

Constitutive IFN α protein production in bats

Vincent Bondet^{1#}, Maxime Le Baut^{2#}, Sophie Le Poder^{2,3}, Alexis Lécu⁴, Thierry Petit⁵, Rudy Wedlarski⁶, Darragh Duffy^{1^*}, Delphine Le Roux^{2,7^*}

¹Translational Immunology Lab, Institut Pasteur, Paris, France

²BioPôle Alfort, Ecole Nationale Vétérinaire d'Alfort, Maisons-Alfort, F-94700, France

³Anses, INRAE, Ecole Nationale Vétérinaire d'Alfort, UMR VIROLOGIE, Laboratoire de Santé Animale, Maisons-Alfort, F-94700, France

⁴Parc Zoologique de Paris, Muséum National d'Histoire Naturelle, 53 Avenue de Saint-Maurice, F-75012, Paris, France.

⁵Parc Zoologique de La Palmyre, 6 Avenue de Royan, F-17570 Les Mathes, France

⁶Bioparc Zoo de Doué La Fontaine, 103 Rue de Cholet, F-49700 Doué-la-Fontaine, France

⁷Anses, INRAE, Ecole Nationale Vétérinaire d'Alfort, UMR BIPAR, Laboratoire de Santé Animale, Maisons-Alfort, F-94700, France

These authors contributed equally to this work.

^ These authors contributed equally to this work

* Corresponding authors:

Darragh Duffy, Translational Immunology Lab, Institut Pasteur, 25, rue du Dr. Roux, 75724 Paris Cedex 15, France. E-mail: darragh.duffy@pasteur.fr

Delphine Le Roux, UMR BIPAR, BioPôle Alfort, Ecole Nationale Vétérinaire d'Alfort, 7 avenue du Général de Gaulle, 94700 Maisons-Alfort, France. E-mail: delphine.le-roux@vet-alfort.fr

Abstract

Bats are the only mammals with self-powered flight and account for 20% of all extant mammalian diversity. In addition, they harbor many emerging and reemerging viruses, including multiple coronaviruses, several of which are highly pathogenic in other mammals, but cause no disease in bats. How this relationship between bats and viruses exists is not yet fully understood. Existing evidence supports a specific role for the innate immune system, in particular type I interferon (IFN) responses, a major component of antiviral immunity. Previous studies in bats have shown that components of the IFN pathway are constitutively activated at the transcriptional level. In this study, we tested the hypothesis that the type I IFN response in bats is also constitutively activated at the protein level. For this we utilized highly sensitive Single Molecule (Simoa) digital ELISA assays, previously developed for humans that we adapted to bat samples. We prospectively sampled four non-native chiroptera species from French zoos. We identified a constitutive expression of IFN α protein in the circulation of healthy bats, and concentrations that are physiologically active in humans. Expression levels differed according to the species examined, but was not associated with age, sex, or health status suggesting constitutive IFN α protein expression independent of disease. These results confirm a unique IFN response in bat species that may explain their ability to coexist with multiple viruses in the absence of pathology. These results may help to manage potential zoonotic viral reservoirs and potentially identify new anti-viral strategies.

Keywords

Chiroptera, bats, type I IFN, Simoa digital ELISA, protein levels, antiviral immunity

Abbreviations

AEB: average enzyme per bead

HAU: hemagglutinating units

IFN: interferon

LOD: limit of detection

1. Introduction

With more than 1,200 species, representing about 20% of the total diversity of Mammals, bats are among the most abundant, diverse and geographically dispersed vertebrates on the planet. They are the only mammals capable of active flight and present numerous anatomical variations (1-4). Bats also act as reservoirs for a multitude of viruses, some recognized as highly pathogenic to humans and animals (5-10). Moreover, bats have been shown to be involved in the emergence and re-emergence of numerous highly pathogenic zoonotic viruses such as Rhabdoviridae, Paramyxoviridae (Nipah and Hendra viruses), Filoviridae (Ebola and Marburg viruses), and Coronaviridae (11-19) for which they are suspected to be involved in the emergence of the original SARS-CoV-2 viral strain (20). Moreover, most of the bats experimentally infected with viral doses of Hepinaviruses or Lyssaviruses, which are lethal to other mammals, did not show apparent clinical signs. (21-23). It is likely that viruses and their bat hosts have undergone a long process of co-evolution that began several million years ago with the appearance of the first Chiroptera (9, 24, 25). These mechanisms, along with their specific characteristics including longevity, migratory activity, active flight, and population density, may have shaped both the bat immune system and their ability to thwart host responses to viruses, resulting in a balance between persistent infection and absence of pathophysiology (5, 6, 10, 26, 27). One hypothesis is that bats are able to control viral replication through the existence of specific innate antiviral mechanisms (28). Among them, the production of IFN is known to be the first line of defence against viral infections (29) and there is evidence of a strong constitutive genomic expression of type I IFN, mainly of IFN α , in at least two species of Chiroptera (*P. alecto* and *C. brachyotis*) (30). This difference is unique since it has not been observed in other mammals, however, it remains to be confirmed whether this constitutive expression occurs at the protein level.

