

## Information transfer and recovery for the sense of touch

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## Abstract

12 Transformation of postsynaptic potentials (PSPs) into action potentials (APs) is the rate-limiting step of  
13 communication in neural networks. The efficiency of this intracellular information transfer also powerfully  
14 shapes stimulus representations in sensory cortices. Using whole-cell recordings and information-theoretic  
15 measures, we show herein that somatic PSPs accurately represent stimulus location on a trial-by-trial basis  
16 in single neurons even 4 synapses away from the sensory periphery in the whisker system. This information  
17 is largely lost during AP generation but can be rapidly (<20 ms) recovered using complementary  
18 information in local populations in a cell-type-specific manner. These results show that as sensory  
19 information is transferred from one neural locus to another, the circuits reconstruct the stimulus with high  
20 fidelity so that sensory representations of single neurons faithfully represent the stimulus in the periphery,  
21 but only in their PSPs, resulting in lossless information processing for the sense of touch in the primary  
22 somatosensory cortex.

23

## 24 **Introduction**

25 Neural information processing requires signal transformation every time the information is  
26 transferred from one neuron to another. This transformation is performed in postsynaptic neurons by  
27 integrating spatiotemporally distributed synaptic inputs and generating action potentials, which then  
28 propagate information across synaptically coupled neurons. For each processing step, how much  
29 information is retained, how much of it is transferred to a postsynaptic neuron, how much is lost, and  
30 whether local networks can fully recover the lost information during this intracellular sub-to-suprathreshold  
31 information transfer are questions that have yet to be answered. In an accompanying paper (Zeldenrust et  
32 al., 2020) we show on a single neuron level that how much information is lost during action potential  
33 generation depends on the cell-class. On a network level, the effectiveness of this input-to-spike operation  
34 depends on the connectivity, the code used between the sender (i.e. presynaptic neurons), and the receiver  
35 (i.e. postsynaptic neurons) as well as the noise characteristics of the channel. Since many of these are  
36 currently impossible to assess experimentally, the rules of information transfer in biological circuits, with  
37 the exception of cell-type-specific intracellular information transfer in single neurons as outlined in the  
38 accompanying article (Zeldenrust et al., 2020), are still largely unknown.

39 Sensory systems in particular offer unique opportunities to study information processing in neural  
40 circuits. If the primary function of a sensory circuit is to faithfully and reliably represent the environment  
41 (Azarfar et al., 2018; DeCharms and Zador, 2000; Diamond et al., 1999; Knudsen et al., 1987), a substantial  
42 part of the sensory information in the periphery should be represented throughout the sensory circuits in the  
43 form of neural signals. Sensory systems are commonly organized in the form of topographical maps, where  
44 sensory receptors in the periphery are represented by topographically organized groups of neurons along  
45 the sensory axis (Harding-Forrester and Feldman, 2018; Kole et al., 2018; Petersen, 2019). However, the  
46 functional role of these topographical maps for sensory processing is still not clear (Chklovskii and  
47 Koulakov, 2004; Diamond et al., 1999; Kaas, 1997; Weinberg, 1997). Understanding the mechanisms of

48 information processing, transfer and recovery is particularly important in sensory circuits, as the efficacy  
49 of signal transformation should determine the extent, speed, and accuracy of sensory representations.

50 Stimulating single neurons in the sensory and motor cortices can result in observable behavioral  
51 responses such as whisker movement (Brecht et al., 2004; Doron et al., 2014; Houweling and Brecht, 2008;  
52 Voigts et al., 2008). However, single neurons carry surprisingly little information in the rate and timing of  
53 their action potentials (Alenda et al., 2010; Panzeri et al., 2001; Petersen et al., 2001; Quiroga and  
54 Panzeri, 2009). Given that pooling information across simultaneously recorded neighboring neurons  
55 minimally contributes to the information carried in local populations because neighboring neurons carry  
56 largely redundant information (Petersen et al., 2002, 2001), the target postsynaptic neurons are likely to  
57 reconstruct the stimulus by spatiotemporal integration across behaviorally relevant spatial and temporal  
58 scales (Azarfar et al., 2018; Celikel and Sakmann, 2007).

59 Here, we performed intracellular recordings and used computational modeling to address the  
60 principles of information processing in the somatosensory cortex. Surprisingly (to us), we found that the  
61 sensory stimulus can be fully reconstructed with the information available in the *subthreshold* responses of  
62 single excitatory neurons (i.e. the recorded EPSPs in L2/3 neurons). Up to 90% of this information is lost  
63 during intracellular information transfer, i.e. when an *action potential* is generated from these subthreshold  
64 responses, in agreement with previous observations on the information content of action potentials in barrel  
65 cortical neurons (Alenda et al., 2010; Panzeri et al., 2001; Petersen et al., 2002). *In vivo* information loss is  
66 likely to exceed this value, due to background ongoing activity (Destexhe et al., 2003). Next, we assessed  
67 information recovery on the *population level* using an analysis based on bootstrapped groups of neurons  
68 recorded *in vitro*. We found that information lost during action potential generation can be fully recovered  
69 by as little as 100 neurons with a time resolution of 2-3 ms. Finally, we turned to a realistic and well-  
70 constrained simulation of a barrel column (Huang et al., 2020) to study the relation between encoding  
71 strategies in L4 and decoding strategies in L2/3 to determine the mechanisms of information recovery.

72 Comparing candidate encoding strategies in the L4 population, we found that a population rate code (using  
73 peri-stimulus time histograms obtained in the *in vivo* recordings) is unsuitable for information transfer in a  
74 cortical network because the trial-to-trial reliability is too low to fit the high information recovery that we  
75 found as in our experiments. Codes with higher trial-to-trial reliability in timing and rate perform  
76 substantially better, with optimal performance reached if neurons fire reliably across trials. In this case, the  
77 L4 activity can be fully decoded by small groups (~25 cells) of both excitatory and inhibitory neurons in  
78 L2/3 within ~20 ms after stimulus onset, and within a few ms after the first spike response. In summary,  
79 we show that intracellular information transfer is highly lossy, and thus potentially selective. However, by  
80 combining the limited but complementary information in the spike trains of L2/3 inhibitory and excitatory  
81 neurons, single neurons could fully reconstruct stimulus resulting in lossless representation of sensory  
82 information in their subthreshold responses.

