

1 **Title:**

2 **Predictors of cognitive impairment in primary age-related tauopathy: an autopsy study**

3 **Authors:** Megan A. Iida BS<sup>1\*</sup>, Kurt Farrell PhD<sup>1\*</sup>, , Jamie M. Walker MD, PhD<sup>2</sup>, Timothy E. Richardson  
 4 DO, PhD<sup>2</sup>, Gabe Marx<sup>1</sup>, Clare H. Bryce MD<sup>1</sup>, Dushyant Purohit MD<sup>1</sup>, Gai Ayalon PhD<sup>4</sup>, Thomas G. Beach  
 5 MD-PhD<sup>5</sup>, Eileen H. Bigio MD<sup>6</sup>, Etty Cortes MD<sup>1</sup>, Marla Gearing PhD<sup>7</sup>, Vahram Haroutunian PhD<sup>8</sup>, Corey  
 6 T. McMillan PhD<sup>9</sup>, Eddie B. Lee<sup>9</sup>, Dennis Dickson MD<sup>10</sup>, Ann C. McKee MD<sup>11</sup>, Thor D. Stein MD-PhD<sup>11</sup>,  
 7 John Q. Trojanowski MD-PhD<sup>12</sup>, Randall L. Woltjer MD<sup>13</sup>, Gabor G. Kovacs MD-PhD<sup>14,15,16</sup>, Julia K. Kofler  
 8 MD<sup>17</sup>, Jeffrey Kaye MD<sup>18</sup>, Charles L. White III MD<sup>19</sup>, John F. Crary MD-PhD<sup>1\*\*</sup>

9  
 10 1) Department of Pathology, Neuropathology Brain Bank & Research CoRE, Nash Family Department of  
 11 Neuroscience, Friedman Brain Institute, Ronald M. Loeb Center for Alzheimer's Disease, Icahn School  
 12 of Medicine at Mount Sinai, USA;

13  
 14 2) Department of Pathology and Laboratory Medicine and The Glenn Biggs Institute for Alzheimer's &  
 15 Neurodegenerative Diseases, UT Health San Antonio, San Antonio, TX, USA;

16  
 17 4) Ultragenyx Pharmaceuticals USA;

18  
 19 5) Neuropathology, Banner Sun Health Research Institute, Sun City, Arizona, USA;

20  
 21 6) Department of Pathology, Northwestern Cognitive Neurology and Alzheimer Disease Center,  
 22 Northwestern University Feinberg School of Medicine, Chicago, Illinois, USA;

23  
 24 7) Department of Pathology and Laboratory Medicine, Emory University School of Medicine, Atlanta,  
 25 Georgia, USA;

26  
 27 8) Departments of Psychiatry and Neuroscience; Alzheimer's Disease Research Center, Icahn School of  
 28 Medicine at Mount Sinai, New York, New York, USA; and JJ Peters VA Medical Center (MIRECC), Bronx,  
 29 NY.

30  
 31 9) Department of Neurology, Perelman School of Medicine, Penn FTD Center, Center for  
 32 Neurodegenerative Disease Research, University of Pennsylvania, Philadelphia, Pennsylvania, USA;

33  
 34 10) Department of Neuroscience, Mayo Clinic, Jacksonville, Florida, USA;

35  
 36 11) Department of Pathology, VA Medical Center & Boston University School of Medicine, Boston,  
 37 Massachusetts, USA;

38  
 39 12) Center for Neurodegenerative Disease Research, Department of Pathology and Laboratory Medicine,  
 40 Perelman School of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania, USA;

41  
 42 13) Department of Pathology, Oregon Health Sciences University, Portland, Oregon, USA;

43  
 44 14) Laboratory Medicine Program & Krembil Brain Institute University Health Network Toronto Ontario  
 45 Canada;

46

15) Tanz Centre for Research in Neurodegenerative Disease and Department of Laboratory Medicine and Pathobiology, University of Toronto, Toronto, Ontario, Canada;

16) Previous address: Institute of Neurology, Medical University of Vienna, Vienna, Austria;

17) Department of Pathology, University of Pittsburgh Medical Center, Pittsburgh, Pennsylvania, USA;

18) Department of Neurology, Oregon Health & Science University, Portland USA;

19) Neuropathology Laboratory, Department of Pathology, University of Texas Southwestern Medical Center, USA;

\* These authors contributed equally to this work

\*\* Correspondence:

John F. Crary, MD-PhD

Professor, Department of Pathology

Director, Neuropathology Brain Bank & Research CoRE

Nash Family Department of Neuroscience

Friedman Brain Institute

Ronald M. Loeb Center for Alzheimer's Disease

Icahn School of Medicine at Mount Sinai

1 Gustave L. Levy Place Box 1194 New York, NY 10029, USA

Telephone: (212) 659-8695, Email: [john.crary@mountsinai.org](mailto:john.crary@mountsinai.org)

The authors declare no conflicts of interest

## Acknowledgements

This work was supported by the National Institutes of Health [R01 AG054008, R01 NS095252, R01 AG060961, and R01 NS086736 to J.F.C, F32 AG056098 and P30 AG066514 to K.F., R01 AG062348 to J.F.C., A.M., and D.D., P30 AG010124, P01 AG017586 and U19 AG062418 to J.Q.T, R01 AG066152 to C.T.M, P50 AG005133 to J.K., P50 AG005138, P30 AG066514, and 75N95019C00049 to V.H., U24

80 NS072026 and P30 AG019610 to T.B., P30 AG013854 to E.B., P30 NS055077 and P50 AG025688 to  
81 M.G., P30 AG08017 to R.W. and U54 NS115266 to A.M.], the Alzheimer's Association [NIRG-15-363188  
82 to J.F.C.], the Tau Consortium, Genentech/Roche, David & Elsie Werber, Alexander Saint-Amand  
83 Fellowship, J.M.R. Barker Foundation, The McCune Foundation, and the Winspear Family Center for  
84 Research on the Neuropathology of Alzheimer Disease, The Arizona Department of Health Services, and  
85 the Michael J. Fox Foundation for Parkinson's Research. G.G.K. is supported by the Rossy Foundation  
86 and by the Safra Foundation. The authors would also like to acknowledge Ping Shang, HT(ASCP) QIHC  
87 and Jeff Harris, HTL(ASCP) for histologic and immunohistochemical preparations, and Chan Foong,  
88 M.S., for preparation of whole slide image.  
89

## Abstract

Primary age-related tauopathy (PART) is a form of Alzheimer-type neurofibrillary degeneration occurring in the absence of amyloid-beta (A $\beta$ ) plaques. While PART shares some features with Alzheimer disease (AD), such as progressive accumulation of neurofibrillary tangle pathology in the medial temporal lobe and other brain regions, it does not progress extensively to neocortical regions. Given this restricted pathoanatomical pattern and variable symptomatology, there is a need to reexamine and improve upon how PART is neuropathologically assessed and staged. We performed a retrospective autopsy study in a collection ( $n=174$ ) of post-mortem PART brains and used logistic regression to determine the extent to which a set of clinical and neuropathological features predict cognitive impairment. We compared Braak staging, which focuses on hierarchical neuroanatomical progression of AD tau and A $\beta$  pathology, with quantitative assessments of neurofibrillary burden using computer-derived positive pixel counts on digitized whole slide images of sections stained immunohistochemically with antibodies targeting abnormal hyperphosphorylated tau (p-tau) in the entorhinal region and hippocampus. We also assessed other factors affecting cognition, including aging-related tau astrogliopathy (ARTAG) and atrophy. We found no association between Braak stage and cognitive impairment when controlling for age ( $p=0.76$ ). In contrast, p-tau burden was significantly correlated with cognitive impairment even when adjusting for age ( $p=0.03$ ). The strongest correlate of cognitive impairment was cerebrovascular disease, a well-known risk factor ( $p<0.0001$ ), but other features including ARTAG ( $p=0.03$ ) and hippocampal atrophy ( $p=0.04$ ) were also associated. In contrast, sex, *APOE*, psychiatric illness, education, argyrophilic grains, and incidental Lewy bodies were not. These findings support the hypothesis that comorbid pathologies contribute to cognitive impairment in subjects with PART. Quantitative approaches beyond Braak staging are critical for advancing our understanding of the extent to which age-related tauopathy changes impact cognitive function.