We have previously used ultra-sensitive Single Molecule Array (Simoa) digital ELISA to measure IFN α protein in the serum of human patients with autoimmune diseases or viral infection whose

levels were previously undetectable with conventional ELISA techniques (31). This ultra-sensitive technique is therefore capable of measuring cytokines at very low concentrations in biological fluids, which were previously only measured indirectly by detection of downstream gene induction. Thus, the measurement of IFN α protein in Chiroptera may confirm the hypothesis raised by Zhou *et al.* which is based solely on mRNA measurements and not on direct protein quantification (30). Indeed, constitutively expressed type I IFN mRNA in bats could represent a "ready to use" pool during viral infection, or it could be directly translated into protein resulting in high blood concentrations and viral protection. In this study, we quantified bat IFN α 2 protein using ultra-sensitive digital ELISA on plasma from four species of captive bats: *P. rodricensis*, *P. lylei*, *R. aegyptiacus* and *E. helvum*. We also correlated the IFN α protein expression to the corresponding mRNA levels. This study provides new evidence of how the unique immune system of bats may control viruses in the absence of disease and in doing so constitute a constant viral reservoir for zoonotic transmission and potential new pandemics.

2. Materials and Methods

2.1 Bat cohort and sampling

Four bat species from four French zoos were sampled, during their annual sanitary examination, by the resident veterinary doctor. 0.2 to 0.5 mL of blood was drawn from different veins depending on the species and collected in an EDTA containing tube using a 1mL syringe and a 25G x 5/8 needle (all from Beckton Dickinson, France). Under general anesthesia (O₂ at 1.5L/min and 5% isoflurane for induction and 2% to maintain the anesthesia) blood of *P. rodricensis* and *R. aegyptiacus* was taken from the medial vein and jugular vein, respectively. *E. helvum* and *P. lylei* were vigilant during blood sampling, which was done from the medial vein for both species. The demographic characteristics and the origins of this cohort are indicated in Table 1. Blood was then split into PAXGene tubes (PreAnalytix GmbH, Qiagen, France) for RNA extraction and RT-qPCR analysis,

and Eppendorf tubes to obtain plasma. PAXGene tubes were stored at -20°C until extraction, while Eppendorf tubes were centrifuged at 2500rpm for 10min. Plasma was then removed and stored at -80°C until Simoa analysis.

Number of individuals	Sex, (female)	Species	Origin
108	36 (43%)	<i>Pteropus rodricensis</i> : 21 (19%)	Parc Zoologique de La Palmyre
		<i>Rousettus aegyptiacus</i> : 10 (9.3%)	Parc Zoologique de La Palmyre
		<i>Eidolon helvum</i> : 23 (21%)	Parc Zoologique de Paris
		<i>Pteropus lylei</i> : 54 (50%)	Bioparc Zoo de Doué La Fontaine

Table 1. Bat species sampled from 4 different zoos.

2.2 Bat cell stimulation assays

Bat epithelial cells from Tb 1 Lu cell line (ATCC, United States) were cultured at 37°C, 5% CO₂, in complete culture medium composed of MEM Eagle medium with 2 mM Glutamine supplemented with 50 U/mL penicillin, 50 µg/mL streptomycin (all from Lonza, Belgium) and with 10% of decomplemented fetal calf serum (Gibco, Thermo Fisher Scientific, France). Before stimulation, cells were plated in 1mL of complete medium per well of 24 well plates and maintained at 37°C, 5% CO₂ until they reached 2x10⁶ cells/well. Supernatants were removed and 1mL of complete medium including 500HAU/mL mouse influenza virus (Strain H1N1 A/PR8/1934) was added to the cells or not (unstimulated control). Before stimulation and 1 hour, 3.5 hours, and 23 hours after stimulation, supernatants were sampled and frozen at -80°C for IFNα protein quantification.

2.3 Production of the *Rousettus aegyptiacus* IFNα protein for ELISA calibration

The *Rousettus aegyptiacus* IFNα DNA sequence was obtained from a previously published study (32). Nucleotide bases that correspond to the signal peptide were removed, a start codon, spacers, and codons for a 6His tag and a TEV cleavage site were added in the 5' termination. The cDNA

coding for the recombinant protein was chemically-synthesized with optimization for expression in *Escherichia coli*. The recombinant gene was then introduced in a pT7 expression plasmid under the control of a Lac operator and harboring kanamycin resistance. *E. coli* strains were transformed and kanamycin-resistant clones were selected. After optimization, protein production was done, culturing the selected clones in a Luria–Bertani kanamycin (LBkan) medium at 16°C during 16 hours after induction with 1mM isopropyl β-D-1-thiogalactopyranoside (IPTG). Bacteria were then harvested by centrifugation. The pellet was lysed and the soluble extract was obtained after a second centrifugation. This soluble extract was directly used at different dilutions as a calibrator for the digital ELISA assay. It was aliquoted and stored at -80°C before use.

2.4 Sample preparation for IFN α 2 digital ELISA assay

All plasma samples were first thawed and centrifuged at 10.000g, +4°C for 10 minutes to remove debris. Because bats can harbor many viruses, supernatants were treated in a P2 laboratory for viral inactivation using a standard solvent/detergent protocol used for human blood plasma products (33, 34) and described in (35) and in (36). Briefly, samples were treated with Tri-*n*-Butyl Phosphate (TnBP) 0.3% (v/v) and Triton X100 (TX100) 1% (v/v) for 2 hours at room temperature. After treatment, TnBP was removed by passing the samples through a C18 column (Discovery DSC-18 SPE from Supelco). For digital ELISA assays, inactivated samples and stimulated cell supernatants were diluted in the Detector / Sample Diluent (Quanterix) added with NP40 0,5% (v/v). They were then incubated for one hour at room temperature before analysis. Global dilution factor was generally 1/6 for plasma samples and 1/3 for stimulated cell supernatants depending on the amount of material available and to allow the optimal protein detection.