## 83 **Results**

### 84 **L2/3 single cell responses to *in vivo* whisker stimulation**

85 We performed whole-cell current-clamp recordings of Layer (L) 2/3 pyramidal neurons in the juvenile rat  
86 primary somatosensory cortex, in the barrel cortical subregion under ketamine anesthesia. During these *in*  
87 *vivo* recordings, sensory stimulation was provided by direct stimulation of the principal and 1st order  
88 surround whiskers with a piezo stimulator in 2 directions (up-down: Fig. 1A). The cumulative synaptic  
89 input in response to these stimuli was quantified in properties of the somatic post-synaptic potential (PSP),  
90 i.e. the onset time, slope, and peak amplitude (Fig. 1B-D). Principal whisker stimulation-evoked PSPs  
91 exhibited the shortest latency, as well as the highest slope and amplitude, in comparison to PSPs evoked by  
92 the stimulation of surrounding whiskers, in agreement with previous observations (Brecht et al., 2003).  
93 Although the PSPs were highly reliable (PW: 99.8%, SW: 91.8% of trials evoked PSPs), action potentials  
94 (spikes) were sparse and unreliable, even after principal whisker deflections (PW: 6.2% (SD 8.6%) ; SW:  
95 1.7% (SD 2.9%) of trials included evoked APs).

### 96 **Properties of sub- & suprathreshold responses of L2/3 neurons to L4 stimulation**

97 Mutual information calculations require long sampling durations, which limits the possibilities for unbiased  
98 calculation of information processing with high-dimensional naturalistic stimuli *in vivo*. Therefore we  
99 performed acute slice experiments with simplified stimuli (Fig.1E-P). Bipolar electrodes in L4 were used  
100 to deliver square pulses with varying slopes as described before (Fig.1E-F; Huang et al., (2016); see  
101 Materials and Methods). Visualized L2/3 neurons were recorded in whole-cell current clamp configuration.  
102 PSP responses of L2/3 neurons systematically varied with the four stimulus patterns (Fig. 1G-J). Spike  
103 responses showed a similar dependence on the stimulus slope when averaged over multiple trials, with

104 delayed spike times, increased threshold and decreased spike probability for shallower slopes of stimulation  
105 (Fig. 1K-M).

106 The average properties shown above indicate that individual cells qualitatively correspond to  
107 whisker deflections mimicking spatial stimuli. However, during sensory processing, animals have to deduce  
108 object location from single trials, not from averages over many trials, which is only possible when trial-to-  
109 trial variability is low. Spikes exhibited a far greater temporal trial-to-trial variability than PSPs (Fig. 1N-  
110 P). PSP onset times showed an average progression with stimulus slope, with a small trial-to-trial  
111 variability (Fig. 1N, SD = 0.42-0.63 ms), whereas spike time variability was threefold higher (Fig. 1O, SD  
112 = 1.2-2.3 ms). Therefore, spike times could only to a very limited degree be predicted on the basis of PSP  
113 onset times, with spikes often occurring with significant and variable delays (Fig. 1P, 2.8-4.7 (SD 1.1-2.1)  
114 ms). Spike generation was also failure prone, with average spike failures rates up to 31.7% of the trials  
115 [Range : 0-85%]. While spike failures bear information in rate-based codes, in timing-based codes  
116 information is missing when the neuron fails to fire an action potential. To study the influence of trial-to-  
117 trial variability in timing and rate on stimulus information, we calculated Shannon's mutual information  
118 between the stimulus and several PSP and spike properties. This mutual information provides largely  
119 agnostic estimates of the transmitted information between stimulus and response.

120 **Information transmission between somatic PSPs and Spikes**

121 How much information does a somatosensory neuron carry about the sensory stimulus (S) and how much  
122 of this information does it transfer to its postsynaptic targets? Surprisingly, the information between a single  
123 somatic PSP and the stimulus contains the bulk (~95%,  $I(S;PSP)$  vs.  $H(S)$ , Fig. 2A) of the entropy of the  
124 sensory stimuli. The information between the onset time (81%, Fig. 2B), slope (8.4%, Fig. 2B) and  
125 amplitude (6.4%, Fig. 2B) of the PSP and the stimulus contribute largely independently to the total  
126 information content of a PSP (Fig. 2C). However most of this information is lost upon spike generation

127 (down to 24%,  $I(S;St,Vt)$ , Fig. 2A), where spike timing (St, 16%, Fig. 2D) and voltage threshold (Vt, 6.2%,  
128 Fig. 2D) carry most of the stimulus information contained in the spikes.

129 We can directly quantify how much of the information in the PSP is transferred to the spike (see  
130 Materials and Methods). Unsurprisingly, the total entropy in the PSP (i.e across onset, slope and amplitude  
131 together) exceeds the stimulus information multifold (6.4 bits, 3.2-fold, Fig. 2E). The transferred  
132 information from PSP to spike amounts to 22% and 15% of the PSP entropy for St and Vt respectively (Fig.  
133 2E, comparison of medians). However, most of this information is redundant, since the actual amount of  
134 stimulus information contained in a spike is much lower (0.41 bit, Fig. 2A). The individual PSP properties  
135 on the other hand contribute slightly synergistically to the timing information in the spike (Fig. 2F, 6.4%,  
136  $p < 10^{-5}$ ). Consequently, while a substantial amount of the information about the stimulus in the PSP is  
137 transferred to the spike, this information is insufficient to encode the present stimulus space at the single  
138 neuron level on a single neuron and single trial basis.

139 **Information recovery in local neural populations *in silico* and *in vitro***

140 If the PSP-to-spike transformation causes a dramatic drop in information about the stimulus carried in the  
141 neural activity, how can the somatic PSPs of L2/3 neurons carry near complete information (Fig. 2A)? Since  
142 these neurons are four synapses away from the sensory periphery, a recovery of information has to occur at  
143 the network level. Information recovery was analyzed both in an anatomically and physiologically well-  
144 constrained network model of a rat barrel column (Fig. 3) and on bootstrapped populations of the *in vitro*  
145 data (Fig. 4, see Materials and Methods and Huang et al., (2020)).

