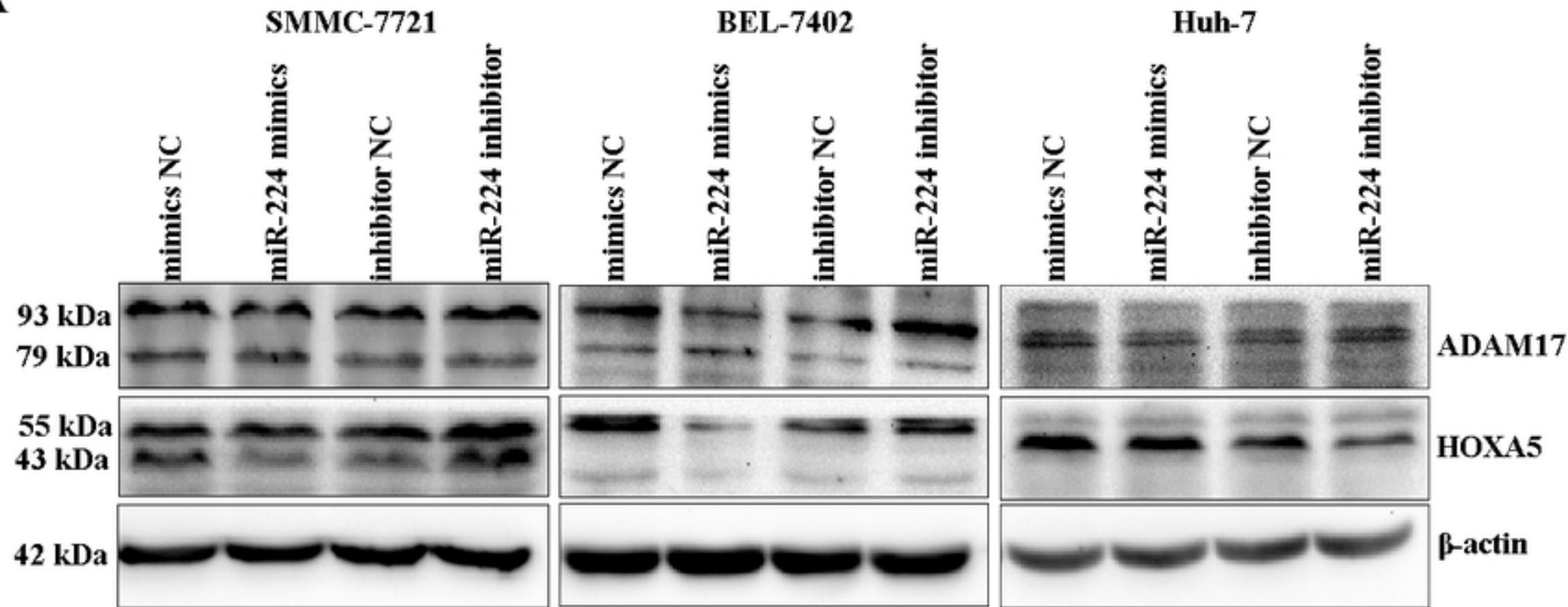
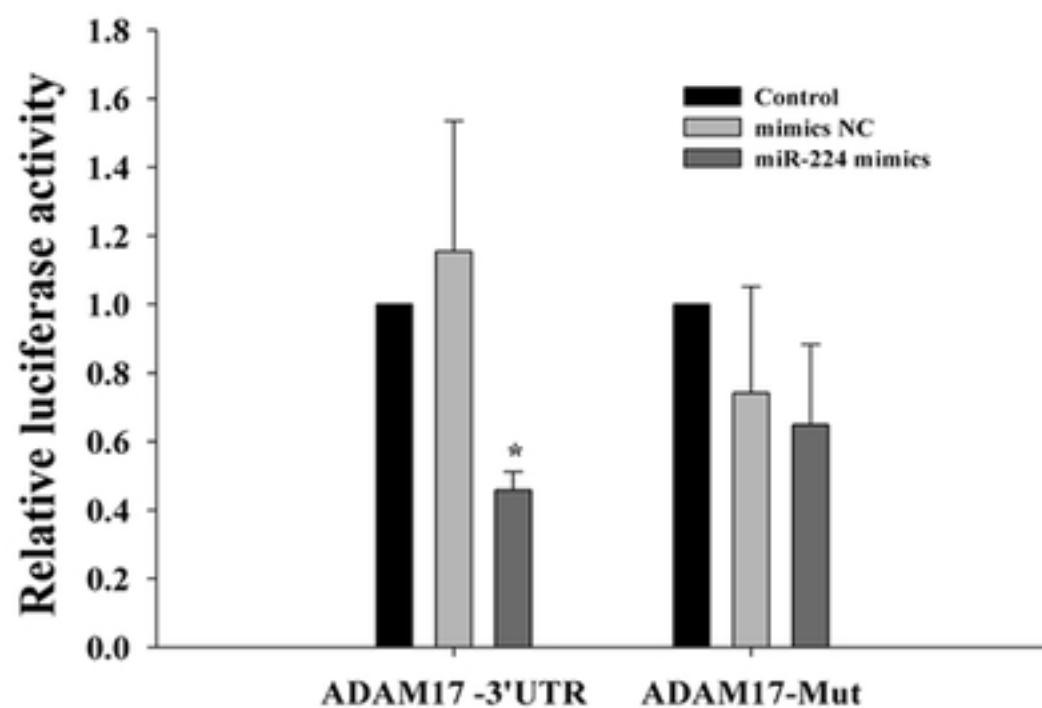
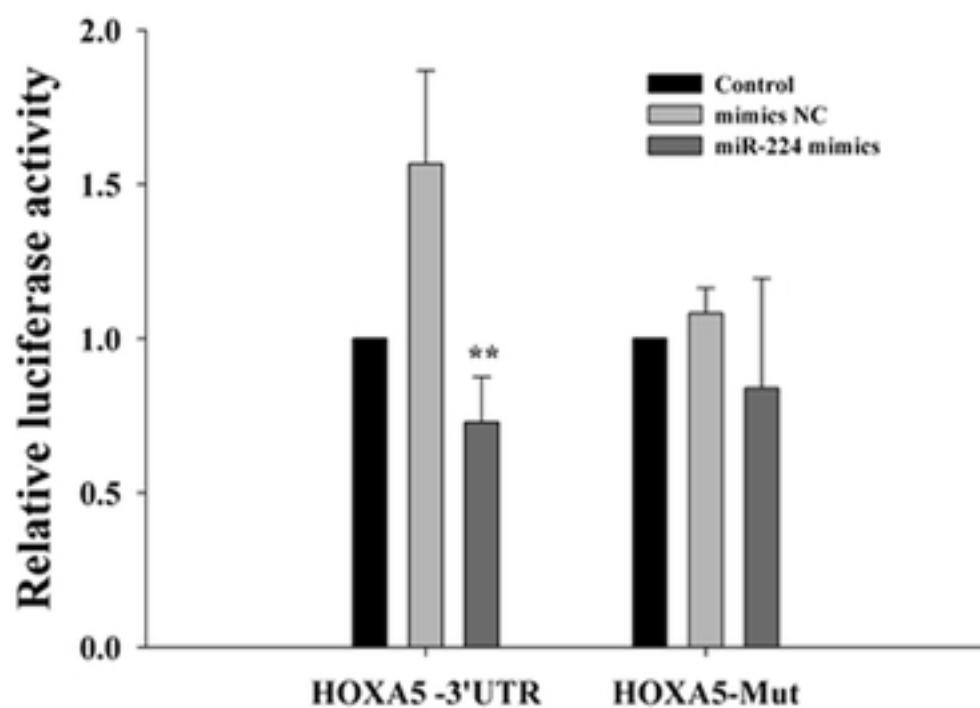
2