**Keywords:** PART, dementia, Aging, Braak, ARTAG

## 114 Introduction

115 It is widely recognized that abnormal hyperphosphorylated tau (p-tau) deposition is a ubiquitous feature  
 116 of the aging human brain, observed in both cognitively normal subjects and in those with a range of  
 117 clinical features, including cognitive, motor and psychiatric symptoms [37]. The causes of tauopathy are  
 118 diverse, and include both genetic and environmental risk factors [48]. Autosomal dominant mutations in  
 119 the tau gene (*MAPT*) cause frontotemporal lobar degeneration and common risk alleles, notably the  
 120 *MAPT* 17q21.31 H1 haplotype, are associated with sporadic tauopathies including progressive  
 121 supranuclear palsy (PSP), corticobasal degeneration (CBD), and argyrophilic grain disease (AGD) [12].  
 122 Abnormal p-tau deposition is also seen following exposure to repetitive head trauma in contact sports  
 123 and other contexts in the setting of chronic traumatic encephalopathy (CTE) [43]. Neurofibrillary tangles  
 124 (NFT) are also a component of Alzheimer disease (AD), where they are associated amyloid-beta deposits  
 125 [16].

126 Although it is generally understood that autopsy studies are critical for establishing definitive  
 127 diagnoses, the neuropathology of the tauopathies is complex and overlapping. Further, non-impaired  
 128 individuals often display significant amounts of p-tau accumulation, complicating our understanding of  
 129 the contribution of such brain changes to symptomatology. Approaches to assessing tauopathy in post-  
 130 mortem tissues continue to evolve. Neuropathologically, tauopathies can be differentiated by the  
 131 neuroanatomical regionality of p-tau aggregates, cell type involvement (i.e., neurons versus glia),  
 132 preferential isoform accumulation, and filament ultrastructure. Based upon these differentiating features,  
 133 validated neuropathological diagnostic consensus criteria have been devised and, in some cases,  
 134 undergone revision. Examples include revision of the AD diagnostic criteria, and consensus criteria for  
 135 CTE [41, 46]. The term aging-related tau astrogliopathy (ARTAG), which was described in recent  
 136 consensus criteria on various patterns of astrocytic p-tau observed in aging, has been especially helpful  
 137 for differentiating age-related changes from CTE, both of which have perivascular p-tau deposits, but with  
 138 differences in cell types involved [38, 42]. The introduction of criteria for primary age-related tauopathy

(PART) to describe individuals who develop AD-type neurofibrillary pathology with or without dementia in the absence of significant amyloid deposition helped to better define this entity and differentiated it from AD [17]. Understanding age-related tauopathy is of critical importance in the context of diagnosis and staging of all the tauopathies given its extremely high prevalence and importance as a co-morbidity in essentially all studies evaluating tauopathy.

There has been controversy surrounding the PART consensus criteria since their introduction [11, 19], and there have been a substantial number of recent clinicopathological studies focused on understanding this pathological presentation [4, 6, 7, 29, 33, 36, 51, 52, 60]. Given the close clinical and neuropathological similarities between PART and AD such that historically the two entities were classified together, accumulating evidence has highlighted differences. Clinically, the average age is higher for individuals who have PART than those with AD and patients with PART are more often female [35]. Patients with PART pathology are more often cognitively normal, but a subset have mild cognitive impairment or amnesic dementia, and this correlates with p-tau severity [17]. Among symptomatic individuals with a neuropathological diagnosis of PART, nearly half had been clinically diagnosed with AD compared with 86% of those with autopsy-confirmed AD, indicating that despite diagnostic uncertainty, clinicians recognize differences between the two [59]. One retrospective study identified other factors including depression, Braak stage, and history of stroke, as independent predictors of cognitive impairment [6]. Another found that those with PART had a sparing of semantic memory compared to those with AD, suggesting that there is a distinct difference in clinical presentation [8]. Longitudinal analyses found that subjects with PART have a significantly slower clinical decline after becoming symptomatic than those with AD across multiple neuropsychological domains [60].

One limitation of most published studies on PART is that they rely on retrospective analysis of previously collected datasets (e.g., the National Alzheimer's Coordinating Center database, NACC) with predefined neuropathological measures that may not fully capture all the clinically relevant features [45]. Further, findings might not be generalizable to other populations, and a lack of uniform analysis and

164 quantitation might lead to bias. Critically, the Braak staging system was specifically developed for  
 165 assessment of tau pathology in the context of AD, and has not been rigorously tested in amyloid-negative  
 166 subjects, so the extent to which it is valid for staging p-tau pathology in PART is unclear. Additionally, the  
 167 Braak stage represents a hierarchical progression of the regional spread of neurofibrillary tangles, but  
 168 does not directly measure the severity or burden of p-tau, but this has been incorporated into some  
 169 operationalized frameworks [2]. Because the pathology in PART generally remains predominantly in the  
 170 medial temporal lobe, this hierarchical pathoanatomical system may sub-optimally measure severity of  
 171 the disease. There are numerous approaches to assessing lesion burden of p-tau and other pathologies  
 172 [10, 28, 30, 31, 40, 41, 44, 63], including cell counting and stereology [3, 5, 13, 21, 27, 64]. While each  
 173 of these approaches have intrinsic advantages, they are limited in that they are labor intensive and for  
 174 this reason and others, these methods have not been widely adopted in neuropathology laboratories [20,  
 175 62]. One approach that may have potential to better assess p-tau in PART is using computer-assisted  
 176 quantitative morphometrics on digital whole slide images, which may be well suited for staging PART.

177 Here, we studied a cohort of autopsy-confirmed subjects with PART, enabling us to reexamine  
 178 how tau pathology manifests in PART. We compared Braak staging with computer-assisted quantitative  
 179 measures of p-tau burden, and used logistic regression to assess their contribution to cognitive  
 180 impairment. Using this cohort, we were able to explore critical co-morbid pathologies (e.g.,  
 181 cerebrovascular disease), and further assess neuropathological changes that are not available in existing  
 182 publicly available datasets, including atrophy and ARTAG.

## 183 **Methods**

184

### 185 Patient samples

186

187 Formalin-fixed paraffin embedded (FFPE) tissue from the frontal cortex and hippocampus as well as  
 188 fresh-frozen tissue from frontal cortex were derived from autopsy brains from a subset of individuals from  
 189 a previously described collection [61]. Specifically, the cohort included cases from the Oregon Health  
 190 Sciences University (Portland, OR, USA), Banner Sun Health Research Institute (Sun City, AZ, USA),  
 191 Emory (Atlanta, GA, USA), Northwestern (Evanston, IL, USA), the University of Pennsylvania  
 192 (Philadelphia, PA, USA), University of Pittsburgh (Pittsburgh, PA, USA), University of Texas  
 193 Southwestern Medical Center (Dallas, TX, USA), and the Medical University of Vienna (Vienna, Austria).  
 194 Clinical inclusion criteria included being cognitively normal or having a diagnosis of mild cognitive  
 195 impairment (MCI) or dementia with a recorded clinical dementia rating (CDR), Mini-Mental State  
 196 Examination (MMSE), or postmortem clinical chart review CDR score within two years of death [22, 47].  
 197 CDR and MMSE scores were used to assign subjects into either cognitively normal or cognitively  
 198 impaired groups. Individuals who had a CDR score of 0.5 or above or MMSE score below 26 were  
 199 considered to be cognitively impaired while subjects with a CDR score of 0 or MMSE score 26 or above  
 200 were considered cognitively normal [39]. If an individual had both MMSE score and CDR score, the most  
 201 recent score was used, and if both scores were given on the same date, the CDR score was used.

202 Comprehensive neuropathological assessments were performed at the contributing institutions.  
 203 Neuropathological criteria for PART included (1) cases that had a Braak stage of 0-IV and (2) Consortium  
 204 to Establish a Registry for Alzheimer's Disease (CERAD) neuritic plaque severity score of 0 [10, 44].  
 205 Neuropathological exclusion criteria consisted of other neurodegenerative diseases including AD, Lewy  
 206 body disease, progressive supranuclear palsy (PSP), corticobasal degeneration (CBD), chronic traumatic  
 207 encephalopathy (CTE), Pick disease, Guam amyotrophic lateral-sclerosis-parkinsonism-dementia,  
 208 subacute sclerosing panencephalitis, globular glial tauopathy. Data pertaining to Braak stage, CERAD,



209 Lewy body pathology (incidental), cerebrovascular disease, infarcts (vascular brain injury), microinfarcts,  
210 and argyrophilic grains, were derived from neuropathologic studies performed at respective centers. The  
211 presence of aging-related tau astrogliopathy (ARTAG) was determined on p-tau immunohistochemical  
212 stains described below [38].

213

## 214 Atrophy score

215

216 Given that no widely accepted validated system for assessing hippocampal atrophy on human brain  
217 sections exists, we devised a semiquantitative scoring system and applied it to low power images of  
218 hematoxylin & eosin-stained sections counterstained with Luxol fast blue. We defined atrophy severity  
219 as the magnitude of ventricular dilatation (hydrocephalus ex vacuo) relative to the size of the hippocampal  
220 formation. If there was no apparent ventricular dilatation or atrophy, then a score of 0 was assigned. If  
221 there was appreciable atrophy, but the dorsoventral height of the ventricle was less than the height of the  
222 thickest section of CA1, then a score of 1 (mild) was assigned. If the magnitude of ventricular dilatation  
223 exceeded the thickness of CA1, then a score of 2 (moderate) was given. If the total area of the ventricle  
224 area was greater than the area of the hippocampus proper, a score of 3 (severe) was assigned. This  
225 score was derived only in the subset of cases where the entire temporal horn of the lateral ventricle was  
226 available included in the provided section ( $n=24$ ).