146 The model has anatomically correct numbers and laminar locations of major classes of inhibitory  
147 and excitatory neurons in L4 and L2/3 (Fig. 3A), single neuron dynamics based on experimental  
148 observations as well as statistically defined connectivity and synaptic transmission parameters. Stimulation  
149 was provided analogously to the *in vitro* stimulation in L4, using previously collected L4 peri-stimulus time

150 histograms (PSTH) of principal and surround whisker stimulation *in vivo* (Fig. 3B, L4 response to principal  
151 whisker in gray (Celikel et al., 2004). PSTHs of simulated L2/3 excitatory and inhibitory neurons  
152 correspond to experimentally observed ones under similar conditions (De Kock et al., (2007), Fig. 3B red  
153 and blue, respectively).

154 In the model, the timing of and information in PSPs and spikes closely matched the properties of  
155 the real neurons in biological networks (compare Fig 3 to Fig.1-2). In the simulations, the trial-to-trial  
156 variability in spike timing was substantially and significantly larger than the variability in PSP timing  
157 (Fig. 3C, compare with Fig. 1A). Stimulus information was nearly fully retained in the somatic PSPs of  
158 excitatory neurons (Fig. 3D, red, 88.8% of the stimulus entropy), yet reduced substantially (20.1%) during  
159 spike generation, similar to our observations in biological neurons (Fig. 2A). Interestingly, inhibitory  
160 neurons carried significantly less information in their PSPs (Fig. 3D, blue, 77.3%), but also exhibited less  
161 information loss during spike generation (43.6%).

162 In the bootstrapped data, the stimulus information could be almost fully recovered from populations  
163 of excitatory L2/3 neurons recorded *in vitro* (>81.1%, Fig. 4B). The amount of information recovered was  
164 substantially greater for decoding including timing (81.1%, in timing of the 1st spike, binned at 2ms, 100  
165 cells), than for rate based only decoding (50.5%, red vs. light red) and was largely independent of the  
166 population size (i.e. the MI saturates quickly as a function of population size). To avoid an overestimation  
167 of information from high-dimensional population data (due to the limited sampling bias), we first decoded  
168 the stimulus from single trial responses using a support vector machine (SVM) based decoder (Fig. 4A)  
169 before computing the MI (Ince et al., 2010b, 2010a; Quiroga and Panzeri, 2009). To verify that this  
170 method did not introduce a positive bias, we computed the information in response to an artificial  
171 uninformative stimulus set (same PSTH for all stimuli, independent Poisson spiking), which yielded near-  
172 zero MI values (Fig. 4C). The performance of the SVM provided significantly better results (correctly  
173 predicting 94% of the stimuli), than linear (79%) or quadratic (80%) decoders. However, better decoders

174 than the SVM may still exist, and the information calculated here therefore constitutes a lower bound on  
175 the available information in the population data. Also, it should be noted that the present timing code does  
176 not automatically include the rate code, since only the first spike is considered (see Materials and Methods).

177 **Contribution of timing/rate reliability for information recovery**

178 While a well-defined stimulus can be provided *in vitro*, the details of the L4 population activity cannot be  
179 controlled. From the perspective of information transmission in single trials, the most important property  
180 of the neural response is the reliability across trials. We utilized the barrel column *in silico* (Huang et al.,  
181 2020) to investigate the influence of the encoding strategy in L4 (reliability of spike timing and spike count)  
182 on the information transfer to L2/3.

183 We first considered three extreme cases of encoding in L4, with a more systematic exploration in  
184 the following section. In the first case ('Rate + Poisson'), stimuli are encoded only by the population PSTH,  
185 but spikes across trials and neurons are drawn with Poisson statistics. In the second case ('Rate + Trial  
186 Reliability'), stimuli are encoded by the population PSTH in L4, but also by the spike timing and count of  
187 its individual neurons (i.e. the spike trains were identical for each trial with the same stimulus conditions).  
188 Within the constraints of the experimentally observed PSTHs, these two cases constitute the lower and  
189 upper bound of trial-to-trial reliability in L4. In the third case ('No Rate + Trial Reliability'), the population  
190 PSTH carries no information about the stimulus, but all information about the stimulus is encoded in the  
191 spike timing and count of individual neurons. The latter case is added for comparison with the other two  
192 encoding paradigms. Here, the population PSTH does not distinguish between stimuli, which happens for  
193 instance for texture recognition tasks (Arabzadeh et al., 2005). These three cases are illustrated in the insets  
194 of Fig. 4D-F. More details regarding the construction of these cases are given in Materials and Methods.

195 In the 'Rate + Poisson' case, information transfer is overall low, with interneurons providing a  
196 superior readout of the information about the stimulus in L2/3 compared to excitatory neurons, both when

197 the information was decoded in rate and in timing (Fig 4D right, light and solid colors respectively,  
198 excitatory (red) vs. inhibitory (blue) neurons). While timing and rate codes are similarly efficient in  
199 interneurons, substantially larger populations of excitatory neurons are required to decode information in  
200 rate than in time. Given that the timing of the stimulus is only present on the population level in L4, the  
201 dominance of this temporal readout in L2/3 is remarkable. Assuming the emulated stimuli *in silico*  
202 approximate the stimuli *in vitro* with high accuracy (see Figs. 1 and 2), the ‘Rate + Poisson’ coding does  
203 not reflect the L4 encoding scheme in the present experiments.

204 In the ‘Rate + Trial Reliability’ case, the information transfer is overall substantially higher than in  
205 the ‘Rate+Poisson’ case (Fig. 4E right). In this condition the number of neurons required to recover the full  
206 stimulus information is the lowest (~25) of the three cases. This was expected, since in this case two sources  
207 of information - rate and timing - are used in the encoding of the stimulus. Remarkably, both cell-types and  
208 both decoding strategies yield very similar information values here, suggesting this encoding strategy is  
209 optimal for information transfer.

210 In the ‘No Rate + Trial Reliability’ case, the information transfer is intermediate between the two  
211 preceding cases. Here, the stimulus is only encoded by the responses of individual L4 neurons, not by the  
212 population PSTH. Interestingly, the opposite from the ‘Rate + Poisson’-case can be observed here in the  
213 decoding efficiency between L2/3 cell-types: contrary to the ‘Rate + Poisson’-case, where interneurons  
214 transfer more stimulus information, here excitatory neurons become substantially more efficient at  
215 representing information (compare Fig. 4D to 4F).