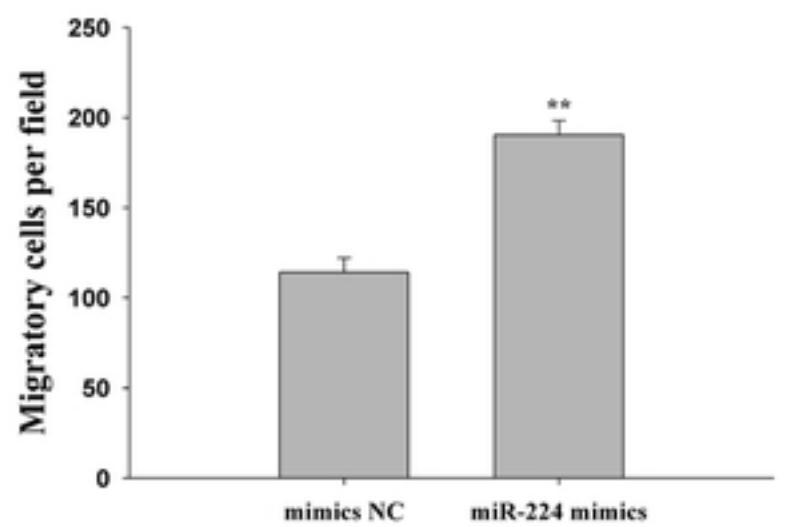
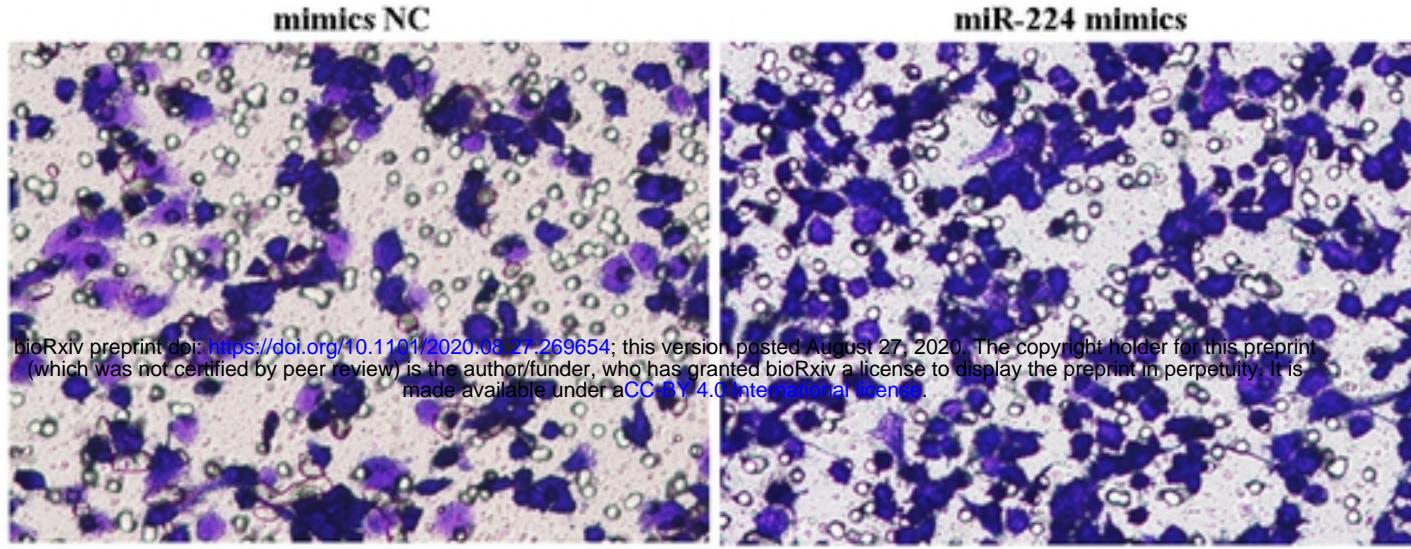
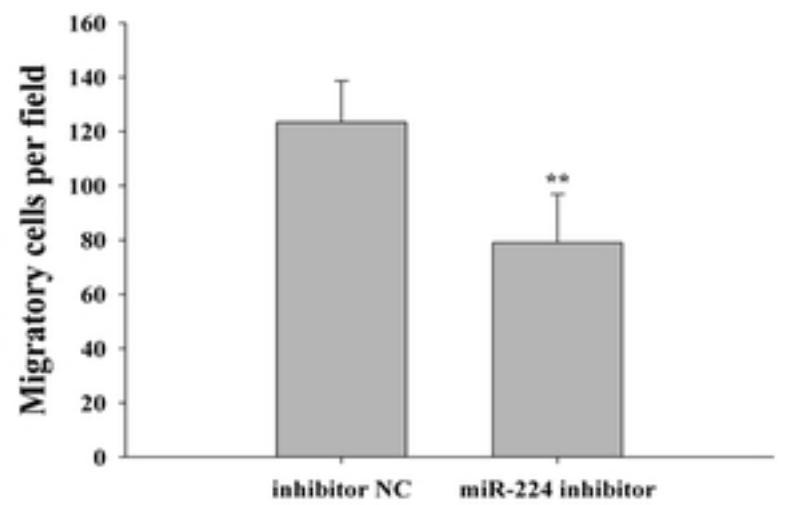