227

## 228 Immunohistochemistry

229

230 Immunohistochemistry (IHC) and hematoxylin & eosin (H&E) stains were performed on FFPE sections  
231 (5  $\mu$ m) that were prepared from blocks of hippocampus and frontal cortex for supplemental  
232 neuropathological analyses (see below). Sections mounted on positively charged slides were dried  
233 overnight at room temperature. IHC was performed on a Leica Bond III automated stainer, according to  
234 the manufacturer's protocols (Leica Microsystems, Buffalo Grove, IL, USA). IHC was performed using

antibodies to hyper-phosphorylated tau (p-tau, AT8, 1:1000, Fisher Scientific, Waltham, MA) and beta-amyloid (A $\beta$ , 6E10, 1:1000, Covance, Princeton, NJ, USA). A $\beta$  stains were confirmed to be negative to ensure that there were no neuritic or diffuse plaques present (CERAD score of 0) for all cases. For each set of slides stained, a known severe AD case was included as a batch control.

239

Computer-assisted morphometric analysis

241

Whole slide images (WSI) were prepared from glass slides that were scanned using an Aperio CS2 (Leica Biosystems, Wetzlar Germany) digital slide scanner. Quantitative analysis of the tau burden was performed in selected regions in the hippocampi using the following methodology; WSI were neuroanatomically segmented using Aperio ImageScope software into the hippocampus proper (i.e., dentate, cornu ammonis, and subiculum) and the adjacent cortex that we termed the entorhinal region, which variably includes posterior portions of the parahippocampal gyrus with remnants of the (trans-)entorhinal region or lingual gyrus. Staining was measured in these areas separately and together using a modified version of the Aperio positive pixel count (Version 9) based on the intensities of the positive control sample in each batch to determine the area of immunoreactivity. Data were normalized using the number of positive pixel counts to the total area creating a 0-1 p-tau burden scale.

252

Genetic analysis

254

High-throughput isolation of DNA was performed using the MagMAX DNA Multi-Sample Ultra 2.0 Kit on KingFisher Flex robotic DNA isolation system (ThermoFisher, Waltham, MA). 20-40 mg of fresh frozen brain tissue were placed into a deep-well plate and treated with 480  $\mu$ l of Proteinase K mix (Proteinase K, Phosphate Buffered Saline [pH 7.4], Binding Enhancer) and incubated overnight at 65°C at 800 rpm on a shaking plate. Genomic DNA was isolated and purified using magnetic particles. DNA quality control was performed using a nanodrop spectrophotometer (concentration > 50ng/ $\mu$ l, 260/280 ratio 1.7-2.2).

261 Genotyping was performed using single nucleotide polymorphism (SNP) microarrays (Infinium Global  
262 Screening Array v2.4. or the Infinium OmniExpress-24, Illumina, San Diego CA). Raw genotype files were  
263 converted to PLINK-compatible files using GenomeStudio software (Illumina, San Diego CA). *MAPT*  
264 haplotype was determined using the rs8070723 H2 tagging SNP. *APOE* genotype was provided by the  
265 collaborating center. For analyses, the *APOE* status was collapsed into a binary variable of the presence  
266 or absence of *APOE* ε4.

267

268 Statistical analysis

269

270 All statistical tests were performed using the statistical software Statistical Package for the Social  
271 Sciences (SPSS) (IBM, Chicago, IL). Data was visualized using the ggplot2 package in project R or Excel  
272 (Microsoft, Redmond, Washington). Binary measurements (yes/no) were created for pathological,  
273 clinical, demographic, and genetic variables. Specifically, variables were extracted from the pathological  
274 diagnosis and binary measurements (yes/no) were created for the following variables: argyrophilic grains,  
275 Lewy body pathology (incidental), cerebrovascular disease, and infarcts (vascular brain injury).  
276 Additionally, the same process was done for clinical variables: history of psychiatric illness and education  
277 (for this study, defined as at least some college).

278 Descriptive statistics were used to identify differences between the cognitively normal and  
279 cognitively impaired PART groups for clinical, pathological, and genetic variables. Differences were  
280 detected using  $\chi^2$  tests or exact  $\chi^2$  if any cell size included < 5 participants. A t-test was performed to  
281 determine if age differed significantly between normal and cognitively groups. Next, an unadjusted binary  
282 logistic regression was performed to determine what genetic, clinical, and pathological variables were  
283 associated with being cognitively impaired within our PART cohort. Lastly, a multivariable model was  
284 created to determine what extent Braak NFT stage and the computer-assisted morphometrics were able  
285 to predict cognitive impairment in PART when adjusting for age. Statistical significance was determined  
286 if  $\alpha < 0.05$ . Not all data was available on the subjects.

## Results

One hundred seventy-four neuropathologically confirmed amyloid-negative subjects were included in this study (Table 1, Figure 1). The overall mean age was 83.2 with a range of 52.9-105.1 years. Of these, 124 subjects (mean age 81.0, range = 52.9-102.4) had no cognitive impairment and 50 (mean age 88.3, range = 69.8-105.1) had some degree of cognitive impairment, with either mild cognitive impairment (MCI) or dementia. The majority of subjects who were cognitively impaired were 80+ years of age (Figure 2). The Braak NFT stage ranged from 0 to IV with the majority of cognitively impaired subjects having a Braak NFT score of II to IV. A higher percentage of females had cognitive impairment (62.0%) compared to those who were cognitively normal (49.2%).

We observed several differences among subjects with cognitive impairment compared to those who were cognitively normal. First, cognitively impaired PART subjects were more likely to be older (age of testing 81.0 vs. 88.3,  $p < 0.0001$ ), have cerebrovascular disease (42.0% vs. 4.8%,  $p < 0.0001$ ) and have hippocampal age-related tau astroglipathy (ARTAG; 38.3% vs. 21.6%,  $p < 0.05$ ) compared to cognitively normal subjects (Table 1). However, education, history of psychiatric illness, argyrophilic grains, incidental Lewy body pathology, infarcts, presence of an *APOE*  $\epsilon 2$  allele, presence of *APOE*  $\epsilon 4$  allele, and *MAPT* haplotype status did not significantly affect cognitive status ( $p > 0.05$  for all conditions).

In our main unadjusted analysis, we assessed the extent to which a series of clinical, neuropathological, and genetic variables predicted cognitive impairment in our PART cohort (Table 2). We found that age and cerebrovascular disease were the strongest predictors of cognitive impairment ( $p < 0.0001$  for both cases). ARTAG and hippocampal atrophy were also significant predictors, but to a lesser extent ( $p < 0.05$  for both cases). There were more reported men and subjects with a history of psychiatric illness, argyrophilic grains, incidental Lewy body pathology, infarcts, and microinfarcts in the cognitively impaired PART group, however none of these predictors was significantly different ( $p > 0.05$  for all conditions). *APOE*  $\epsilon 4$  (at least 1  $\epsilon 4$  allele) was reported more in the cognitively normal PART group but did not reach significance. Braak NFT stage significantly predicted cognitive impairment ( $p < 0.05$ ).

313 Additionally, the computer-assisted morphometrics in the entorhinal region, hippocampus proper, and  
 314 the combined region were significantly associated with cognitive impairment ( $p = 0.0001$ , Figure 3A-C,  
 315 Table 3). Lastly, when the Braak NFT stage was correlated with computer-assisted morphometrics in the  
 316 combined region ( $p < 0.001$ ), there was a high degree of variability between the Braak NFT stage and  
 317 the computer-assisted combined region morphometrics (Figure 3D).

318 Finally, using a multivariable model, we assessed whether any measurements for p-tau predicted  
 319 cognitive impairment when controlling for age. In this adjusted analysis, we found that computer-assisted  
 320 morphometrics used to capture p-tau burden in the hippocampus proper and combined region were  
 321 significantly associated with cognitive impairment in PART ( $p < 0.05$  for both cases). However, the  
 322 computer-assisted morphometrics in the entorhinal region were not associated with cognitive impairment  
 323 yet there was a trend toward statistical significance ( $p = 0.068$ ). The Braak NFT stage was not able to  
 324 predict cognitive impairment when controlling for age ( $p = 0.73$ , Table 3, Figure 4).

