# *AI334 and AQ806 antibodies recognize the spike S protein from SARS-CoV-2 by ELISA*

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## Abstract

We tested 10 recombinant antibodies directed against the spike S protein from SARS-CoV-1. Among them, antibodies AI334 and AQ806 detect by ELISA the spike S protein from SARS-CoV-2.

## Introduction

The spike (S) glycoprotein (UniProt #P0DTC2) mediates attachment of coronaviruses to the host ACE2 receptor (through the Receptor-Binding Domain [RBD] in the S1 subunit) and fusion with the host cell membrane (through the S2 subunit) (Yan *et al.*, 2020). Here we describe the ability of two recombinant antibodies (AI334 and AQ806) to detect by ELISA the soluble ectodomain of the S protein from SARS-CoV-2.

## Materials & Methods

**Antibodies:** ABCD_AA831, ABCD_AF167, ABCD_AF618, ABCD_AH286, ABCD_AH287, ABCD_AH971, ABCD_AH974, ABCD_AI334, ABCD_AQ601, ABCD_AQ602, and ABCD_AQ806 antibodies (ABCD nomenclature, https://web.expasy.org/abcd/) were produced by the Geneva Antibody Facility (http://www.unige.ch/medecine/antibodies/) as mini-antibodies with the antigen-binding scFv portion fused to a mouse IgG2A Fc. The synthesized scFv sequences (GeneArt, Invitrogen) correspond to the sequences of the variable regions joined by a peptide linker $(GGGGS)_3$ (see Table 1 for clone names and references). HEK293 suspension cells (growing in FreeStyle™ 293 Expression Medium, Gibco #12338) were transiently transfected with the vector coding for the scFv-Fc of each antibody. Supernatants (see Table 1 for individual yields) were collected after 5 days.

**Table 1**: Clone number, epitope, reference and production yields for the antibodies used in this study.

| ABCD | Clone | Epitope | Reference | Yield (mg/L) |
|---|---|---|---|---|
| **AA831** | 80R | S1/RBD | Sui *et al.*, 2004 | 100 |
| **AF167** | S230.15 | S1/RBD | Rockx *et al.*, 2008 | 80 |
| **AF618** | S227.14 | S1/RBD | Rockx *et al.*, 2008 | 80 |
| **AH286** | m396 | S1/RBD | Prabakaran *et al.*, 2006 | 120 |
| **AH287** | F26G19 | S1/RBD | Berry *et al.*, 2004 | <5 |
| **AH974** | CR3013 | S1 | van den Brink *et al.*, 2005 | 120 |
| **AI334** | CR3022 | S1 | ter Meulen *et al.*, 2006 | 50 |
| **AQ601** | 3C7 | S1/RBD | Coughlin *et al.*, 2007 | 50 |
| **AQ602** | 4D4 | S1 | Coughlin *et al.*, 2007 | <5 |
| **AQ806** | VHH-72 | S1/RBD | Wrapp *et al.*, 2020a | 100 |
| **AH971** | CR3009 | N protein | van den Brink *et al.*, 2005 | 150 |

**Antigen:** The prefusion ectodomain (residues 1-1208) of the SARS-CoV-2 S protein, with a KV->PP substitution at residues 986/987, a RRAR->GSAS substitution at residues 682-685, and C-terminal T4 fibritin trimerization motif, protease cleavage site, TwinStrepTag and 8xHisTag (PDB #6VSB; Wrapp *et al.*, 2020b), was transiently transfected into $25x10^8$ suspension-adapted ExpiCHO cells (Thermo Fisher) using 1.5 mg plasmid DNA and 7.5 mg of PEI MAX (Polysciences) in 500 mL ProCHO5 medium (Lonza). Incubation with agitation was continued at 31°C and 4.5% $CO_2$ for 5 days. The clarified supernatant was purified in two steps: via a Strep-Tactin XT column (IBA Lifesciences) followed by Superose 6 10/300 GL column (GE Healthcare) to a final concentration of 180 µg/ml in PBS.