2.5 IFN α 2 digital ELISA assay

The Simoa IFN α 2 assay was developed using the Quanterix Homebrew kit and described in (36). The BMS216C (eBioscience) antibody clone was used as a capture antibody after coating on

paramagnetic beads (0.3mg/mL), and the BMS216BK biotinylated antibody clone was used as the detector at a concentration of 0.3ug/mL. The SBG revelation enzyme concentration was 150pM. The assay follows a 2-step ELISA configuration. Two calibrators were used; recombinant human IFN α 2c (hIFN α 2c) purchased from eBioscience and *Rousettus aegyptiacus* IFN α (bIFN α) produced in *Escherichia coli* for this study. The limit of detection (LOD) was calculated by the mean value of all blank runs + 2SD after log conversion.

2.6 RNA extraction and IFN α RT-qPCR

Whole blood RNA was extracted manually from PAXGene tubes, following manufacturer's instructions (Blood RNA extraction kit, Qiagen, France). After extraction, samples were inactivated at 65°C for 5min then stored at -80°C until RT-qPCR. RT-qPCR was done using the qScript XLT One-Step RT-qPCR mix following manufacturer's instructions (Quanta BioSciences, Inc., United States). Taqman probes (Applied Biosystems, ThermoFisher Scientific, France) and primers (Eurofins, France) for IFN- α 1, IFN α 2 and IFN α 3 were described previously for *P. alecto* bat species in (30) and used here. Probes and primers for the bat GAPDH housekeeping gene were designed using Primer-BLAST from NCBI (<https://www.ncbi.nlm.nih.gov/tools/primer-blast/>) and are presented in table S2. All data were normalized relative to the housekeeping gene (GAPDH) as indicated. The expression level of the target genes was calculated using the standard curve method and expressed as copy numbers relative to the housekeeping gene.

2.7 Nested RT-qPCR for pan-coronaviruses in mRNA from bat whole blood

RT-qPCR for potential coronaviruses in bat whole blood RNA was performed from the mRNA extracted previously and following the protocol previously published in (37).

2.8 Statistical analyses

GraphPad Prism 8 was used for statistical analysis. Mann-Whitney tests were used to compare two

groups such as female and male, or healthy and disease. ANOVA tests (Kruskal–Wallis) with Dunn's post testing for multiple comparisons were used to test for differences between multiple bat species. For all analyses, p values less than 0.05 were considered statistically significant, with *p<0.05; **p<0.01; ***p<0.001; ****p<0.0001. Median values were reported on figures. Spearman correlations are used to compare continuous variables such as mRNA level or age and protein production.

2.9 Data availability

All available data from the bat cohort are shown in Supplementary Table S1.

3. Results

3.1 Bat IFN α protein detection with an IFN α 2 digital ELISA assay

Anti-bat IFN α antibodies are not commercially available for the development of bat-specific IFN α ELISA. However, given the ultra-sensitivity of human IFN α digital ELISA which detects protein at attomolar concentrations (31), and potential cross-species reactivity, we hypothesized that our existing human assay could also detect bat IFN α . As a first proof of concept, we stimulated a bat lung epithelial cell line with influenza virus (Strain H1NI A/PR81934) and tested the recovered supernatant with a human IFN α 2 digital ELISA. We observed a significant induction of IFN α 2 protein at 1hr and 3.5 hrs as compared to the unstimulated control (Fig 1a).

These initial results were extrapolated from a standard curve of a human recombinant IFN α 2 protein. To better adapt our assay to bat species, we produced a recombinant *Rousettus aegyptiacus* IFN α protein (bIFN α) from *Escherichia coli* competent bacteria. SDS-Page analysis of the soluble and insoluble extracts obtained from the bacteria pellet showed that bIFN α was mainly produced as an insoluble form even at low induction temperature (Fig S1a). Comparing profiles before and after induction of the protein expression, SDS-Page analysis showed that the unique

bIFN α band appeared alone at this mass (Fig S1a). Western-Blot analysis of the soluble fraction after induction at 16°C using the IFN α 2 assay detection antibody revealed that the protein was expressed in a single band at the expected molecular weight (Fig S1a). The purification from the soluble extract failed: the bIFN α protein was not selected at the expected molecular weight (Fig S1b) and the western-Blot analysis revealed no affinity at the purified molecular weight (Fig S1a). The purification from the insoluble extract succeeded, but the renaturation of the protein failed (Fig S1c). So we used the soluble extract itself as a calibrator after quantification of bIFN α . Global protein quantification of the soluble extract was done using the BCA assay, and the bIFN α protein concentration in the soluble extract was estimated after gel densitometry for potential use as a digital ELISA calibrator.

To explore the ability of the IFN α 2 digital ELISA assay designed for the quantification of human interferons, to quantify bat IFN α species, we compared the responses of the assay to bIFN α protein and all 13 human IFN α subtypes (Fig. 1b). As expected the assay revealed a weaker response for bIFN α in comparison with hIFN α 2c. However, the affinity of the human mAb for the bat protein was comparable to the human subtypes, with bIFN α and hIFN α 21 showing very similar affinities, and two human species showing weaker responses (Fig. 1b). Using the bIFN α protein as the calibrator, we re-calculated the cellular response after *in vitro* influenza stimulation and observed similar results with the highest concentrations present after 1hr of influenza stimulation (Fig 1c). The only difference observed was related to the scale of these results, due to the lower affinity of the mAb for the bIFN α 2 calibrator.