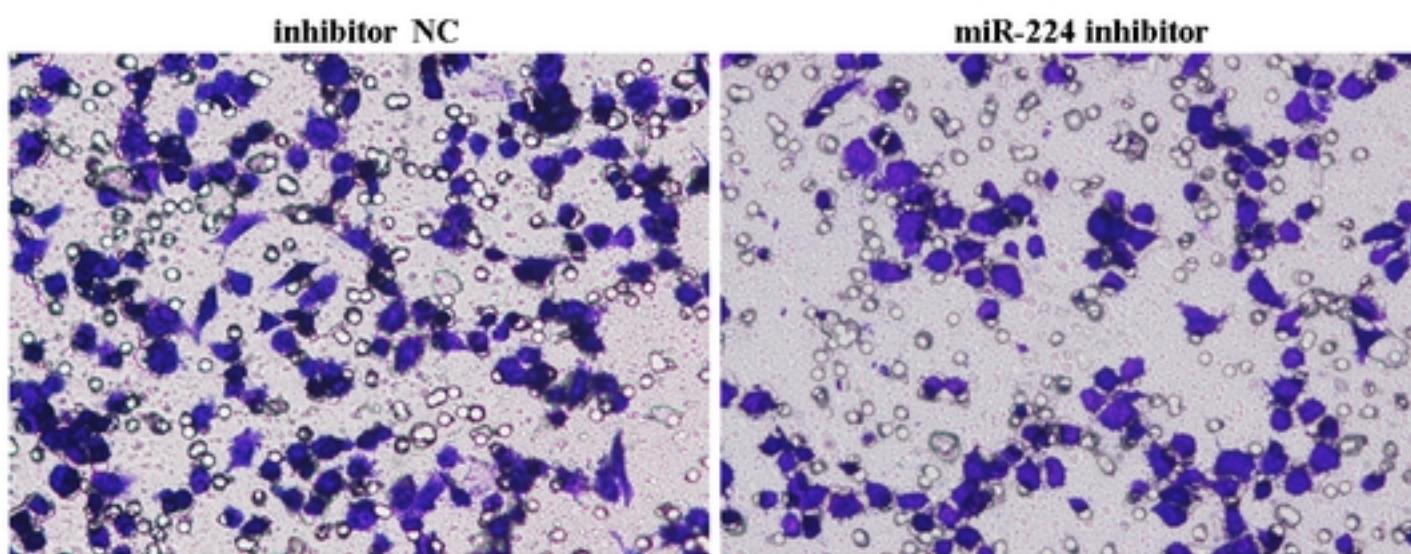
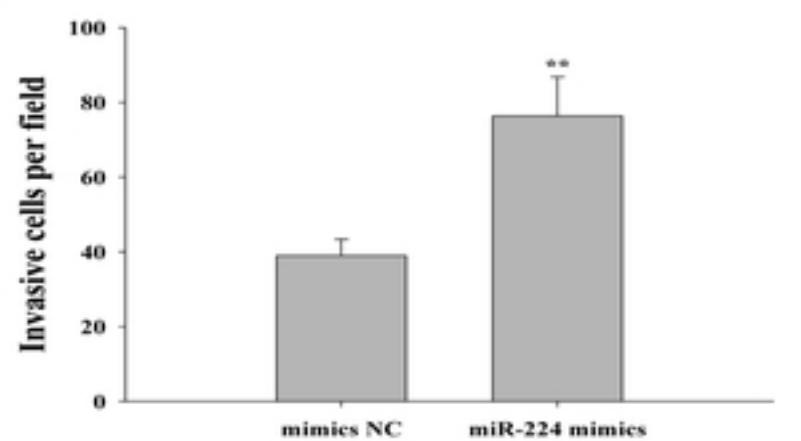
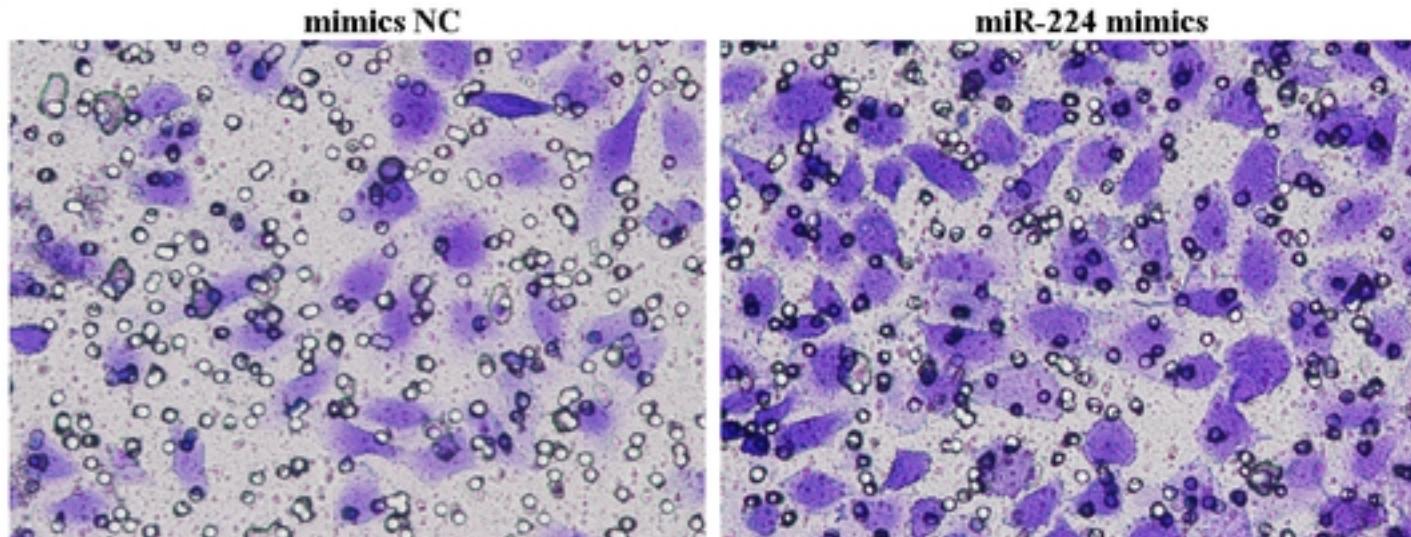