216 In summary, the availability of stimulus information in L2/3 spike trains is highly dependent on 1)  
217 the encoding properties of L4, 2) the decoding strategy in L2/3 and 3) the identity of the L2/3 neuronal  
218 populations (inhibitory or excitatory, for a summary see Table 1). Information about the stimulus in the  
219 spikes of L4 single units is best recovered by L2/3 excitatory neurons (Fig. 4F), given that there is a

220 reasonable trial-to-trial reliability. Next, we systematically modulated the information content in L4 spike  
221 trains to investigate the consequences for L2/3 information availability.

222 **Population and single unit information selectively influence inhibitory or excitatory cells**

223 While we only considered the extreme cases of L4 encoding above, realistic encoding will necessarily cover  
224 a range of cases between these extremes. Different stimuli will often, but not always, lead to different  
225 population PSTHs (but different surface textures may well lead to similar population PSTHs while finely  
226 modulating single unit responses (Arabzadeh et al., 2005). Conversely, even in cases where the population  
227 PSTH carries substantial information about the stimulus identity, spiking may well not be Poisson, but more  
228 reliable (especially at the response onset see e.g. (Amarasingham et al., 2006)).

229 We investigated the contributions from the population and the single unit separately. Information  
230 on the population level was represented as the average PSTH of the population (see Figure 6A1). Stimulus  
231 information was encoded in L4 spike trains either as timing or rate differences. Timing differences were  
232 implemented as shifts of the PSTHs ( $\Delta_T$ ), whereas firing rate differences were implemented as rate factors  
233 between the PSTHs ( $\Delta_C$ ). If  $\Delta_T = 0$  ms and  $\Delta_C = 1$ , then no information is contained in the population PSTH.  
234 Conversely, if  $\Delta_T = 4$  ms and  $\Delta_C = 4$ , the combined differences between the PSTHs are similar to the  
235 experimentally observed ones. These parameters allow us to study the susceptibility of L2/3 neurons to the  
236 different encoding strategies of L4 neurons (examples of spike patterns are shown in Fig. 5A1 above the  
237 PSTHs, 10 trials each).

238 For decoding using spike times, inhibitory neurons exhibited a substantially greater susceptibility  
239 to variations in the distinguishability of stimuli on the L4 population level compared to excitatory neurons  
240 (blue vs. red, Fig. 5A2). This was true for both variations in time ( $\Delta_T$ ) and rate ( $\Delta_C$ ) in L4. Similarly, for  
241 rate decoding, inhibitory neurons were more susceptible to changes in rate than excitatory neurons (Fig.

242 5A3). Timing had little effect on rate decoding, since this corresponds mostly to a shift in the analysis  
243 window (unrestricted here), with no change in rate information in L2/3.

244 Next, we considered the influence of various levels of single unit information on the information  
245 availability in L2/3. Here, the population PSTH is kept fixed, but the temporal and count reliability are  
246 varied on a neuron-to-neuron basis (see Fig. 5B1). The timing reliability was varied by introducing a  
247 temporal jitter to individual spikes across trials ( $SD_T$ ), while contracting spiking patterns to remain  
248 consistent with the population PSTH. Count reliability was varied selectively by a linear transition between  
249 a completely reliable and a Poisson model, while maintaining the population PSTHs. This was done by  
250 shifting spikes between neurons while maintaining the temporal variability across neurons.

251 For decoding using spike times, excitatory neurons showed much greater susceptibility to single  
252 unit differences in reliability in L4, both for rate and time (Fig. 5B2). Interestingly, this carried over to the  
253 rate decoding to an even greater degree, which may be the domain of action for inhibitory neurons (Fig.  
254 5B3).

255 In summary, L2/3 excitatory neurons are much more sensitive than L2/3 inhibitory neurons to the  
256 spike timing of L4 single neurons, whereas the information encoded by the population PSTH in L4 is carried  
257 mostly by inhibitory L2/3 neurons. Hence, we propose that the inhibitory and excitatory populations  
258 perform stimulus decoding in parallel, extracting stimulus information from distinct features in L4 activity  
259 (see Table 1). Together they have the ability to represent the entire information efficiently in small  
260 populations.

261 **Information recovery occurs rapidly within a few milliseconds**

262 Information processing in the sensory cortices is under severe temporal constraints, especially in S1, where  
263 the sensory input is tightly integrated with the motor output for the purpose of precise and adaptive whisking  
264 control (Li et al., 2015; Proville et al., 2014; Voigts et al., 2015, 2008). The state of processing at a given

265 time can be estimated by computing the mutual information over limited time windows, which  
266 progressively include a larger proportion of the neural response (Fig. 6A, all excitatory and inhibitory  
267 neurons separated for a single trial). In combination with varying the group size, we can thus obtain a  
268 ‘neurotemporal’ overview over the process of information availability in L2/3 as a function of neuronal  
269 class.

270 We consider three different encoding strategies by L4, ‘Rate + Poisson’, ‘Rate + Trial Reliability’  
271 and ‘No Rate + Trial Reliability’ as in the previous section (Fig. 6B-D). In the ‘Rate + Poisson’ case, the  
272 mutual information begins to increase with interneurons leading over excitatory neurons (Fig. 6B, left,  
273 group size = 10 cells) around 12-14ms after stimulus onset (in L4). The inhibitory neurons reach maximal  
274 stimulus information, and do not achieve full stimulus information. Groups of inhibitory neurons encode  
275 more information than excitatory neurons, independent of the time relative to the stimulus onset and almost  
276 independent of group size (Fig. 6B, right).

277 For the ‘Rate + Trial Reliability’ encoding condition in L4, the difference in the information content  
278 between cell types in L2/3 is small, with inhibitory neurons carrying slightly more information at early peri-  
279 stimulus times and across all group-sizes (Fig. 6C, right). The difference in onset timing renders the  
280 information content of the inhibitory neurons higher only during the initial 1-2ms after response onset, due  
281 to the earlier response times of the inhibitory neurons (Fig. 6C, left).

282 In the ‘No Rate + Trial Reliability’ condition, the times when the information content increases are  
283 very comparable for excitatory and inhibitory neurons. However, after a few milliseconds, excitatory  
284 neurons prevail over inhibitory neurons. This advantage is preserved over time, whereas the difference in  
285 information content as a function of group size is strongly reduced, with inhibitory neurons eventually  
286 catching up with excitatory neurons (Fig. 6D, right).