To better understand the cross-species reactivity we compared the sensitivities of the 13 IFN α subtypes as previously described (36), the 5 other human IFN β , IFN λ 1, IFN λ 2, IFN ω and IFN γ , and the 5 mouse IFN α 1, IFN α 3, IFN α 4, IFN α 11 and IFN α 13, with their available online sequences in the UniProtKB database (www.uniprot.org) after alignment using the CLUSTALW software (www.genome.jp/tools-bin/clustalw). We also considered the fact that epitopes must be accessible to the antibodies and so studied the IFN α 2, IFN α 14 and IFN ω three-dimensional

structures published online in the PDB database (www.rcsb.org) after spacial alignment using the PyMOL software (www.pymol.org). This analysis suggests that the epitope recognized by the IFN α 2 mAb could be the ¹¹⁰LMKED sequence in the human IFN α 2 molecule. *Rousettus aegyptiacus* IFN α and *Pteropus rodricensis* IFN α 1, IFN α 2 and IFN α 3 protein sequences were previously described by Omatsu *et al* (32) and Zhou *et al.* (30) respectively. Alignment of these molecules showed that the corresponding amino-acids in the IFN α 2 assay epitope position are LMNED for the 3 IFN α species from *Pteropus* and LLDED for *Rousettus* (Fig S2a). The LMNED sequence also appears in human IFN α 16 and IFN α 17, two species for which the IFN α 2 assay shows a positive response. The M→L substitution concerns two apolar amino-acids. The K→D substitution changes a positive with a negative charged amino-acid, but these two amino-acids are then hydrophilic and so do not produce a detrimental α -helix coil in the structure. This *in silico* analysis provides support for how the IFN α 2 antibody assay may recognize *Rousettus* and *Pteropus* IFN α protein.

3.2 IFN α proteins are constitutively elevated in plasma of bat species

Having validated the assay for its ability to detect bat IFN α , we analyzed plasma samples from 4 bat species sampled from French zoos (Table 1) with the digital ELISA assay. Results are presented using the two calibrators; hIFN α 2c (Fig. 2a) and bIFN α (Fig. 2b). A greater number of samples were above the assay limit of detection using the bat protein calibrator as compared to the human protein, confirming the interest of using a bat specific protein. IFN α protein responses obtained within species were relatively consistent. *Pteropus rodricensis* and *Rousettus aegyptiacus* showed significantly elevated IFN α protein plasma levels as compared to *Eidolon helvum* and *Pteropus lylei*, with both the hIFN α 2c ($p<0.05$) and bIFN α ($p<0.0001$) calibrators. Notably in *Eidolon helvum* plasma IFN α was largely undetectable with both assays revealing interesting inter-species differences. When available, we assessed whether age, sex, or presence of mild clinical symptoms

(Table S1) were associated with IFN α protein levels in all species but found no significant associations (Fig S3). We also tested for presence of corona viruses in the blood but found no evidence (data not shown). These results support the hypothesis that certain bat species have physiological levels of circulating IFN α protein in healthy conditions.

3.3 IFN α mRNA are constitutively expressed in bat leukocytes

The constitutive mRNA expression of bat IFN α genes has been previously described for *Rousettus aegyptiacus* (32) and *Pteropus rodricensis* (30). To test whether the protein plasma levels we observed were linked with leukocyte mRNA expression, and to extend these observations to *Eidolon helvum* and *Pteropus lylei*, we quantified gene expression of IFN α 1, IFN α 2 and IFN α 3 in our cohort using RT-qPCR and normalizing the results using GAPDH mRNA. Results from *Rousettus aegyptiacus* were negative, perhaps explained by a lack of specificity of the primers utilized. In the other bat species examined the number of IFN α mRNA copies were globally similar to GAPDH, supporting that IFN α species are constitutively expressed at the mRNA level. The IFN α 2 subtype mRNA was the most expressed, while IFN α 3 was the most variable between species. *Eidolon helvum* had the lowest IFN α mRNA levels, reflecting the absence of detectable protein in this species. *Pteropus lylei* had overall the highest IFN α RNA levels. Significant differences ($p<0.05$) were observed between *Eidolon helvum* and *Pteropus lylei* for IFN α 2 and IFN α 3, and between *Pteropus rodricensis* and *Pteropus lylei* for IFN α 3. This also indicates that high levels of variation could be observed within a same genus, and also within the same species.

3.4 IFN α mRNA expression and protein production could be linked within each species

While *Eidolon helvum* showed lower mRNA and protein levels as compared to the other species, and *Pteropus rodricensis* medium mRNA levels and a high protein level, *Pteropus lylei* showed the highest RNA and the lowest protein levels (Fig. 2a, b and Fig. 3a). Therefore, we observed no overall direct link between IFN α mRNA expression and protein plasma concentrations across all

species examined. To directly explore this hypothesis, we tested the correlations for each gene and protein for *Pteropus rodricensis* samples where sufficient detectable measurements were available for both parameters. In this relatively small sample size (n=8 for paired samples) we observed a strong statistically significant correlation ($Rs=0.952$, $p=0.001$) between IFN α 2 RNA and protein levels (Fig. 3b). No correlation was observed with IFN α 1 and IFN α 3 genes (Fig. 3b). This result may reflect that the ELISA assay is designed for the IFN α 2 subtype.

4. Discussion

Type I interferons trigger the downstream activation of hundreds of critical genes as part of the anti-viral immune response. Because of this potency, IFN proteins are secreted at relatively low concentrations as compared to other major cytokines, and multiple regulatory mechanisms exist to control their effects (38). Potential negative consequences of over activation of interferon responses is reflected by their direct implication in multiple autoimmune conditions such as lupus and interferonopathies (39). Furthermore, long-term treatment of patients (ie chronic HCV patients) with type I IFN based therapies can induce serious side effects including depression (40).