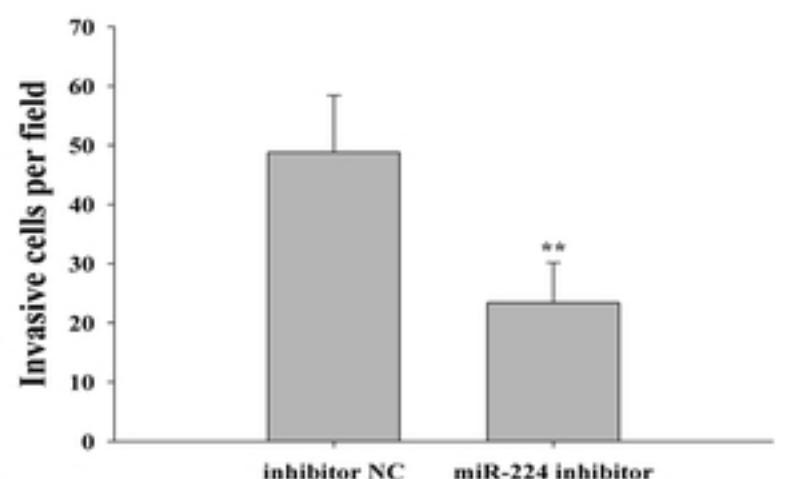
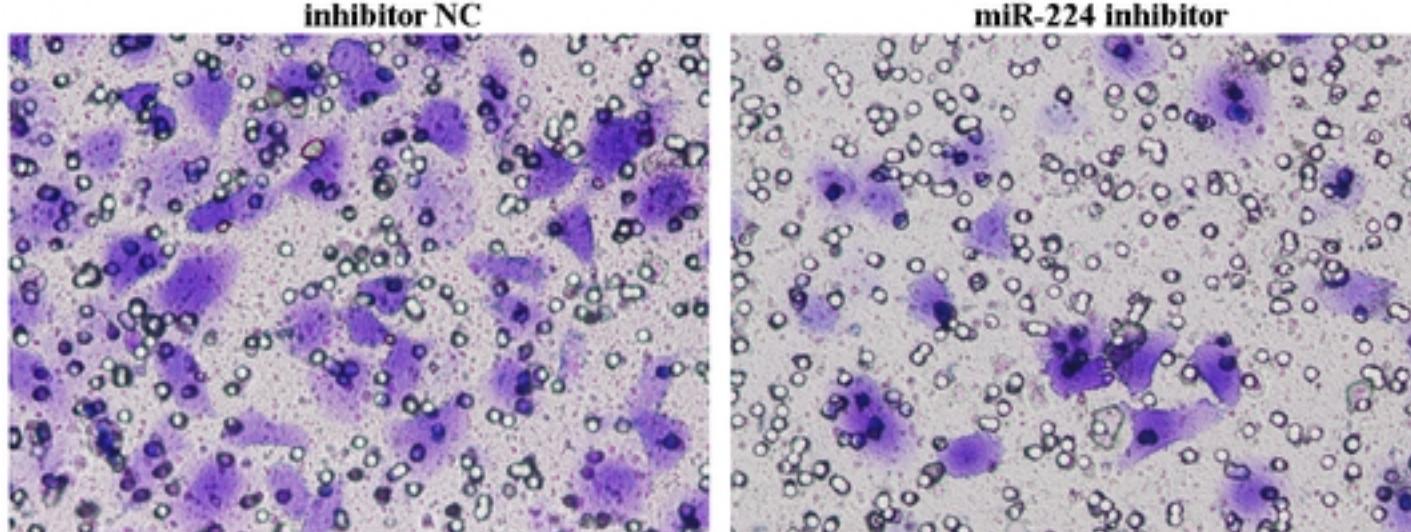
3

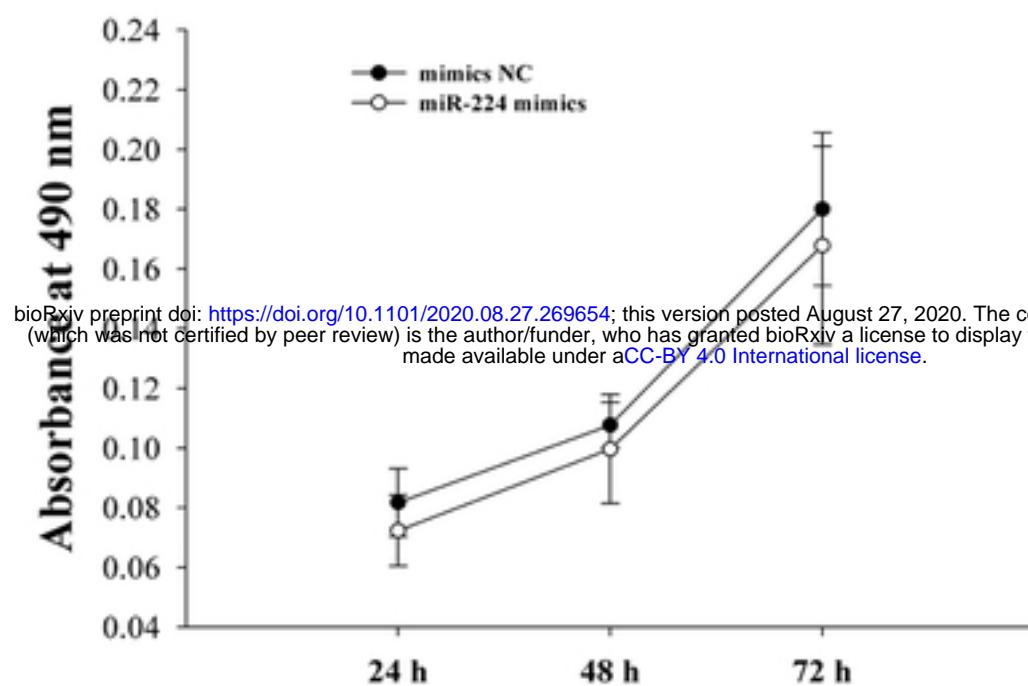
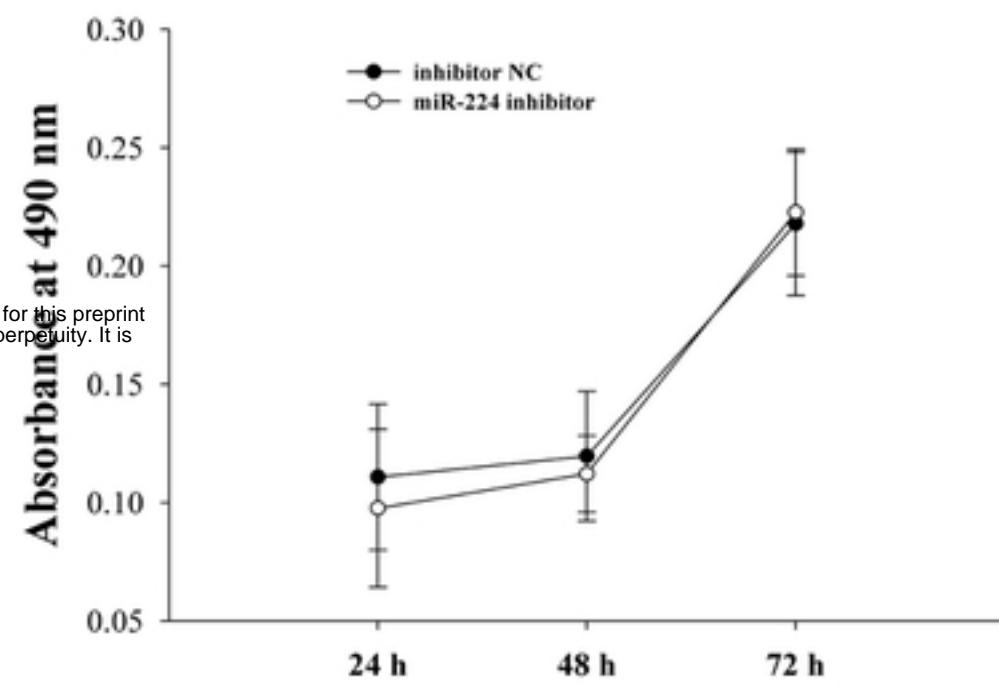