## Discussion

Since the neuropathological criteria for PART were proposed, the terminology has been widely adopted, but controversy persists, especially around its relationship to Alzheimer disease (AD). Delineating the histological/cellular features that are associated with cognitive impairment in PART is critical for advancing our understanding of the pathology and determining the extent to which it overlaps with AD. The fact that subjects with PART, as with AD neuropathologic change, can range in their cognitive status from normal to demented, raises the question as to whether cognitive reserve/resilience plays a role or alternatively whether we are not adequately capturing the relevant features, such as common comorbidities or other factors. This study, by using a large autopsy cohort with multivariate analyses, directly addresses these critical questions. The goal was to leverage our collection of post-mortem PART brains to characterize the clinical, pathological, and genetic features that are associated with cognitive impairment in PART. Additionally, we sought to compare Braak stage with pathology burden measures derived from p-tau immunohistochemistry that quantifies severity independently of neuroanatomical vulnerability. To overcome intra-center variability in tau pathology measures, we reassessed each case histologically to maximize accuracy.

We found that all of our PART definite cases had p-tau restricted mainly to the MTL (Braak NFT stage <IV), which is consistent with and supports other previous studies investigating PART [4, 17, 34]. Cases ranged in cognitive impairment with the majority of subjects being cognitively normal, and consistent with prior data, the PART subjects tended to be older than individuals with AD [17, 59]. The results of our study confirm those of previous autopsy studies showing that cerebrovascular disease predicts cognitive impairment in PART [6, 49]. Interestingly, we did not see a strong correlation between cognitive impairment and microinfarcts, while others have shown a correlation with cognition in the oldest old [14]. We did however, find novel, unreported associations of increased age, hippocampal atrophy, and ARTAG with cognitive impairment in our PART definite cohort. Similar to what has been reported by those utilizing the NACC database, our results verify those with a higher Braak NFT stage are associated with more rapid cognitive decline [33].

While these associations have yet to be reported in PART, there are numerous studies showing that age, atrophy, and ARTAG may be associated with cognitive impairment [9, 23, 32, 50, 53]. Surprisingly, we did not see increased odds of the Braak NFT stage being associated with cognitive impairment when controlling for age as has been reported in other studies [6]. However, we did find that using computer-assisted morphometrics to assess p-tau burden in the entorhinal region, hippocampus, and combined region was able to significantly predict cognitive impairment, similar to other studies [1, 13]. While Braak NFT staging is the most widely employed approach for assessing p-tau, it is limited in that it primarily focuses on regionality and not disease burden [25]. Other studies have employed both manual and computer assisted quantitative approaches that may capture aspects of pathological features with more power [24, 26, 56]. However, a majority of these approaches focuses on AD which may not be relevant in the context of PART, where p-tau pathology does not progress in the same hierarchical manner proposed by Braak in AD [10, 16]. Hence this study highlights several new methodologies to assess p-tau burden, which our results suggest to be a more accurate predictor of clinical symptomology in those with PART.

In addition to assessing p-tau burden, we also examined the effect of *APOE* status in PART as a predictor for cognitive impairment. *APOE*  $\epsilon 4$  has been strongly suggested as an important predictor of cognitive decline in AD while *APOE*  $\epsilon 2$  has been shown to be protective [15, 18, 54, 58]. However, many of these studies have been performed in AD cohorts, and in aging cohorts there has been evidence suggesting the  $\epsilon 4$  allele is not a risk factor for cognitive impairment [57]. Our data agree with that reported by Small *et al.* as we did not see an association with *APOE*  $\epsilon 4$  and cognitive impairment, which might be explained by the fact that we studied a pathologically confirmed amyloid-negative cohort. Recent work has suggested that *APOE* may exacerbate tau pathology independently of amyloid deposition [55]. Here, we failed to detect an association of cognitive impairment in PART with the *MAPT* H1 haplotype; future larger studies with more statistical power are required to delineate the genetic architecture of PART.

This study had notable limitations. There was a relatively small number of subjects in the cognitively impaired PART group ( $n=50$ ), which may weaken our power to predict cognitive impairment.

377 Additionally, because a majority of our subjects were not from longitudinally studied prospective cohorts,  
378 we were unable to obtain certain lifestyle variables, such as actual years of education and concussion  
379 history, which could potentially significantly affect our model. However, given that diagnosing PART pre-  
380 mortem is currently challenging, it would be impractical to create such a prospective cohort. We would  
381 also like to highlight the association we observed with ARTAG and cognitive status might be only due to  
382 collinearity between p-tau severity and ARTAG, with p-tau probably the driving pathology and the ARTAG  
383 association being significant because of its potential dependence on p-tau. Lastly, our study was limited  
384 to pathology of the medial temporal lobe and frontal cortex. A more exhaustive study would have  
385 incorporated a greater number of brain regions to more extensively address other potential tau-related  
386 pathologies.

387 In summary, our findings are consistent with the hypothesis that PART is an amyloid-independent  
388 tauopathy, primarily affecting the medial temporal lobe, which can present with cognitive impairment.  
389 Several demographic and neuropathological variables including age, ARTAG, cerebrovascular disease,  
390 hippocampal atrophy, Braak NFT stage, and p-tau computer assessments were significantly associated  
391 with cognitive impairment in our PART cohort. The Braak NFT stage was not a significant predictor of  
392 cognitive impairment when controlling for age, while the computer-assistant morphometrics were. These  
393 data strongly suggest that neuroanatomical staging used in AD may not be as relevant to PART given  
394 the pathology minimally spreads beyond the medial temporal lobe. Novel techniques to measure p-tau  
395 burden can further our understanding of PART pathology and associated clinical and genetic features.



**Table 1. Patient data**

|                                      | Overall           | Cognitive Status  |                   | p        |
|--------------------------------------|-------------------|-------------------|-------------------|----------|
|                                      |                   | Normal            | Impaired*         |          |
| Demographics                         |                   |                   |                   |          |
| Average age at testing (range)       | 83.2 (52.9-105.1) | 81.0 (52.9-102.4) | 88.3 (69.8-105.1) | <0.0001  |
| Total (Male / Female)                | 174 (82 / 92)     | 124 (63 / 61)     | 50 (19 / 31)      | 0.126*** |
| Age at last visit (%)                |                   |                   |                   |          |
| <60                                  | 7 (4.0)           | 7 (5.6)           | 0 (0.0)           |          |
| 60-69                                | 15 (8.6)          | 14 (11.3)         | 1 (1.7)           |          |
| 70-79                                | 33 (19.0)         | 30 (24.2)         | 3 (5.2)           |          |
| 80-89                                | 76 (43.7)         | 45 (36.3)         | 31 (53.4)         |          |
| 90+                                  | 51 (29.3)         | 28 (22.6)         | 23 (39.7)         |          |
| Education, at least some college (%) | 32 (18.4)         | 15 (78.9)         | 17 (77.3)         | 0.89     |
| History of psychiatric illness (%)   | 45 (25.9)         | 29 (31.9)         | 17 (45.9)         | 0.13     |
| Neuropathological data               |                   |                   |                   |          |
| Argyrophilic grains                  | 32 (18.4)         | 12 (9.7)          | 10 (20.0)         | 0.06     |
| Lewy body pathology (incidental)     | 16 (9.2)          | 11 (8.9)          | 5 (10.0)          | 0.82     |
| Cerebrovascular disease**            | 27 (15.5)         | 6 (4.8)           | 21 (42.0)         | <0.0001  |
| Infarcts (vascular brain injury)     | 37 (21.3)         | 24 (19.4)         | 13 (26.0)         | 0.33     |
| Hippocampus ARTAG positive (%)       | 43 (24.7)         | 25 (21.6)         | 18 (38.3)         | 0.03     |
| Genetic Data                         |                   |                   |                   |          |
| Presence of ≥1 APOE ε4 allele        | 22 (12.6)         | 16 (12.9)         | 6 (11.3)          | 0.77     |
| Presence of ≥1 APOE ε2 allele        | 46 (26.4)         | 27 (21.8)         | 19 (35.8)         | 0.06     |
| Presence of ≥1 MAPT H2               | 59 (33.9)         | 42 (36.2)         | 17 (36.2)         | 1        |

\* Mild cognitive impairment or dementia, \*\* excluding cerebral amyloid angiopathy, \*\*\*Male sex, significant values in bold (Chi squared test)