Because of these potential negative effects of IFN signaling, recent studies that reported constitutive IFN gene expression in healthy bats were unexpected. However, it provided strong evidence for how bats may potentially live healthily with multiple viral species that are pathological to other mammals including humans. Nevertheless, it raised many additional questions, namely the one which we addressed in our study as to whether IFN protein is also constitutively elevated in bats. This question is not so trivial to address for multiple reasons. IFN proteins have been challenging to directly quantify due to their low physiological concentrations in biological samples, and most studies have utilized proxy readouts such as interferon stimulated gene (ISG) expression or cytopathic assays. The development of digital ELISA such as Simoa overcame these challenges as we demonstrated by the measurement of all 13 IFN α human subtypes, and more recently specifically IFN α 2 in multiple human cohorts (31, 36, 41). However additional challenges exist for

the study of interferon in non-human species, in particular bats, most notably the lack of specific reagents in particular monoclonal antibodies which are required for ELISA technologies. We tested the hypothesis that attomolar digital ELISA sensitivity combined with species cross-reactivity would enable the quantification of bat IFN α protein. After assay validation on virus stimulated bat cell cultures, we further modified the assay with a recombinant bat protein as the standard calibrator. Applying this assay to 4 species of bats conclusively showed constitutive expression of IFN α protein in the circulation of healthy animals. While we cannot yet make a direct comparison, these levels are significantly higher as compared to healthy humans, where IFN α is essentially undetectable (36) even at attomolar sensitivity.

Our study contains some inherent weaknesses. IFN α protein concentrations were calculated using human IFN α 2c or *Rousettus aegyptiacus* IFN α protein (bIFN α) produced in *E. coli* as calibrators, with results respectively in the fg/mL and the ng/mL ranges. Such a difference could be due to different antibody specificities, potentially due to incorrect folding of the bIFN α protein produced in *E. coli* strains. This is supported by the observation that the bIFN α protein is mainly expressed in the insoluble fraction even at 16°C (Fig. S1c), that the protein failed to renature after urea purification (Fig. S1b), and that the viral inactivation solvent/detergent protocol had a greater effect on the IFN α 2 assay response to bIFN α (>1Log) than hIFN α 2c (Fig. S4), suggesting greater insolubility of the bat protein. Additional improvements of the assay could be envisioned such as production of a purer and better folded recombinant protein in mammalian cells, and eventually the production of a bat specific monoclonal antibody against this protein.

Despite these technical limitations, we were able to show elevated levels of plasma IFN α protein in certain bat species, which also correlated with expression levels of the IFN α 2 gene. While additional confirmatory experiments will be required, the inter-species differences in plasma IFN α protein is an interesting observation. It would also be interesting in future studies to assess whether these IFN α protein differences have an impact on viral levels and diversity within the different bat species. Lastly, our results raise additional new questions on the nature of bat

physiology, in particular how the constitutively activated type I IFN response is maintained in bats without resulting in pathological conditions such as those observed in human autoimmune disease. Chronic IFN activation, in particular during growing and development phases, can have significant neurological effects as observed in interferonopathies such as STING mutation patients (42). Finally given the important role of type I interferon for protection to infection with SARS-CoV-2 (35, 43), and the potential role bats have played in seeding the COVID-19 pandemic (20) understanding this host-virus relationship could have major implications for pandemic preparedness. In summary improved knowledge on the special nature of bat IFN regulation could have major implications for our basic understanding of IFN biology, its continued use as a therapeutic, and our capacity to prepare for viral pandemics.

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Figures Legends

Figure 1. Bat IFN α proteins are detected with a human IFN α 2 digital ELISA assay.

(a) IFN α protein levels expressed as equivalent human IFN α 2c concentrations obtained after stimulation of bat epithelial cells Tb 1 Lu with 500HAU/mL mouse influenza virus (FLU) or unstimulated (NS) for 0 to 24 hours at 37°C and 5% CO₂. **(b)** IFN α 2 digital ELISA assay response (AEB) as a function of the IFN α concentration for the *Rousettus aegyptiacus* IFN α calibrator (bIFN α) produced in *Escherichia coli* (red) in comparison with the human IFN α 2c calibrator (blue) and the 12 other human IFN α subtypes. **(c)** IFN α protein levels expressed as equivalent bIFN α concentrations obtained after stimulation of bat epithelial cells as described previously. Box plots represent median and individual values represented by dots are reported on figures, a & c representing pooled results from 3 independent experiments

Figure 2. IFN α protein concentrations in plasma of four bat species.

IFN α concentrations measured in plasma from four bat species expressed (a) as hIFN α 2c or (b) as bIFN α equivalent concentrations. LOD is the limit of detection level of the assay. Median represented by black line with individual animals shown by colour coded dots. Kruskal–Wallis test with Dunn's post testing for multiple comparisons was used, *p<0.05, ***p<0.0001, ****p<0.001. *P. rodricensis* (green, n=12) *R. aegyptiacus* (light blue, n=10), *E. helvum* (dark blue, n=21) and *P. lylei* (purple, n=52).

Figure 3. IFN α mRNA levels in bats and mRNA-protein correlations

(a) Number of copies measured in whole blood for IFN α 1, IFN α 2 and IFN α 3 mRNAs as normalized to GAPDH in three bat species. Probes used here were not suitable for *Rousettus aegyptiacus*. Median represented by black line with individual animals shown by colour coded dots. Kruskal–Wallis test with Dunn's post testing for multiple comparisons was used. *p<0.05. (b) Correlation plots between the Eq. bIFN α protein concentration obtained using the IFN α 2 digital ELISA assay and the GAPDH-normalized number of IFN α 1, IFN α 2 and IFN α 3 mRNA copies. Spearman method is used for correlation analysis with Spearman's Rank Correlation Coefficient R (Rs) and p values reported (n=8).