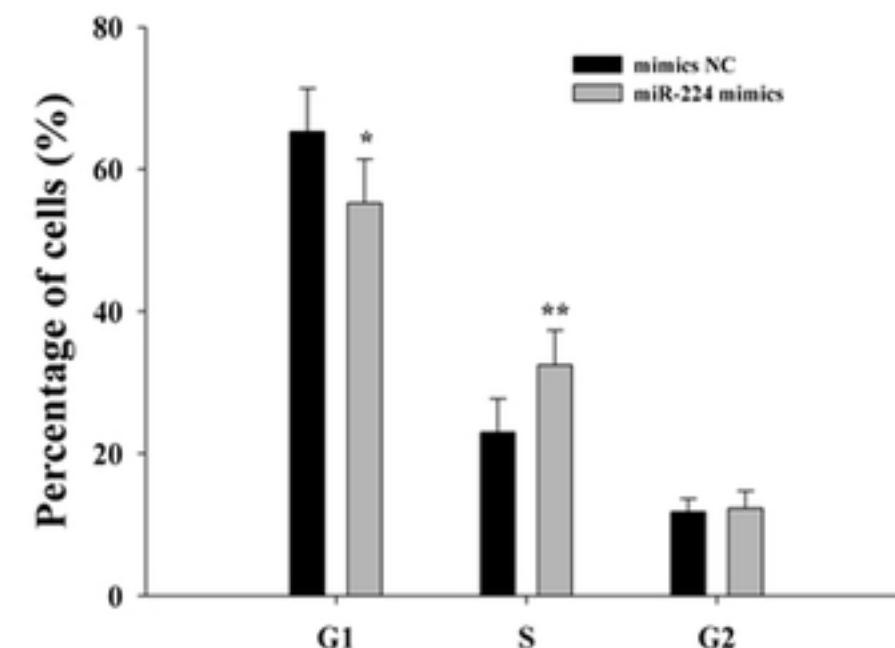
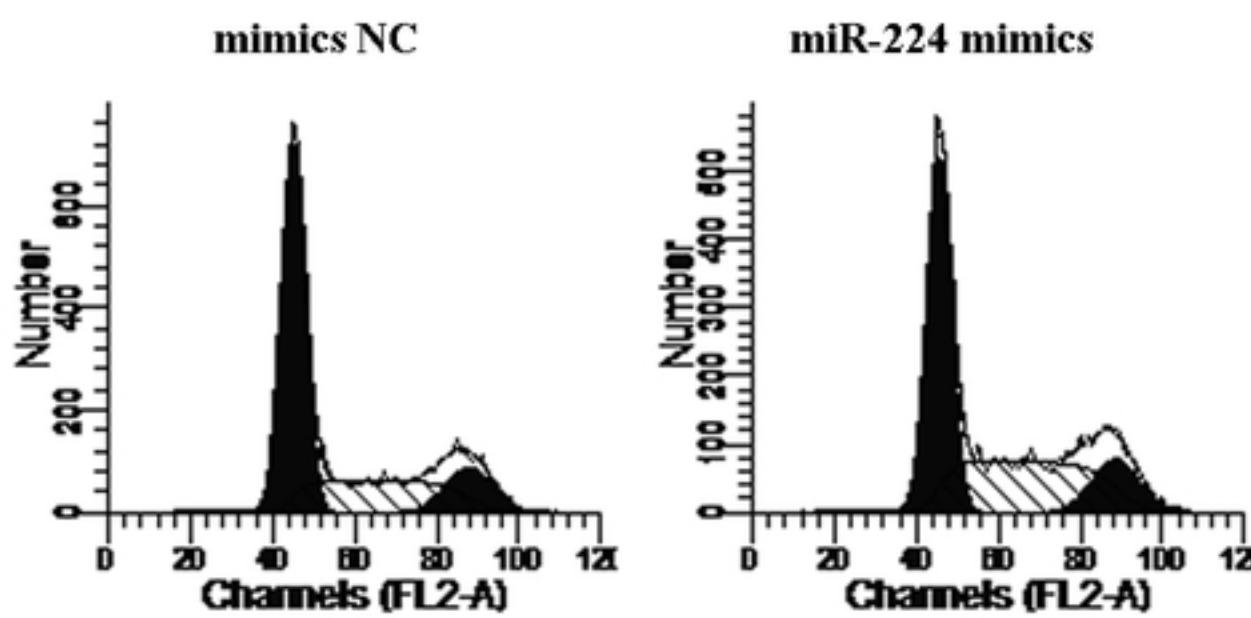
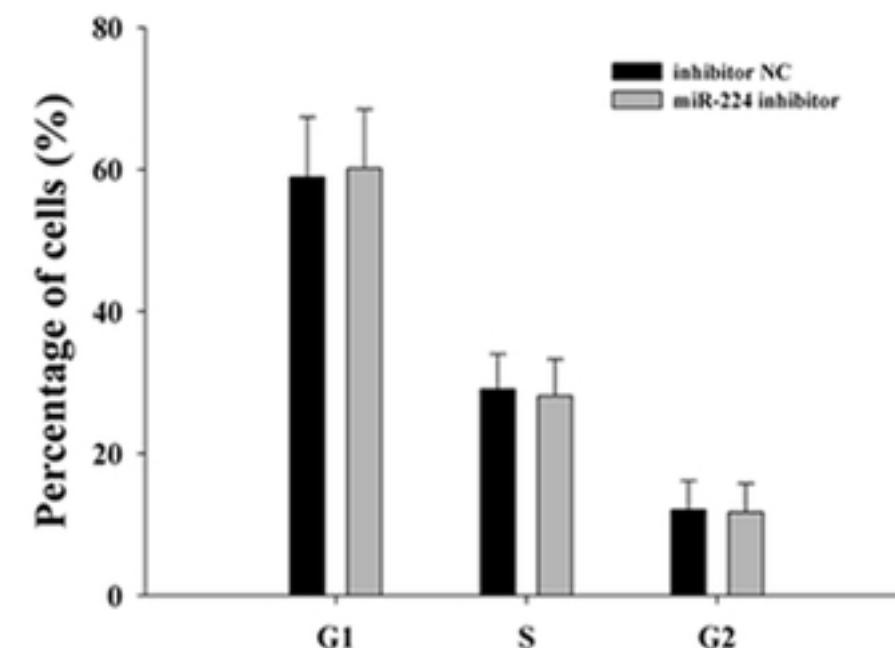