399

**Table 2. Unadjusted odds of being cognitively impaired**

|   | OR    | 95% CI     | p value           |
|---|-------|------------|-------------------|
| <i>Characteristic</i>   |       |            |                   |
| Age, at testing   | 1.08  | 1.04-1.13  | <b>&lt;0.0001</b> |
| Education, y  | 0.87  | 0.67-1.12  | 0.28              |
| Sex   | 1.69  | 0.86-3.30  | 0.13              |
| APOE (at least 1 ε4 allele)   | 0.988 | 0.36-2.70  | 0.98              |
| History of psychiatric diagnosis                                    | 1.82  | 0.83-3.98  | 0.14              |
| Aging-related tau astroglipathy (ARTAG)                             | 2.26  | 1.08-4.72  | <b>0.03</b>       |
| Argyrophilic grains   | 2.33  | 0.94-5.82  | 0.07              |
| Lewy body pathology (incidental)                                    | 1.14  | 0.38-3.47  | 0.82              |
| Cerebrovascular disease*  | 14.24 | 5.27-38.48 | <b>&lt;0.0001</b> |
| Infarcts (vascular brain injury)                                    | 1.46  | 0.68-3.17  | 0.33              |
| Microinfarcts   | 1.05  | 0.43-2.59  | 0.91              |
| Hippocampal atrophy   | 5.32  | 1.04-27.09 | <b>0.04</b>       |
| Braak NFT stage   | 1.37  | 1.03-1.83  | <b>0.03</b>       |
| <i>Computer-assisted p-tau (AT8) burden (positive pixel counts)</i> |       |            |                   |
| Entorhinal region   | 1.90  | 1.31-2.75  | <b>0.001</b>      |
| Hippocampus proper  | 2.17  | 1.48-3.20  | <b>&lt;0.0001</b> |
| Entorhinal region & Hippocampus proper                              | 2.12  | 1.44-3.11  | <b>&lt;0.0001</b> |

\* Excluding cerebral amyloid angiopathy, significant values in bold (logistic regression)

400

401

**Table 3. Odds of being cognitively impaired at death, adjusted**

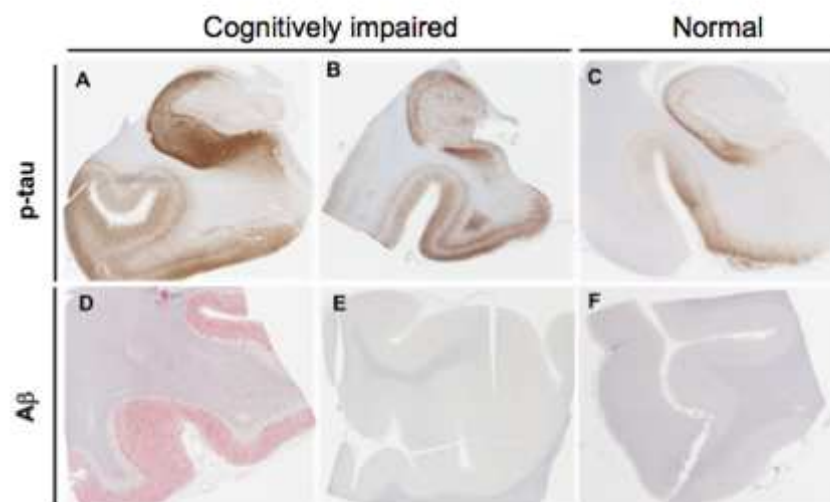
|  | <b>OR</b> | <b>95% CI</b> | <b><i>p</i> value</b> |
|--|-----------|---------------|-----------------------|
| Braak NFT stage  | 1.01      | 0.72-1.41     | 0.98                  |
| P-tau burden (computer-assisted AT8 IHC positive pixels) |           |               |                       |
| Entorhinal region  | 1.46      | 0.97-2.20     | 0.07                  |
| Hippocampus  | 1.66      | 1.07-2.57     | <b>0.02</b>           |
| Entorhinal region & hippocampus                          | 1.62      | 1.06-2.49     | <b>0.03</b>           |
| Significant values in bold (logistic regression)         |           |               |                       |

402

403

404

**Figure 1**



405

406 **Figure 1. Comparison of amyloid and tau pathology in primary age-related tauopathy**  
407 **(PART) versus Alzheimer disease (AD). (A) Immunohistochemical staining using antisera**  
408 **to hyperphosphorylated tau in an AD brain shows marked hyperphosphorylated tau (p-tau)-**  
409 **containing neurofibrillary tangles (NFT) in the hippocampus which extends past the collateral**  
410 **sulcus into the parahippocampal gyrus and other neocortical regions. (B, C) Subjects with mild**  
411 **to severe PART have elevated p-tau levels in the hippocampus predominantly restricted to the**  
412 **medial temporal lobe. (D, E, F) Subjects with AD neuropathologic change have abundant Aβ-**  
413 **containing plaques in neocortical structures, whereas those with PART have sparse or none.**  
414 **These neuropathologic changes in AD and PART are seen in association with varying degree**  
415 **of cognitive impairment ranging from cognitively normal to demented.**

416

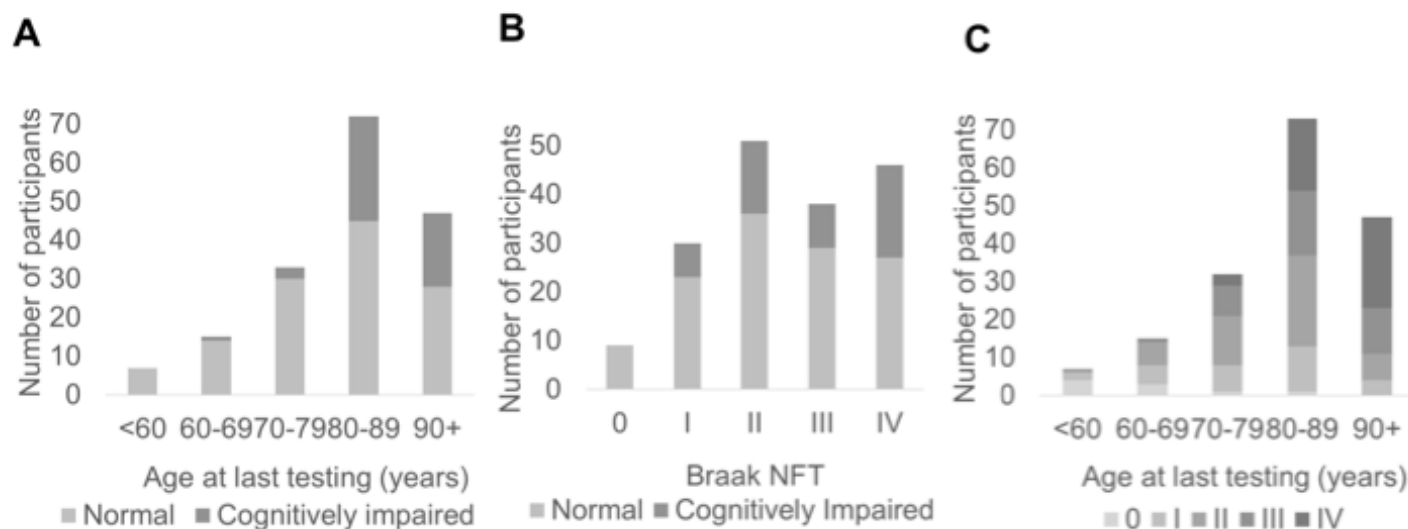
417

418

419

420

**Figure 2**



421

422 **Figure 2. Distribution of age, Braak neurofibrillary tangle (NFT) stage and cognitive**