287 In summary, the representation of stimulus information in L2/3 is rapidly completed within only a  
288 few milliseconds (3-5) after response onset. Which neurons, i.e. excitatory or inhibitory, carry more

289 stimulus information is determined by the encoding strategy in L4 (corresponding potentially to different  
290 types of stimuli), but not much on the peri-stimulus time. As before, pure rate coding on the level of L4 is  
291 identified as an insufficient coding strategy, as it does not fit our experimental results of almost complete  
292 information recovery.

293 **Discussion**

294 We demonstrated that although the intracellular information transfer, i.e. the PSP-to-action potential  
295 transformation, results in a significant loss of information about the stimulus, local networks can overcome  
296 this loss by integrating information from a small, experimentally tractable, number of neurons. Therefore,  
297 the somatic PSPs received by a single cortical neuron contain nearly complete information about the  
298 stimulus, even several synapses away from the sensory periphery. The efficiency of such information  
299 recovery is determined by a conjunction between the encoding scheme, neuronal class and decoding  
300 strategy. Excitatory and inhibitory cells take complementary roles in carrying information in single unit or  
301 population activity, respectively.

302 **Contribution of temporal coding in somatosensory cortex**

303 Encoding information on short temporal scales can enrich the information content of neural activity  
304 relative to coarser average rates (Bialek et al., 1991; Bialek and Rieke, 1992). There has been a long  
305 discussion about whether the brain uses such a ‘spike code’ or ‘rate code’ (for a review, see Brette, 2015).  
306 It has been argued that since cortical networks are both noisy and very sensitive to perturbations, a rate  
307 code is the only way to perform reliable computations (London et al., 2010, but see Denève and Machens,  
308 2016). However, others have pointed to the presence of temporally encoded information in the  
309 somatosensory (Alenda et al., 2010; Panzeri and Diamond, 2010; Petersen et al., 2001) and other sensory  
310 cortices (Kayser et al., 2012, 2010). In particular, the timing of the first (few) spike(s) in response to a

311 stimulus conveys much of the information present in a spike train (Gollisch and Meister, 2008; Johansson  
312 and Birznieks, 2004). Consistently, we find that the majority of the information in the PSP is encoded in its  
313 timing. However, the timing of a spike in response to such a PSP is substantially more variable than the  
314 PSP timing, such that only a small proportion of the information in the PSP is transferred to the spike  
315 (Fig. 2). The amount of information loss could even be more substantial *in vivo* in the presence of  
316 background ongoing activity. On the population level, we find again that the temporal information is highly  
317 relevant during information recovery. In agreement with the previously observed importance of the first  
318 spikes, we find that the information content in the population asymptotes within 5 ms after the first spike  
319 in local populations, consistent with the time-scales of neuronal read-out in whisker cortex estimated before  
320 (5-8 ms, (Stüttgen and Schwarz, 2010)).

321 The temporal information described above can be fully characterized by single neuron variations  
322 in rate, and hence does not include higher order temporal codes, such as the pattern of inter-spike intervals.  
323 Due to the sparse response nature of supragranular excitatory neurons, such a fine-grained higher order  
324 temporal code could only exist in the inter-spike intervals of inhibitory neurons or spike-patterns across  
325 multiple (excitatory or inhibitory) neurons. The term temporal code is however still appropriate for our  
326 results, since the time scales of the response are not only reflecting dynamics in the stimulus, but correspond  
327 to intrinsic computations of the neural network (Nemenman et al., 2004).

328 For the present dataset, Shannon's mutual information was computed with responses aligned to the  
329 stimulus onset. Recent work by Panzeri and colleagues (Panzeri et al., 2010; Panzeri and Diamond, 2010)  
330 have pointed out that such a reference time is not necessarily available to a decoder in S1. How would a  
331 change to an internal reference time, such as the efference copy of the whisking signal (Crochet et al., 2011;  
332 Crochet and Petersen, 2006; Poulet and Petersen, 2008) or a population-based timing (e.g. the “Columnar  
333 Synchronous Response”, CSR, event defined by (Panzeri and Diamond, 2010)) affect the present results?  
334 Assuming that the population response can be approximated by a set of individually recorded neurons (as

335 in (Panzeri et al., 2010; Panzeri and Diamond, 2010)), the influence of such an intrinsic reference on our  
336 results would be only minor, since the relative timing - and thus the relative trial-to-trial variability in timing  
337 - would be the same as in the stimulus locked case. Hence, the information content would not be modified.  
338 If, on the other hand, synchronization between neural groups occurs, results could be significantly  
339 influenced, since then variability could be transferred from spikes to PSPs (in which case the alignment  
340 would be based on the near-synchronous CSR). According to Petersen and colleagues (Petersen et al.,  
341 2001), covariability, measured as noise correlation, was assessed to be ~0.1, and subsequent studies have  
342 found even lower values (Renart et al., 2010), suggesting that stimulus-independent synchronization is not  
343 substantial (note however the results of (Franke et al., 2016)

344 **High information availability and multiplexed codes**

345 To understand ‘how the brain works’, we need to understand what the neural computations are that  
346 make an animal interact with its environment, i.e. how neural activity is transformed from the low-level  
347 response, to perceptual input, to the high-level neural activity that generates behavior (Eliasmith and  
348 Anderson, 2002). For instance, perceptual invariance (an object can be recognized as one and the same  
349 under different circumstances) and selectivity (an object can be distinguished from other, similar objects)  
350 need to be explained by any consistent theory of perception (Seung and Yuste, 2012). A model of how  
351 increasingly abstract features can be recognized by neural networks along the sensory axis was already  
352 explained by for instance the perceptron-model (Rosenblatt, 1958; Seung and Yuste, 2012). When neurons  
353 in each processing layer respond to only a single, increasingly abstract, preferred feature, they disregard  
354 necessarily a lot of information. Therefore, on the *single-neuron level*, the transformation from input to  
355 output is expected to be very sparse, and ‘lossy’. However, whether on a *population level* it is necessary to  
356 be able to fully reconstruct the stimulus, remains an open question. We have shown here that the entire  
357 stimulus information is maintained in layer 2/3 of the barrel cortex and encoded by local populations in a

358 distributed fashion. This information can be recovered already on the basis of a small subset of neurons  
359 (~10-20, if single unit information is present) on short time-scales (~5ms relative to response onset),  
360 ensuring a lossless representation of the sensory world in real-time, i.e. before the next sensory information  
361 arrives from the periphery (in the case of active tactile exploration in freely behaving rodents, the inter-  
362 contact intervals are >30 ms (Voigts et al., 2015, 2008)). In different setups (Dagleish et al., 2020), in  
363 mouse visual cortex (Sriram et al., 2020) and in salamander retina (Marre et al., 2015), comparable values  
364 have been reported . This suggests that the full stimulus information is needed for the computations at  
365 several levels. Combined with the single neuron selectivity, our results suggest that this network performs  
366 a form of coordinate transformation (Denève and Pouget, 2003). However, what the exact nature of the  
367 computations of this and downstream networks is, remains an open question.