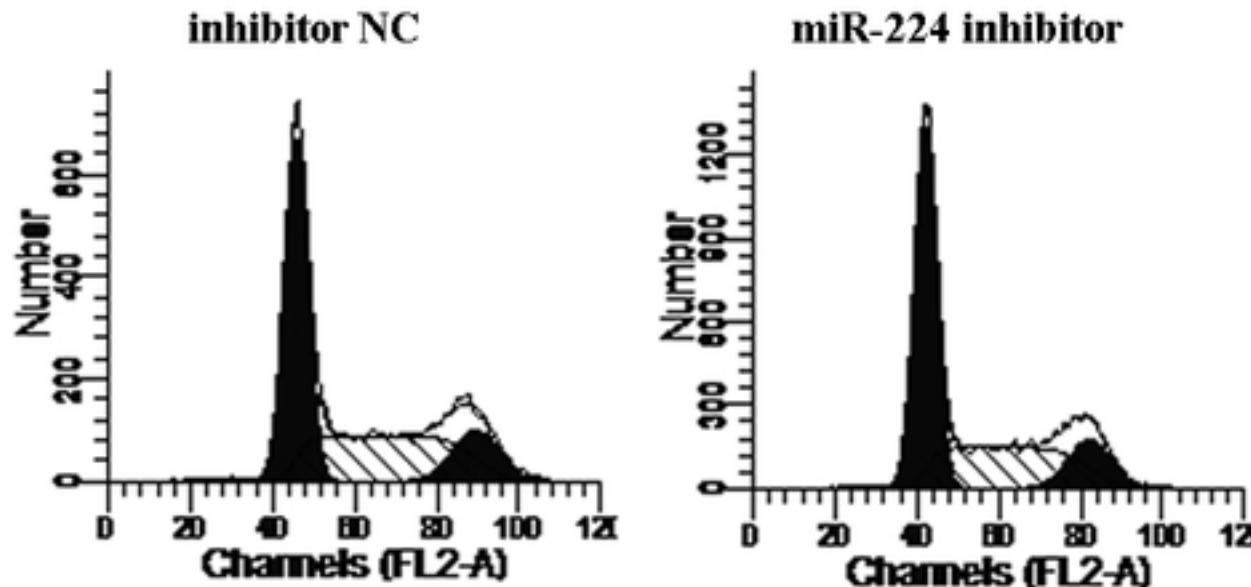
4

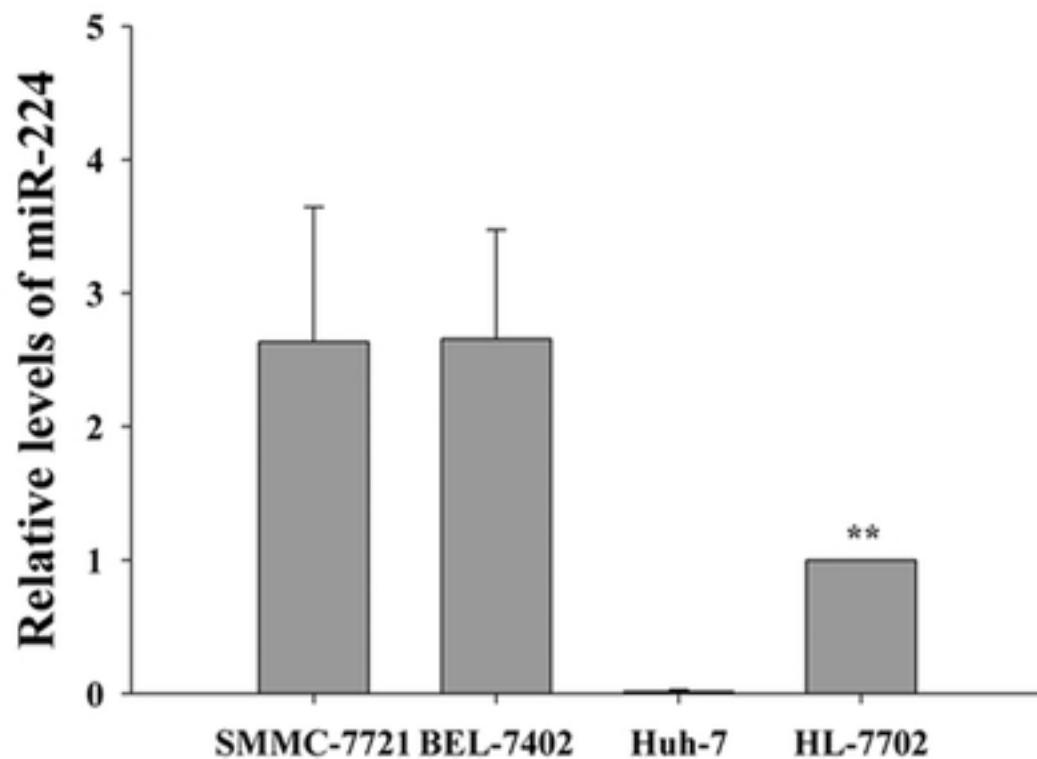
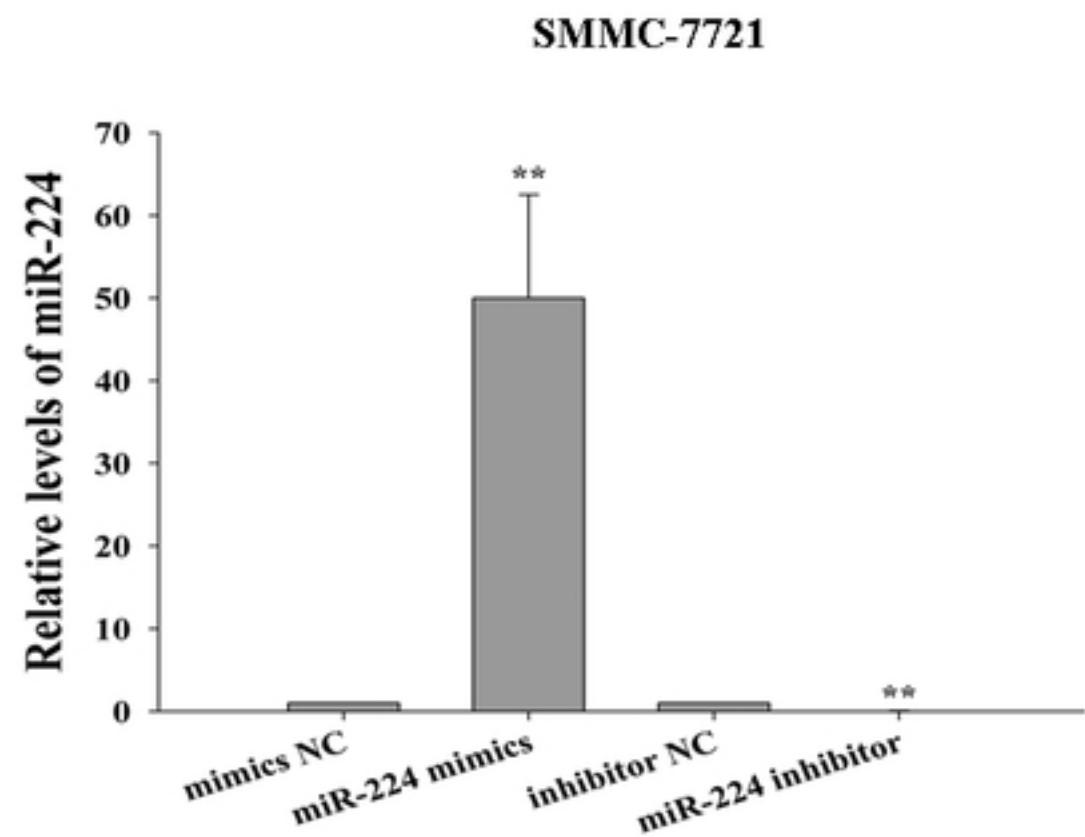