Figure S1. Production and purification of the bIFN α calibrator.

(a) SDS-Page profile obtained before IPTG induction (\emptyset) and after induction at 16, 30 or 37°C in the soluble or insoluble extract of *Escherichia coli* cells transformed to produce bIFN α . Molecular weights (MW) are indicated in kDa (left panel). Western-Blot profile obtained from the soluble fraction after induction at 16°C (right panel). After SDS-Page migration, proteins were transferred on a nitrocellulose membrane then incubated with the IFN α 2 assay detection antibody. Streptavidin-conjugated horseradish peroxidase was added to the membrane and signal was revealed using chemiluminescence. (b) SDS-Page profile for each bIFN α purification step from the

soluble fraction (left panel) after IPTG induction at 16°C. Column load (IN), column flow-through (FT), washes (W1, W2 and W3) and elution fractions (E1 to E9). The middle panel shows the SDS-Page profile for the pools of eluted fractions. The red arrow indicates the bIFN α protein at its right molecular weight. (c) SDS-Page profile for each bIFN α purification step from the insoluble fraction (left panel) after IPTG induction at 16°C as previously described. Right panel shows SDS-Page profile for each renaturation step of the pool of eluted fractions obtained from the insoluble *E. coli* extract : 4M, then 2M, then 1M urea and finally without urea (ØM), the ultimate step where the protein is lost. Red arrows indicate the bIFN α protein at his right molecular weight.

Figure S2. Bat IFN α proteins have a similar IFN α 2 assay epitope to human IFN α proteins

Protein sequence alignment obtained for human IFN α 1, IFN α 2, IFN α 16 and IFN α 17, for *Pteropus alecto* IFN α 1, IFN α 2 and IFN α 3, for *Rousettus aegyptiacus* IFN α , and for mouse IFN α 1, IFN α 3, IFN α 4, IFN α 11 and IFN α 13 using the CLUSTALW software from position 90 to 130 referred to hIFN α 2. The red rectangle highlights the suspected epitope recognized by the antibodies of the IFN α 2 assay.

Figure S3. Age, sex and clinical symptoms were not associated with IFN α protein levels

(a) Correlation plot between age of all studied bat specimens and IFN α protein levels expressed as bIFN α equivalent concentration. Spearman method is used. (b) IFN α protein levels expressed as bIFN α equivalent concentration for female and male groups, all species combined. Median represented by black line. Mann-Whitney test is used, ns: p>0.05. (c) IFN α protein levels expressed as bIFN α equivalent concentration for healthy and mild clinical symptom groups, all species combined. Median represented by black line. Mann-Whitney test is used, ns: p>0.05.

Figure S4. Viral inactivation decreases the affinity of the assay for bIFN α but not hIFN α 2c.

IFN α 2 assay response (AEB) as a function of IFN α concentrations for hIFN α 2c and bIFN α untreated or after viral inactivation.

Tables Legends

Table 1. Demographic characteristics and origins of the bat cohort

Number of individuals, gender, species and origins for the bat cohort. Data are shown as the n (%).

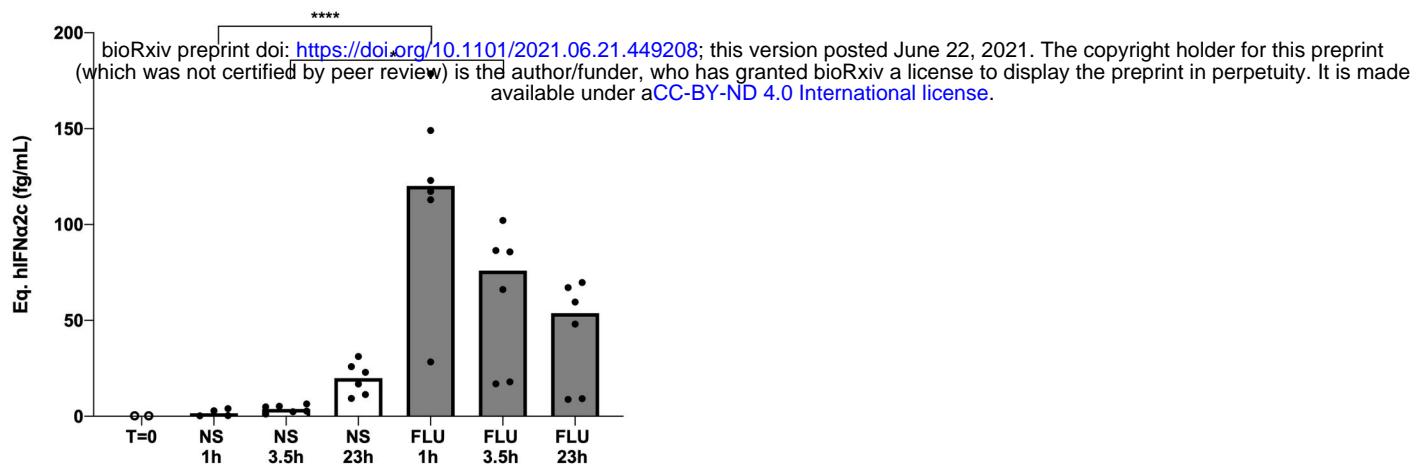
Table S1. All data available for each sample

Species, gender, GAPDH-normalized number of IFN α 1, IFN α 2 and IFN α 3 mRNA copies, interferon α concentrations expressed as hIFN α 2c or bIFN α equivalent concentrations and notes available for each sample.