368 The neural activity of excitatory neurons in cortical layers 2/3 is generally considered to be sparser  
369 than in Layer 4 (see (Barth and Poulet, 2012) for a review, although the evidence is not yet fully conclusive).  
370 This sparsity has been linked to higher selectivity of encoding, in terms of fewer, more specific features  
371 represented per neuron. This increased selectivity could be the reason for the observed information loss  
372 during the transformation of PSPs to spikes. It has been argued that the sparsity of the transformation of  
373 presynaptic spike trains to PSPs to postsynaptic spikes is the result of optimal non-linear processing: only  
374 redundant information, that has been gained before and can be predicted from previous activity, is  
375 discarded, and postsynaptic neurons only respond to ‘new’ information (Denève, 2008; Ujfalussy et al.,  
376 2015). Our result that the postsynaptic membrane potential still contains the full stimulus information, is in  
377 agreement with this argument, and the observation that most information is contained in the first spikes  
378 mentioned before could also be explained this way. However, whether the information that is lost in the  
379 spike-generating process is truly ‘discarded’ information, or, contrarily, redundant information, depends on  
380 the presumed decoding of the neuron: which information is redundant or essential depends on the message  
381 that needs to be conveyed.

382        The distributed persistence of complete stimulus information could provide a practical solution to  
383        one of the classical dilemmas of neural encoding: the compatibility between a specific feature and the  
384        context of the entire stimulus space. Concretely, a readout neuron in L2/3 may have privileged access to L4  
385        neurons selective for one type of feature, with in addition access to a wide range of inputs from a random  
386        subset of the population. It could thus act as a comparator and evaluate the dominant feature in relation to  
387        a representation of the entire stimulus. This becomes especially relevant in the case of multiple concurrent  
388        stimulations on different whiskers, corresponding to the natural situation an animal is exposed to during  
389        active exploration (Voigts et al., 2015). In this case, multiple signals (i.e. the signals from multiple whiskers,  
390        that carry different spatial and temporal information) have to be processed by a single population of  
391        neurons. If this population can be separated into independent-subpopulations, this implies that a population  
392        consists of multiple channels (in the information-theory sense), but if this is not the case, multiple signals  
393        are coded by a single population, so the code becomes multiplexed: a single channel (population) carries  
394        complementary information through different codes. The observation that multiple subsets of neurons carry  
395        complete stimulus information and the observation that spike timing and firing rate of the same population  
396        (channel) can contain independent information about the stimulus hints at such multiplexing ((Panzeri et  
397        al., 2001; Quian Quiroga and Panzeri, 2009), for a review, see (Panzeri et al., 2010)). Our finding that  
398        different postsynaptic populations can decode the timing-encoded and rate-encoded information shows that  
399        the information from both coding schemes (rate and timing by inhibitory and excitatory neurons) of such  
400        multiplexed encoded information can also be used by the brain for further processing in later stages.  
401        Information in inhibitory populations can then be forwarded by for instance disynaptic (dis)inhibition and  
402        the modulation of firing rates or spike probabilities of excitatory populations. Multiplexed codes have been  
403        discussed recently in the context of local field oscillations (Alenda et al., 2010), and the presence of  
404        selective and general information as described herein may provide an additional example of multiplexing  
405        (Fig. 7).

406        Given the information content across the excitatory and inhibitory neural populations calculated  
407 herein, we speculate that distinct tactile features are encoded by rate and timing of spiking during  
408 information encoding and decoded by excitatory and inhibitory neurons separately (Fig. 7A). If an animal  
409 were to use its whiskers to locate a tactile target in space for example (Celikel and Sakmann, 2007; Peron  
410 et al., 2015), this model predicts that inhibitory neurons would carry the largest amount of information  
411 during the first contact as the animal detects the edge of the tactile target. Similarly, at the detection of a  
412 contact during passive whisking (Clem et al., 2008), inhibitory neurons would preferentially respond to the  
413 onset of touch, serving as an edge detector. The information content of different signals within the L4-to-  
414 L2/3 channel is temporally constrained as the animal continues to explore its immediate environment, and  
415 makes additional whisker contacts with the tactile target (Fig. 7B; (Voigts et al., 2015, 2008)), presumably  
416 to predict object distance and extract additional surface feature information about the target. With the  
417 change of whisking pattern, the sensory history and the statistics of the local network activity, relative  
418 information in the excitatory population will eventually dominate the neural representation of touch (Fig.  
419 7C).

420        How to convey stimulus information both lossless and efficiently, and how this depends on physical  
421 properties such as network connectivity and node (neuron) properties, is an important open question in  
422 network science, and neuroscience specifically (Maheswaranathan et al., 2018; Mastrogiuseppe and  
423 Ostojic, 2017), in which computational models like the one we present here (Huang et al., 2020) play an  
424 invaluable role. Recently, it has been shown that there is a trade-off between the sparsity and the amount  
425 of recovered information in neural coding (Billings et al., 2014): lossless coding is only possible if the  
426 connectivity in the network is not too sparse. Specifically, the authors showed that optimal connectivity  
427 included only a few excitatory synapses and strong inhibition. Moreover, activity-dependent thresholds

428 appear to play an invaluable role in such efficient information transmission (Billings et al., 2014; Huang et  
429 al., 2016).