**Protocol:** S protein (10 µg/ml, 50 µl/well in PBS 0.5% (w/v) BSA, 0.1% (w/v) Tween20) was immobilized on streptavidin-coated ELISA plates (Pierce #15124) for 30 min. Each well was rinsed three times with 100 µl of washing buffer (PBS + 0.5% (w/v) BSA + 0.05% (w/v) Tween20), then incubated for 1 hour with 50 µl of each antibody-containing supernatant diluted in washing buffer (Fig. 1). After rinsing 3 times (100 µl washing buffer), wells were incubated with horseradish peroxidase-coupled goat anti-mouse IgG (Bio-Rad #170-6516, dilution 1:1000, 50 µl per well) for 30 min. After 3 rinses, Tetramethylbenzidine (TMB) substrate (Sigma #T5569) was added (50 µl per well). The reaction was stopped by the addition of 25 µl of 2 M $H_2SO_4$. The absorbance (OD) was measured at 450 nm, and the absorbance at 570 nm was subtracted.

## Results

We tested 10 antibodies, originally developed against the SARS-CoV-1 S protein, for detection of the SARS-CoV-2 S protein by ELISA. From these, only two (AI334 and AQ806) bound in a concentration-dependent manner to the SARS-CoV-2 protein (Fig. 1). Two other antibodies, used as negative control (AH917 against the SARS-CoV-1 N protein and RB168 against an amoeba protein), also did not bind the S protein. AI334 and AQ806 antibodies have recently been shown to bind the RBD of the SARS-CoV-2 spike protein (Yuan *et al.*, 2020; Wrapp *et al.*, 2020a).

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## Conflict of interest

The authors declare no conflict of interest.

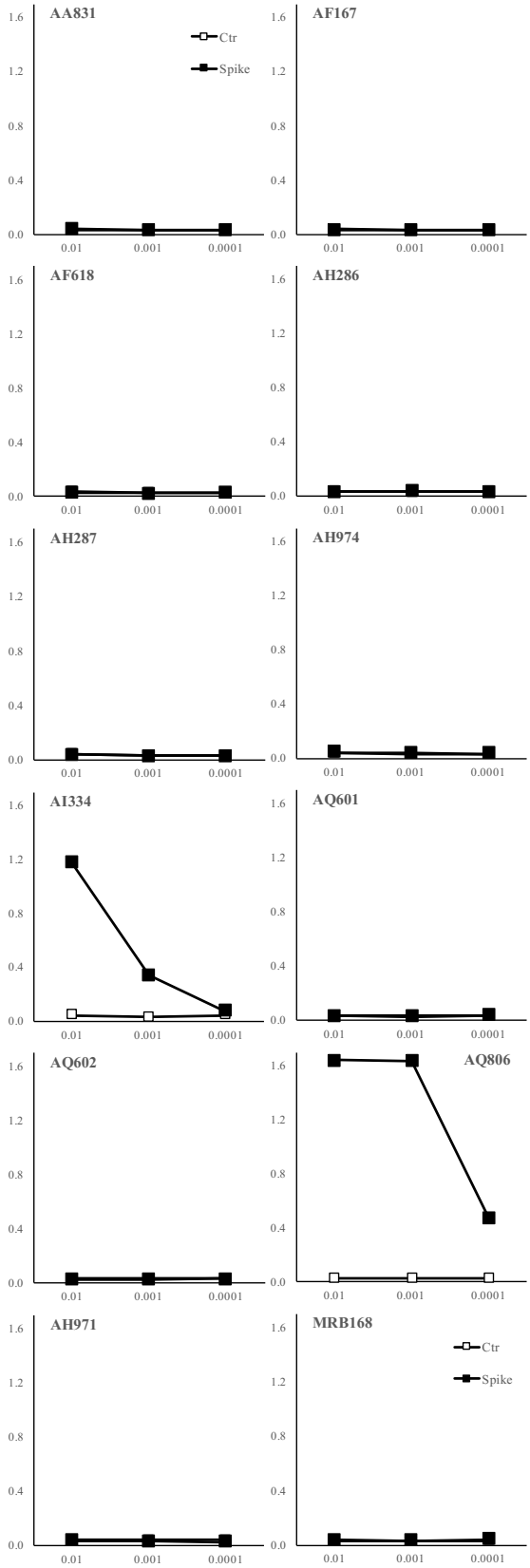


**Fig. 1.** Specific binding of AI334 and AQ806 antibodies to the SARS-CoV-2 S protein, as detected by ELISA. On the Y axis, ELISA signal (in arbitrary units). On the X axis, the antibody dilution (1:100, 1:1'000 and 1:10'000). 'Spike' refers to the binding to the spike S protein; 'Ctr' refers to the binding to biotinylated BSA.