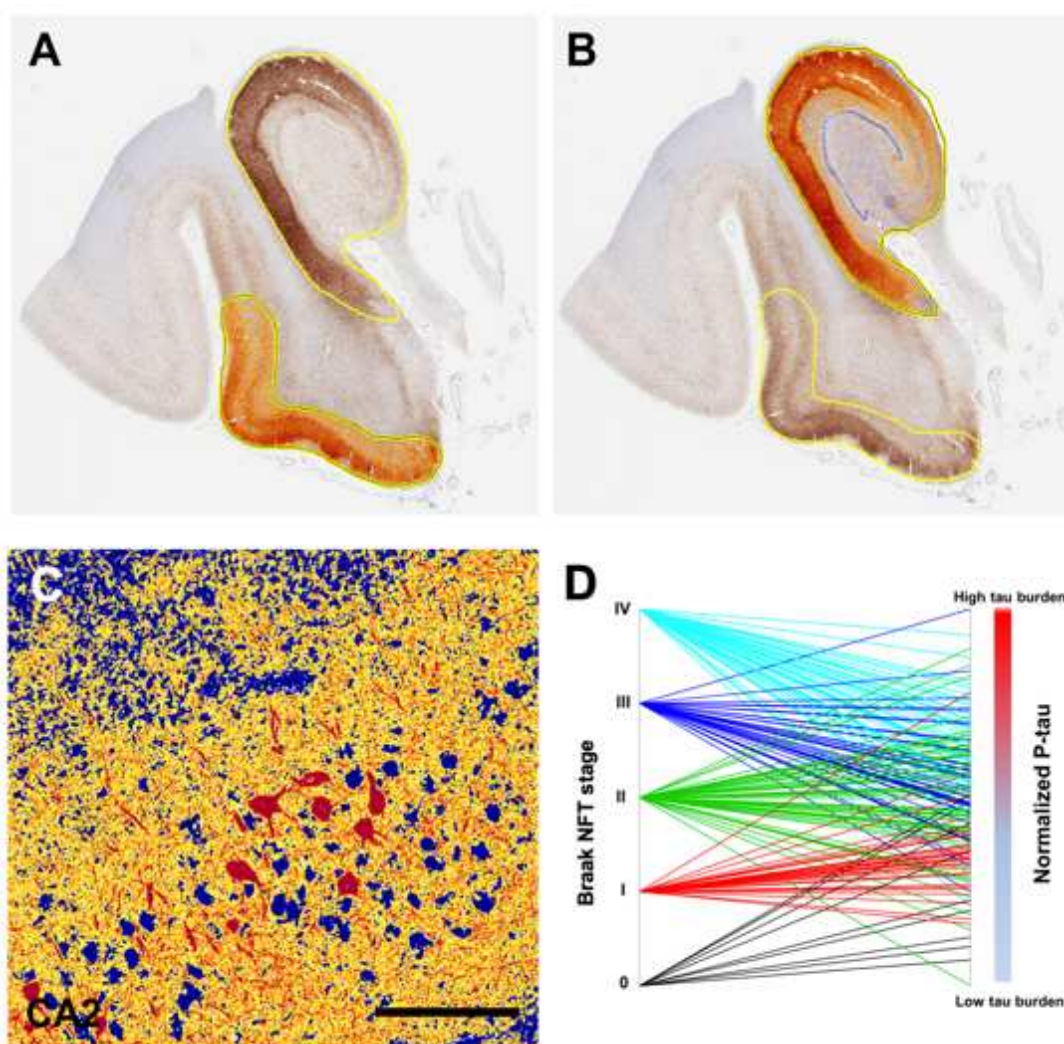
423 **status. (A)** The number of normal and cognitively impaired subjects across the age spectrum.

424 **(B)** The number of cognitively normal and impaired subjects by Braak stage. **(C)** The number

425 of subjects across the aging spectrum by Braak stage.

426

**Figure 3**



427

428 **Figure 3. Computer-assisted morphometrics to assess pathological tau burden. (A, B)**

429 Quantitative assessment of hyperphosphorylated tau (p-tau) burden was performed on whole

430 slide images of the hippocampus stained for p-tau (AT8) using immunohistochemistry. Positive

431 pixel counts were determined in two regions (hippocampus proper and entorhinal region).

432 Results were normalized to the total area assessed. A third summary score of the total p-tau

433 burden of the medial temporal lobe was calculated by summing positive pixels in both. (C) High

434 power image shows high intensity in red, medium intensity in yellow and negative staining in

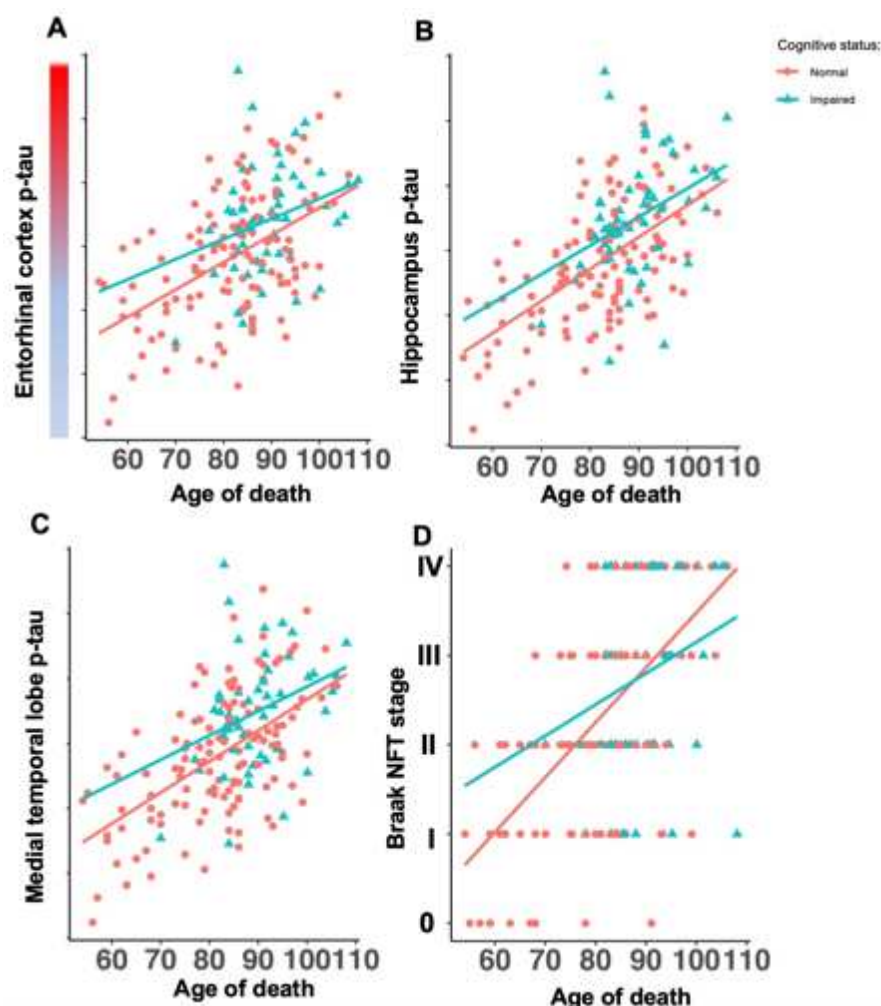
435 blue. (D) Parallel plot showing the relationship between Braak stage and the computer

436 morphometric quantification of p-tau using the normalized medial temporal lobe (hippocampus  
437 and entorhinal region). Scale bar = 150  $\mu$ m.



438

Figure 4



439

440 **Figure 4. Pathological tau burden in normal and cognitively impaired subjects across the**  
441 **aging spectrum. (A-C)** Generalized linear models of age versus tau burden show significant  
442 differences between cognitively normal and cognitively impaired subjects in the hippocampus  
443 proper ( $p = 0.047$ ), and combined entorhinal region and hippocampus regions ( $p < 0.048$ ), but  
444 not in the entorhinal region alone ( $p = 0.07$ ). **(D)** Generalized linear model of age vs Braak NFT  
445 staging did not show significant differences between cognitively normal and cognitively impaired  
446 subjects ( $p = 0.73$ ).



## 447    **References**

- 448    1        Abner EL, Neltner JH, Jicha GA, Patel E, Anderson SL, Wilcock DM, Van Eldik LJ, Nelson  
449           PT (2018) Diffuse Amyloid-beta Plaques, Neurofibrillary Tangles, and the Impact of APOE  
450           in Elderly Persons' Brains Lacking Neuritic Amyloid Plaques. *J Alzheimers Dis* 64: 1307-  
451           1324 Doi 10.3233/JAD-180514
- 452    2        Alafuzoff I, Arzberger T, Al-Sarraj S, Bodi I, Bogdanovic N, Braak H, Bugiani O, Del-  
453           Tredici K, Ferrer I, Gelpi E et al (2008) Staging of neurofibrillary pathology in Alzheimer's  
454           disease: a study of the BrainNet Europe Consortium. *Brain pathology* 18: 484-496 Doi  
455           10.1111/j.1750-3639.2008.00147.x
- 456    3        Attems J, Neltner JH, Nelson PT (2014) Quantitative neuropathological assessment to  
457           investigate cerebral multi-morbidity. *Alzheimers Res Ther* 6: Doi ARTN 85  
458           10.1186/s13195-014-0085-y
- 459    4        Bell WR, An Y, Kageyama Y, English C, Rudow GL, Pletnikova O, Thambisetty M, O'Brien  
460           R, Moghekar AR, Albert M Set al (2019) Neuropathologic, genetic, and longitudinal  
461           cognitive profiles in primary age-related tauopathy (PART) and Alzheimer's disease.  
462           *Alzheimers Dement* 15: 8-16 Doi 10.1016/j.jalz.2018.07.215
- 463    5        Bennett DA, Buchman AS, Boyle PA, Barnes LL, Wilson RS, Schneider JA (2018)  
464           Religious Orders Study and Rush Memory and Aging Project. *Adv Alzh Dis* 6: 159-187  
465           Doi 10.3233/978-1-61499-876-1-159
- 466    6        Besser LM, Crary JF, Mock C, Kukull WA (2017) Comparison of symptomatic and  
467           asymptomatic persons with primary age-related tauopathy. *Neurology* 89: 1707-1715 Doi  
468           10.1212/WNL.0000000000004521
- 469    7        Besser LM, Mock C, Teylan MA, Hassenstab J, Kukull WA, Crary JF (2019) Differences  
470           in Cognitive Impairment in Primary Age-Related Tauopathy Versus Alzheimer Disease. *J*  
471           *Neuropathol Exp Neurol*: Doi 10.1093/jnen/nly132
- 472    8        Besser LM, Mock C, Teylan MA, Hassenstab J, Kukull WA, Crary JF (2019) Differences  
473           in Cognitive Impairment in Primary Age-Related Tauopathy Versus Alzheimer Disease. *J*  
474           *Neuropathol Exp Neurol* 78: 219-228 Doi 10.1093/jnen/nly132
- 475    9        Bishop NA, Lu T, Yankner BA (2010) Neural mechanisms of ageing and cognitive decline.  
476           *Nature* 464: 529-535 Doi 10.1038/nature08983