430 For future *in vivo* studies, an important question will be, whether complete information  
431 representation persists if larger stimulus sets/spaces are considered, since it is expected that the  
432 dimensionality of the response, and hence the number of neurons needed for complete information  
433 recovery, depends on the complexity of the stimulus (Gao and Ganguli, 2015). Due to the requirements of  
434 accurate estimation of mutual information, we had to restrict the stimulus space to four stimuli in the context  
435 of whole-cell recordings (leading to ~300 trials per recorded cell). Note, however, that even under these  
436 conditions, trial-to-trial variability could have prevailed and prevented complete stimulus reconstruction on  
437 the single neuron and population level. Moreover, the present results can only provide a lower bound on  
438 the available information, since not all possible codes were explored and the decoding step between  
439 stimulus and response renders all results lower bounds (Quiroga and Panzeri, 2009). In contrast to a  
440 previous study in the auditory cortex (Ince et al., 2013), we find that more complex decoding methods  
441 provide an improved decoding quality and hence more mutual information. Concretely, support vector  
442 machine decoding with radial basis functions provided superior performance (94%) than either diagonal  
443 linear (77%), linear (79%), or quadratic (80%) decoders. In order for the neural system to achieve this  
444 quality of decoding, it would, however, need to have readout mechanisms which use decoding strategies  
445 beyond linear or quadratic combinations.

446 **Predictions for cell-type specific coding strategies**

447 The cortical population of neurons is composed of various cell-types, which differ in their morphology,  
448 location and physiology (De Kock et al., 2007; Narayanan et al., 2015; Oberlaender et al., 2012; Staiger et  
449 al., 2015). These differences suggest distinct roles in information processing, some of which have recently

450 been elegantly elucidated (Ko et al., 2011). Coarsely, on the level of their firing patterns, inhibitory neurons  
451 can be distinguished from excitatory neurons, by more dense responses, based on a greater convergence of  
452 connections (reviewed in Harris and Mrsic-Flogel (2013)). The connectivity in the present model was set  
453 in precise accordance with the latest results from the literature from identified, pairwise recordings (see  
454 Huang et al. (2020) for detailed references), and consequently recreates these differences in firing behavior.  
455 Going beyond previous work, we find the coding balance to lean to either cell class, depending on the  
456 encoding strategy used in L4.

457 We explored these encoding strategies in L4, finding that excitatory neurons more effectively  
458 convey information encoded in L4 single units, requiring a level of reliability in L4 beyond Poisson-spiking  
459 (Figure 6B). Conversely, inhibitory neurons are more effective in carrying L4 population rate information  
460 (Figure 6A). Hence, together, excitatory and inhibitory neurons make effective use of the combined  
461 information in population rate and single unit responses in L4.

462 The L4 encoding is likely to depend on the stimulus condition: Many stimuli will induce time-  
463 varying population rates, which distinguish them from other stimuli. However, exceptions exist, such as the  
464 comparison of similar textures (Arabzadeh et al., 2005), which have only small differences in population  
465 rate, and differ more in their fine-structure. On the other hand, temporally structured inputs (e.g. many  
466 natural stimuli) lead to stronger time locking between neurons in L4 (Amarasingham et al., 2011; Litwin-  
467 Kumar and Doiron, 2012). Based on our results, we suggest that excitatory and inhibitory neurons might  
468 focus on distinct individual and population information to optimize the availability of stimulus information  
469 in local networks. Since long-range projections of inhibitory neurons are rare (Thomson and Lamy, 2007),  
470 the information content in the spiking of the inhibitory neurons is likely to be most relevant for local  
471 processing. Testing this hypothesis will not be trivial, since the inhibitory neurons cannot be removed from  
472 the network without influencing the overall network dynamics. Nonetheless, transient optogenetic  
473 modulation of the rate and timing of select inhibitory neurons' activity while studying neural encoding of

474 stimuli in the rest of the network will help to answer the question about which inhibitory neurons contribute  
475 more to the transfer and recovery of information within a column (i.e. local) and across columnar networks.

476

477

478 In summary, the results presented here suggest that single neurons are efficient real-time encoders  
479 of stimuli even several synapses away from the sensory periphery, but intracellular information transfer  
480 results in a substantial loss in the information transmitted to the postsynaptic neurons. The lost information  
481 can be recovered rapidly, i.e. within 20 ms, by comparatively small numbers of neurons in local populations,  
482 so that lossless information transfer along the sensory axis is ensured. The information recovery depends  
483 critically on the type of the neuron as well as the coding properties of both the presynaptic and postsynaptic  
484 pools of neurons, such that excitatory and inhibitory populations process complementary information about  
485 the stimulus in the somatosensory cortex.

486

## 487 Materials and Methods

### 488 Experimental procedures

489 Rats from either sex were used according to the Guidelines of National Institutes of Health and were  
490 approved by the local Institutional Animal Care and Use Committee. All data can be found in this online  
491 repository: <https://doi.org/10.34973/59my-jm24>, the relevant code can be found here:  
492 [https://github.com/DepartmentofNeurophysiology/Information-transfer-and-recovery-for-sense-of-touch-  
493 code-for-figures](https://github.com/DepartmentofNeurophysiology/Information-transfer-and-recovery-for-sense-of-touch-code-for-figures)

### 494 In vitro recordings

495 In vitro whole-cell current-clamp recordings were performed in acutely prepared slices of the barrel cortex  
496 between P18-21, after maturation of evoked neurotransmitter release (Martens et al., 2015) as described  
497 before (Allen et al., 2003; Celikel et al., 2004; Clem et al., 2008). Oblique thalamocortical slices (300 mm,  
498 (Finnerty and Connors, 2000)) were cut 45° from the midsagittal plane in chilled low-calcium, low-sodium  
499 Ringer's solution (in mM; sucrose, 250; KCl, 2.5; MgSO<sub>4</sub>.7H<sub>2</sub>O, 4; NaH<sub>2</sub>PO<sub>4</sub>.H<sub>2</sub>O, 1; HEPES, 15; D-(+)-  
500 glucose, 11; CaCl<sub>2</sub>, 0.1). Slices were first incubated at 37°C for 45 minutes and were subsequently kept in  
501 room temperature in carbonated (95% O<sub>2</sub>/5% CO<sub>2</sub>) bath solution (pH 7.4, normal Ringer's solution: in  
502 mM, NaCl, 119; KCl, 2.5; MgSO<sub>4</sub>, 1.3; NaH<sub>2</sub>PO<sub>4</sub>, 1; NaHCO<sub>3</sub>, 26.3; D-(+)-glucose, 11; CaCl<sub>2</sub>, 2.5).  
503 Visualized whole-cell recordings were performed using an Axoclamp-2B amplifier under IR-DIC  
504 illumination. A custom-made tungsten bipolar extracellular stimulation electrode (inter-tip distance 150  
505 micrometer) was placed in the lower half of a L4 barrel. Stimulation protocol was as described before  
506 (Huang et al., 2016). In short, 10 ms long current pulses were delivered using a bipolar electrode located in  
507 the lower half of a mystacial whisker's barrel. The pulses were square and had equal maximal amplitude  
508 although the rising phase of the stimulus had different slopes. It took 0,2,4 or 6 ms for the pulse to reach