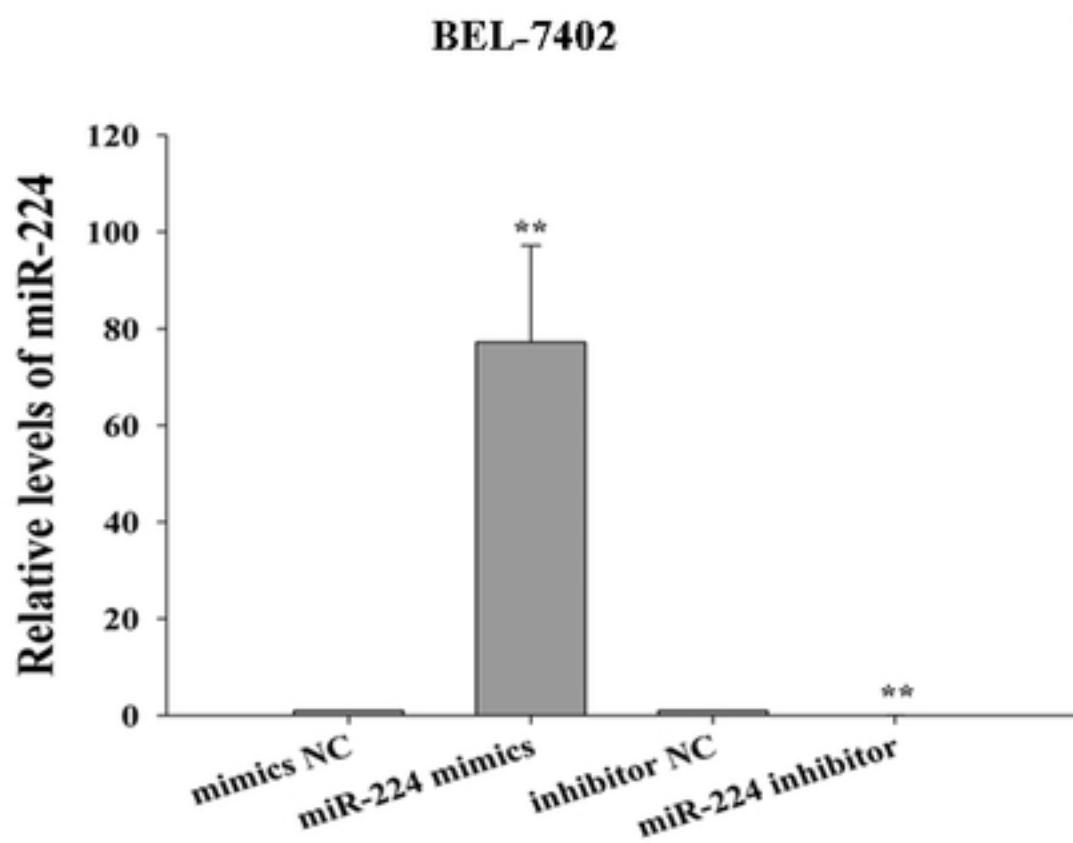
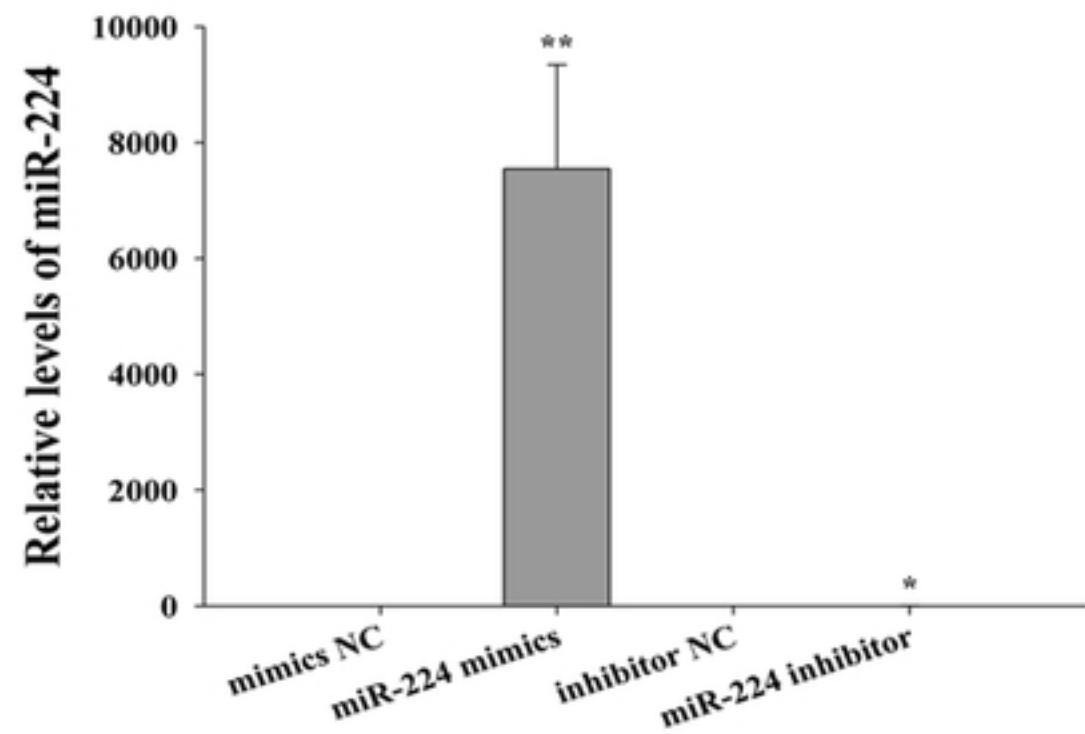
A**B****C****D****Figure**

A**B****E****C****F****D****G****Figure**

A**B****C****Figure**

A**B****C****D****Figure**

A**B****C****D****Figure**

A**B****C****D****Figure**