477 10 Braak H, Braak E (1991) Neuropathological stageing of Alzheimer-related changes. *Acta*  
478 *neuropathologica* 82: 239-259 Doi 10.1007/bf00308809

479 11 Braak H, Del Tredici K (2014) Are cases with tau pathology occurring in the absence of  
480 Abeta deposits part of the AD-related pathological process? *Acta neuropathologica* 128:  
481 767-772 Doi 10.1007/s00401-014-1356-1

482 12 Caillet-Boudin ML, Buee L, Sergeant N, Lefebvre B (2015) Regulation of human MAPT  
483 gene expression. *Molecular neurodegeneration* 10: 28 Doi 10.1186/s13024-015-0025-8

484 13 Cherry JD, Tripodis Y, Alvarez VE, Huber B, Kiernan PT, Daneshvar DH, Mez J,  
485 Montenegro PH, Solomon TM, Alosco MLet al (2016) Microglial neuroinflammation  
486 contributes to tau accumulation in chronic traumatic encephalopathy. *Acta Neuropathol*  
487 *Com* 4: Doi UNSP 112 10.1186/s40478-016-0382-8

488 14 Corrada MM, Sonnen JA, Kim RC, Kawas CH (2016) Microinfarcts are common and  
489 strongly related to dementia in the oldest-old: The 90+ study. *Alzheimers Dement* 12:  
490 900-908 Doi 10.1016/j.jalz.2016.04.006

491 15 Cosentino S, Scarmeas N, Helzner E, Glymour MM, Brandt J, Albert M, Blacker D, Stern  
492 Y (2008) APOE epsilon 4 allele predicts faster cognitive decline in mild Alzheimer  
493 disease. *Neurology* 70: 1842-1849 Doi 10.1212/01.wnl.0000304038.37421.cc

494 16 Crary JF (2016) Primary age-related tauopathy and the amyloid cascade hypothesis: the  
495 exception that proves the rule? *J Neurol Neuromedicine* 1: 53-57

496 17 Crary JF, Trojanowski JQ, Schneider JA, Abisambra JF, Abner EL, Alafuzoff I, Arnold SE,  
497 Attems J, Beach TG, Bigio EH et al (2014) Primary age-related tauopathy (PART): a  
498 common pathology associated with human aging. *Acta neuropathologica* 128: 755-766  
499 Doi 10.1007/s00401-014-1349-0

500 18 Dik MG, Jonker C, Comijs HC, Bouter LM, Twisk JW, van Kamp GJ, Deeg DJ (2001)  
501 Memory complaints and APOE-epsilon4 accelerate cognitive decline in cognitively  
502 normal elderly. *Neurology* 57: 2217-2222 Doi 10.1212/wnl.57.12.2217

503 19 Duyckaerts C, Braak H, Brion JP, Buee L, Del Tredici K, Goedert M, Halliday G, Neumann  
504 M, Spillantini MG, Tolnay Met al (2015) PART is part of Alzheimer disease. *Acta*  
505 *neuropathologica* 129: 749-756 Doi 10.1007/s00401-015-1390-7

- Farfel JM, Yu L, De Jager PL, Schneider JA, Bennett DA (2016) Association of APOE with tau-tangle pathology with and without beta-amyloid. *Neurobiology of Aging* 37: 19-25 Doi 10.1016/j.neurobiolaging.2015.09.011
- Farrell K, Cosentino S, Iida MA, Chapman S, Bennett DA, Faust PL, Louis ED, Crary JF (2019) Quantitative Assessment of Pathological Tau Burden in Essential Tremor: A Postmortem Study. *J Neuropath Exp Neur* 78: 31-37 Doi 10.1093/jnen/nly104
- Folstein MF, Robins LN, Helzer JE (1983) The Mini-Mental State Examination. *Arch Gen Psychiatry* 40: 812 Doi 10.1001/archpsyc.1983.01790060110016
- Fox NC, Scahill RI, Crum WR, Rossor MN (1999) Correlation between rates of brain atrophy and cognitive decline in AD. *Neurology* 52: 1687-1689 Doi 10.1212/Wnl.52.8.1687
- Giannakopoulos P, Herrmann FR, Bussiere T, Bouras C, Kovari E, Perl DP, Morrison JH, Gold G, Hof PR (2003) Tangle and neuron numbers, but not amyloid load, predict cognitive status in Alzheimer's disease. *Neurology* 60: 1495-1500 Doi 10.1212/01.wnl.0000063311.58879.01
- Gold G, Bouras C, Kovari E, Canuto A, Glaria BG, Malky A, Hof PR, Michel JP, Giannakopoulos P (2000) Clinical validity of Braak neuropathological staging in the oldest-old. *Acta neuropathologica* 99: 579-582; discussion 583-574 Doi 10.1007/s004010051163
- Gomez-Isla T, Hollister R, West H, Mui S, Growdon JH, Petersen RC, Parisi JE, Hyman BT (1997) Neuronal loss correlates with but exceeds neurofibrillary tangles in Alzheimer's disease. *Ann Neurol* 41: 17-24 Doi 10.1002/ana.410410106
- Hamasaki H, Honda H, Okamoto T, Koyama S, Suzuki SO, Ohara T, Ninomiya T, Kiyohara Y, Iwaki T (2017) Recent Increases in Hippocampal Tau Pathology in the Aging Japanese Population: The Hisayama Study. *J Alzheimers Dis* 55: 613-624 Doi 10.3233/Jad-160521
- Hauw JJ, Daniel SE, Dickson D, Horoupian DS, Jellinger K, Lantos PL, McKee A, Tabaton M, Litvan I (1994) Preliminary NINDS neuropathologic criteria for Steele-Richardson-Olszewski syndrome (progressive supranuclear palsy). *Neurology* 44: 2015-2019 Doi 10.1212/wnl.44.11.2015

- Hickman RA, Flowers XE, Wisniewski T (2020) Primary Age-Related Tauopathy (PART): Addressing the Spectrum of Neuronal Tauopathic Changes in the Aging Brain. *Curr Neurol Neurosci Rep* 20: 39 Doi 10.1007/s11910-020-01063-1
- Hyman BT (1998) New neuropathological criteria for Alzheimer disease. *Arch Neurol* 55: 1174-1176 Doi 10.1001/archneur.55.9.1174
- Jack CR, Jr., Bennett DA, Blennow K, Carrillo MC, Dunn B, Haeberlein SB, Holtzman DM, Jagust W, Jessen F, Karlawish J et al (2018) NIA-AA Research Framework: Toward a biological definition of Alzheimer's disease. *Alzheimers Dement* 14: 535-562 Doi 10.1016/j.jalz.2018.02.018
- Jack CR, Petersen RC, Xu Y, O'Brien PC, Smith GE, Ivnik RJ, Boeve BF, Tangalos EG, Kokmen E (2000) Rates of hippocampal atrophy correlate with change in clinical status in aging and AD. *Neurology* 55: 484-489 Doi 10.1212/Wnl.55.4.484
- Jefferson-George KS, Wolk DA, Lee EB, McMillan CT (2017) Cognitive decline associated with pathological burden in primary age-related tauopathy. *Alzheimers Dement* 13: 1048-1053 Doi 10.1016/j.jalz.2017.01.028
- Jellinger KA, Alafuzoff I, Attems J, Beach TG, Cairns NJ, Crary JF, Dickson DW, Hof PR, Hyman BT, Jack CR, Jr. et al (2015) PART, a distinct tauopathy, different from classical sporadic Alzheimer disease. *Acta neuropathologica* 129: 757-762 Doi 10.1007/s00401-015-1407-2
- Jellinger KA, Attems J (2007) Neurofibrillary tangle-predominant dementia: comparison with classical Alzheimer disease. *Acta neuropathologica* 113: 107-117 Doi 10.1007/s00401-006-0156-7
- Josephs KA, Murray ME, Tosakulwong N, Whitwell JL, Knopman DS, Machulda MM, Weigand SD, Boeve BF, Kantarci K, Petrucelli L et al (2017) Tau aggregation influences cognition and hippocampal atrophy in the absence of beta-amyloid: a clinico-imaging-pathological study of primary age-related tauopathy (PART). *Acta neuropathologica* 133: 705-715 Doi 10.1007/s00401-017-1681-2
- Kovacs GG (2015) Invited review: Neuropathology of tauopathies: principles and practice. *Neuropathology and applied neurobiology* 41: 3-23 Doi 10.1111/nan.12208
- Kovacs GG, Ferrer I, Grinberg LT, Alafuzoff I, Attems J, Budka H, Cairns NJ, Crary JF, Duyckaerts C, Ghetti B et al (2016) Aging-related tau astrogliopathy (ARTAG):

harmonized evaluation strategy. *Acta neuropathologica* 131: 87-102 Doi 10.1007/s00401-015-1509-x