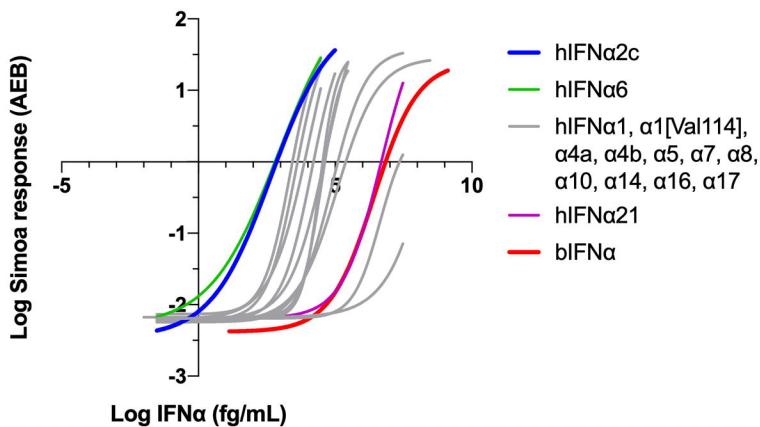
Table S2. Primers and probes sequences

List of primers and probes used in RT-qPCR for bat IFN α 1, IFN α 2, IFN α 3 and GADPH, based on published data from Zhou et al. 2016 for the bat IFN genes described for *P. alecto* and designed for this study for GADPH, from PrimerBlast and available data for GADPH gene in *P. alecto* and *R. aegyptiacus* species.

(a)



(b)



(c)

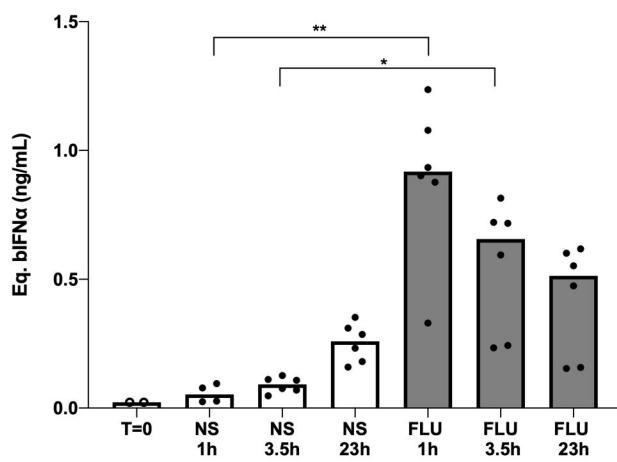
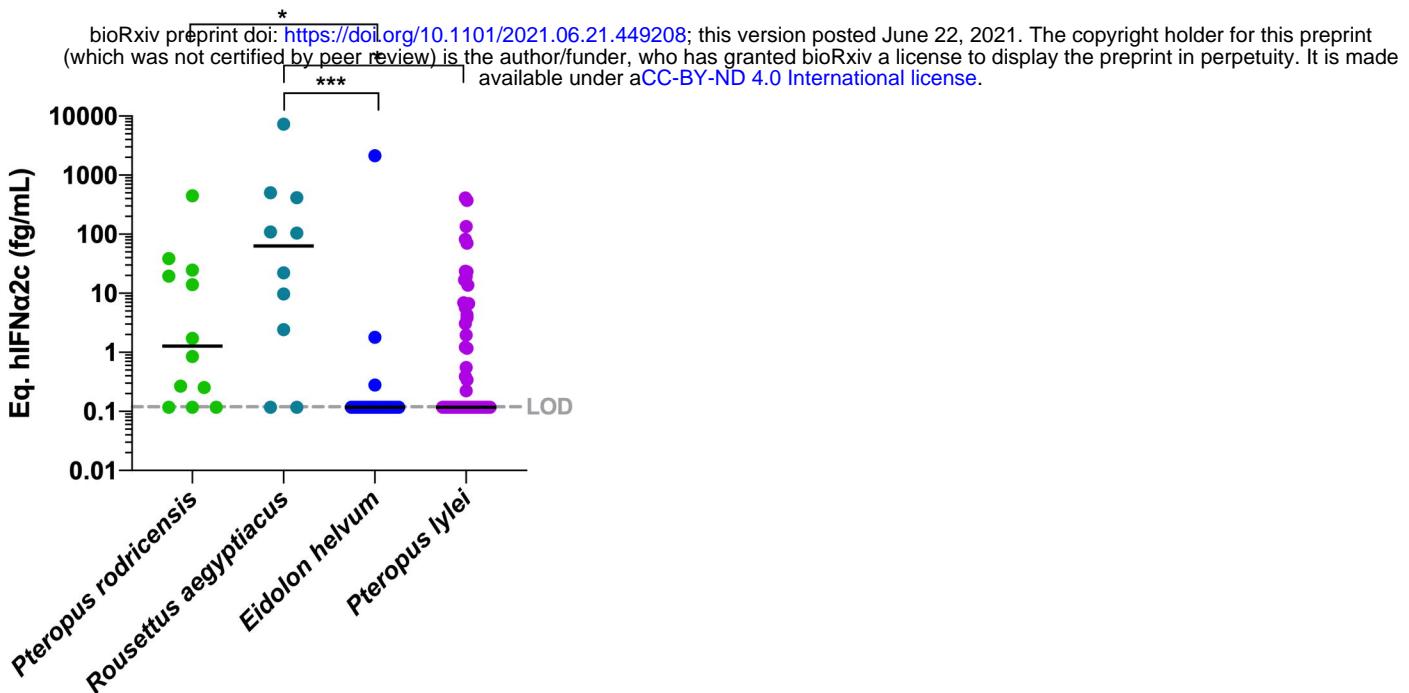
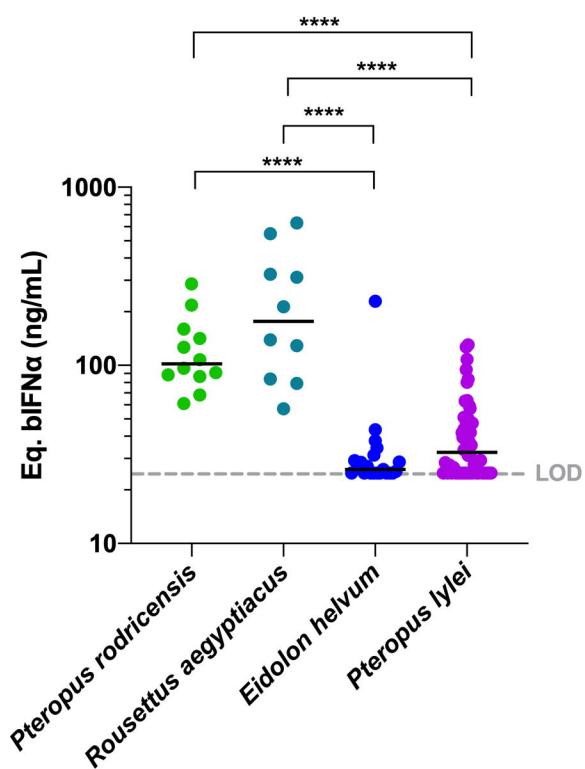