39 Kvitting AS, Fallman K, Wressle E, Marcusson J (2019) Age-Normative MMSE Data for Older Persons Aged 85 to 93 in a Longitudinal Swedish Cohort. *J Am Geriatr Soc* 67: 534-538 Doi 10.1111/jgs.15694

40 Markesbery WR (1997) Neuropathological criteria for the diagnosis of Alzheimer's disease. *Neurobiol Aging* 18: S13-19 Doi 10.1016/s0197-4580(97)00064-x

41 McKee AC, Cairns NJ, Dickson DW, Folkerth RD, Keene CD, Litvan I, Perl DP, Stein TD, Vonsattel JP, Stewart Wet al (2016) The first NINDS/NIBIB consensus meeting to define neuropathological criteria for the diagnosis of chronic traumatic encephalopathy. *Acta neuropathologica* 131: 75-86 Doi 10.1007/s00401-015-1515-z

42 McKee AC, Stein TD, Crary JF, Bieniek KF, Cantu RC, Kovacs GG (2020) Practical Considerations in the Diagnosis of Mild Chronic Traumatic Encephalopathy and Distinction From Age-Related Tau Astroglipathy. *J Neuropathol Exp Neurol* 79: 921-924 Doi 10.1093/jnen/nlaa047

43 McKee AC, Stein TD, Nowinski CJ, Stern RA, Daneshvar DH, Alvarez VE, Lee HS, Hall G, Wojtowicz SM, Baugh CMet al (2013) The spectrum of disease in chronic traumatic encephalopathy. *Brain* 136: 43-64 Doi 10.1093/brain/aws307

44 Mirra SS, Heyman A, McKeel D, Sumi SM, Crain BJ, Brownlee LM, Vogel FS, Hughes JP, van Belle G, Berg L (1991) The Consortium to Establish a Registry for Alzheimer's Disease (CERAD). Part II. Standardization of the neuropathologic assessment of Alzheimer's disease. *Neurology* 41: 479-486 Doi 10.1212/wnl.41.4.479

45 Mock C, Teylan M, Beecham G, Besser L, Cairns NJ, Crary JF, Katsumata Y, Nelson PT, Kukull W (2020) The Utility of the National Alzheimer's Coordinating Center's Database for the Rapid Assessment of Evolving Neuropathologic Conditions. *Alzheimer Dis Assoc Disord* 34: 105-111 Doi 10.1097/WAD.0000000000000380

46 Montine TJ, Phelps CH, Beach TG, Bigio EH, Cairns NJ, Dickson DW, Duyckaerts C, Frosch MP, Masliah E, Mirra SSet al (2012) National Institute on Aging-Alzheimer's Association guidelines for the neuropathologic assessment of Alzheimer's disease: a practical approach. *Acta neuropathologica* 123: 1-11 Doi 10.1007/s00401-011-0910-3

47 Morris JC (1993) The Clinical Dementia Rating (CDR): current version and scoring rules.  
 48 Neurology 43: 2412-2414 Doi 10.1212/wnl.43.11.2412-a

48 Morris M, Maeda S, Vessel K, Mucke L (2011) The many faces of tau. Neuron 70: 410-  
 426 Doi 10.1016/j.neuron.2011.04.009

49 Pierce AL, Kawas CH (2017) Dementia in the oldest old: Beyond Alzheimer disease.  
 PLoS Med 14: e1002263 Doi 10.1371/journal.pmed.1002263

50 Planche V, Coupe P, Helmer C, Le Goff M, Amieva H, Tison F, Dartigues JF, Catheline  
 G (2019) Evolution of brain atrophy subtypes during aging predicts long-term cognitive  
 decline and future Alzheimer's clinical syndrome. Neurobiology of Aging 79: 22-29 Doi  
 10.1016/j.neurobiolaging.2019.03.006

51 Quintas-Neves M, Teylan MA, Besser L, Soares-Fernandes J, Mock CN, Kukull WA,  
 Crary JF, Oliveira TG (2019) Magnetic resonance imaging brain atrophy assessment in  
 primary age-related tauopathy (PART). Acta Neuropathol Commun 7: 204 Doi  
 10.1186/s40478-019-0842-z

52 Robinson AC, Davidson YS, Roncaroli F, Minshull J, Tinkler P, Horan MA, Payton A,  
 Pendleton N, Mann DMA (2021) Early changes in visuospatial episodic memory can help  
 distinguish primary age-related tauopathy from Alzheimer's disease. Neuropathology and  
 applied neurobiology: Doi 10.1111/nan.12726

53 Robinson JL, Corrada MM, Kovacs GG, Dominique M, Caswell C, Xie SX, Lee VMY,  
 Kawas CH, Trojanowski JQ (2018) Non-Alzheimer's contributions to dementia and  
 cognitive resilience in The 90+Study. Acta neuropathologica 136: 377-388 Doi  
 10.1007/s00401-018-1872-5

54 Serrano-Pozo A, Qian J, Monsell SE, Betensky RA, Hyman BT (2015) APOE epsilon 2 is  
 associated with milder clinical and pathological Alzheimer disease. Annals of Neurology  
 77: 917-929 Doi 10.1002/ana.24369

55 Shi Y, Yamada K, Liddel SA, Smith ST, Zhao L, Luo W, Tsai RM, Spina S, Grinberg  
 LT, Rojas JC et al (2017) ApoE4 markedly exacerbates tau-mediated neurodegeneration  
 in a mouse model of tauopathy. Nature 549: 523-527 Doi 10.1038/nature24016

56 Signaevsky M, Prastawa M, Farrell K, Tabish N, Baldwin E, Han N, Iida MA, Koll J, Bryce  
 C, Purohit D et al (2019) Artificial intelligence in neuropathology: deep learning-based  
 assessment of tauopathy. Lab Invest 99: 1019-1029 Doi 10.1038/s41374-019-0202-4



- Small BJ, Graves AB, McEvoy CL, Crawford FC, Mullan M, Mortimer JA (2000) Is APOE-epsilon 4 a risk factor for cognitive impairment in normal aging? *Neurology* 54: 2082-2088  
Doi 10.1212/Wnl.54.11.2082
- Small BJ, Rosnick CB, Fratiglioni L, Backman L (2004) Apolipoprotein E and cognitive performance: a meta-analysis. *Psychol Aging* 19: 592-600 Doi 10.1037/0882-7974.19.4.592
- Teylan M, Besser LM, Crary JF, Mock C, Gauthreaux K, Thomas NM, Chen YC, Kukull WA (2019) Clinical diagnoses among individuals with primary age-related tauopathy versus Alzheimer's neuropathology. *Lab Invest* 99: 1049-1055 Doi 10.1038/s41374-019-0186-0
- Teylan M, Mock C, Gauthreaux K, Chen YC, Chan KCG, Hassenstab J, Besser LM, Kukull WA, Crary JF (2020) Cognitive trajectory in mild cognitive impairment due to primary age-related tauopathy. *Brain* 143: 611-621 Doi 10.1093/brain/awz403
- Walker JM, Richardson TE, Farrell K, Iida MA, Foong C, Shang P, Attems J, Ayalon G, Beach TG, Bigio EH et al (2021) Early Selective Vulnerability of the CA2 Hippocampal Subfield in Primary Age-Related Tauopathy. *J Neuropathol Exp Neurol* 80: 102-111 Doi 10.1093/jnen/nlaa153
- West MJ, Slomianka L, Gundersen HJ (1991) Unbiased stereological estimation of the total number of neurons in the subdivisions of the rat hippocampus using the optical fractionator. *Anat Rec* 231: 482-497 Doi 10.1002/ar.1092310411
- Zhukareva V, Trojanowski JQ, Lee VM (2004) Assessment of pathological tau proteins in frontotemporal dementias: qualitative and quantitative approaches. *Am J Geriatr Psychiatry* 12: 136-145
- Zhukareva V, Trojanowski JQ, Lee VM (2004) Assessment of pathological tau proteins in frontotemporal dementias - Qualitative and quantitative approaches. *Am J Geriatr Psychiatry* 12: 136-145 Doi 10.1176/appi.ajgp.12.2.136