Figure 1

(a)



(b)

**Figure 2**

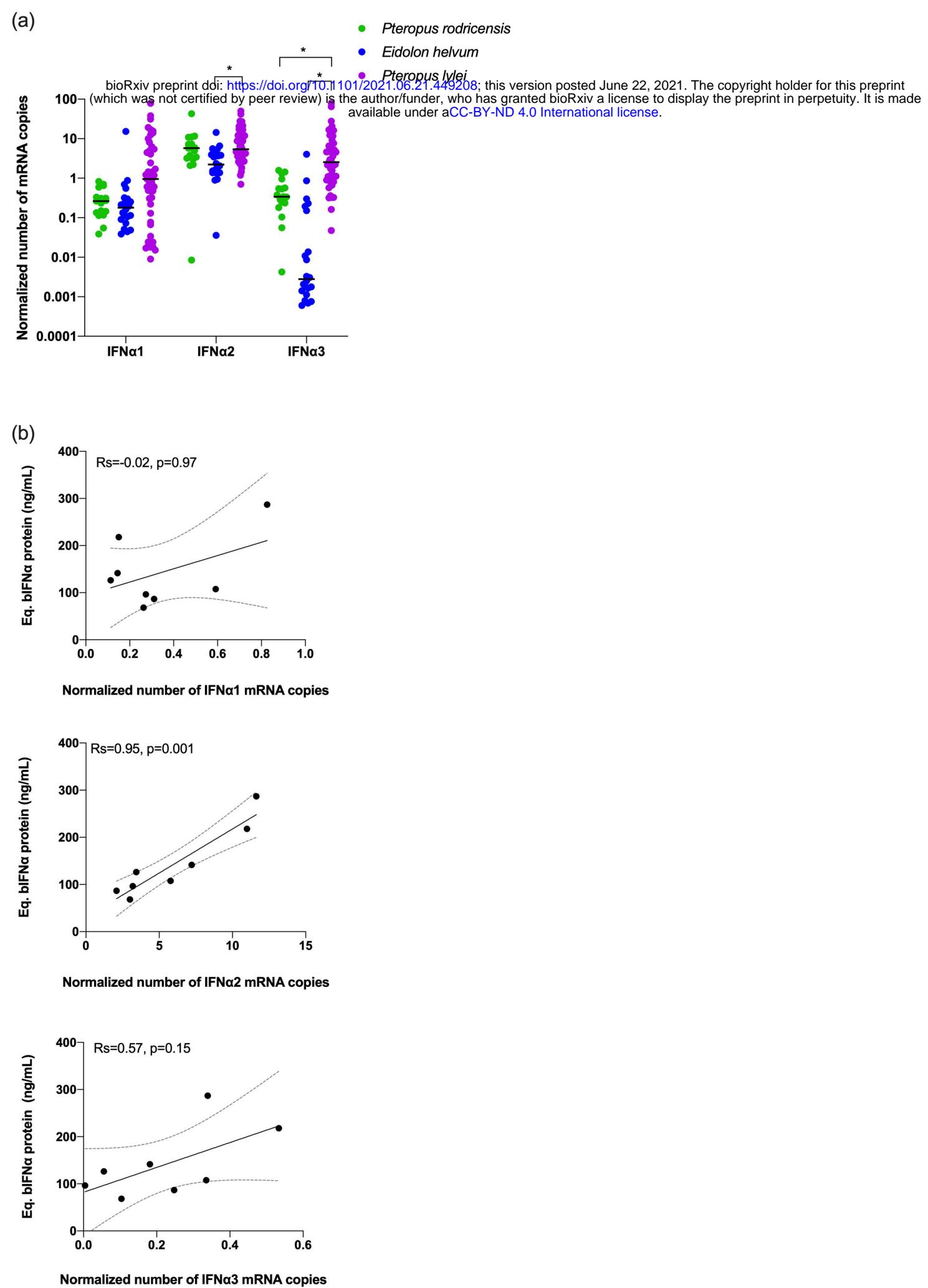


Figure 3