509 the maximum amplitude for stimulus (S)1, S2, S3 and S4, respectively. All intracellular recordings (pipette  
510 resistance 3-4 M $\Omega$ ) were performed in L2/3, orthogonal to the stimulation electrode within 150-300  $\mu$ m  
511 of the cortical surface. The internal solution (pH 7.25) consisted of, in mM, potassium gluconate, 116; KCl,  
512 6; NaCl, 2; HEPES, 20 mM; EGTA, 0.5; MgATP, 4; NaGTP, 0.3. For whole cell recordings, putative  
513 excitatory cells were selected based on pyramidal shaped somata, apical dendrites and distal tuft orientation,  
514 and regular pattern of spiking to somatic current injections (500 ms; data not shown). Data was low-pass  
515 filtered (2 kHz), digitized at 5 kHz using a 12-bit National Instruments data acquisition board and acquired  
516 using Strathclyde Electrophysiology Suite for offline data analysis.

517 **In vivo recordings**

518 In vivo whole-cell current-clamp recordings were performed under ketamine/xylazine anesthesia at P28-  
519 30. Anesthesia was induced using 100 mg/kg (ketamine) and 10 mg/kg (xylazine) mixture and maintained  
520 with intraperitoneal ketamine-only injections (20% of the initial dose) as necessary. Upon complete loss  
521 of facial and hind-limb motor reflexes, the skull was exposed. A head-bolt was fixed posterior to lambda  
522 using cyanoacrylate and was used to immobilize the animal during experiments.

523 The surface over the primary somatosensory cortex (from Bregma, -0.5mm to -2.5mm, from Midline -  
524 2.5mm to -4.5mm was thinned using a dental drill. The surface was kept moist with a thin layer of low-  
525 viscosity mineral oil to maintain the transparency of the thinned skull. Cortical representation of the D2  
526 whisker was localized in the contralateral hemisphere using intrinsic optical imaging as described before  
527 (Stewart et al., 2013) while deflecting individual whiskers using piezoelectric actuators as described  
528 elsewhere (Celikel et al., 2004). The skull above the center of mass of the functional whisker representation  
529 was punctured using a 28 gauge needle to allow patch electrodes to access the cortical region of interest.  
530 All electrode penetrations were perpendicular to the cortical surface. In vivo whole-cell recordings were  
531 performed as described before (Margrie et al., 2002) with recording electrodes (6-7 M $\Omega$ ) filled with the

532 same intracellular solution used in slice experiments. Two different whisker deflection protocols were  
533 used: During optical mapping experiments single whiskers were deflected along the dorsoventral axis at 5  
534 Hz with 8° deflections for 20 times with an inter-trial interval of 20 sec (Stewart et al., 2013). During  
535 electrophysiological recordings single dorsoventral whisker deflections were delivered at 0.2 Hz for 200  
536 times. In each trial 4° whisker deflections were delivered at 10 Hz for 1s. Throughout the experiment the  
537 animal's core body temperature was maintained at  $36.5 \pm 0.5^\circ\text{C}$ .

538 **Data analysis**

539 All analyses were performed off-line in Matlab (Mathworks, Inc), the code for the figures can be found  
540 online: <https://github.com/DepartmentofNeurophysiology/Information-transfer-and-recovery-for-sense->  
541 of-touch-code-for-figures. Raw voltage traces were smoothed using running window averaging (1ms  
542 window size) and the following variables were calculated for all evoked responses: Onset time (Ot, in ms):  
543 Latency of the postsynaptic potential (PSP) onset in respect to onset of the stimulus; Rise time (Rt, in ms):  
544 Time it takes for the membrane to reach 90% of the PSP amplitude relative to the onset of PSP; PSP slope  
545 (Sl, in mV/ms) between 10-90% of the PSP amplitude and amplitude of the EPSP (Amp, in mV). If the  
546 trial included an action potential, the peak of the EPSP was set to the spike threshold (Vt). The spike  
547 threshold was defined as the membrane potential value at which the second derivative of the membrane  
548 potential reached a maximum as described before (Wilent and Contreras, 2004). In slice recordings, resting  
549 membrane potential (Vm, in mV) was calculated as the average membrane potential in a 40 ms time window  
550 prior to the stimulus onset. For *in vivo* recordings the same time window was used but the sweep was  
551 included in the data analysis only if the variance of the membrane potential was  $< 0.5\text{mV}$  during the time  
552 window. For those sweeps in which a spike was observed, the spike threshold and spike latency (St) were  
553 also calculated.

554 **Mutual information analysis for single neurons.**

555 Only cells with more than 250 acceptable sweeps (summed across all stimulus conditions) were  
556 used to perform Shannon information analysis. The mutual information (MI) between any two  
557 variables  $S, R$  can be calculated as

$$558 \quad I(S; R) = H(R) - H(R | S) , \quad (1)$$

559 in which  $H$  is the entropy of a given variable  $R$ :

$$560 \quad H(R) = - \sum_{i=1}^n p(r_i) \log_2(p(r_i)) \quad (2)$$

561 and  $H(R|S)$  is defined as

$$562 \quad H(R | S) = - \sum_{i=1}^n p(s_i) \sum_r P(r | s) \log_2 p(r | s) \quad (3)$$

563 where  $i$  ranges over the stimulus/response types. Note that the stimulus entropy shows a small variability  
564 due to rejected trials. Similarly, the mutual information between one variable  $S$  and multiple  $\mathbf{R}$  (joint mutual  
565 information) can also be calculated using equation (1). In this case, the synergistic effect of  $\mathbf{R}$  can be  
566 expressed as the difference between the linear sum of the mutual information between  $S$  and each individual  
567  $R$  and the joint information  $I(S; \mathbf{R})$ :

$$568 \quad \text{Syn}(S; \mathbf{R}) = I(S; \mathbf{R}) - \sum_{i=1}^n I(S; R_i) \quad (4)$$

569 Information calculations were performed using the Information Breakdown toolbox (Magri et al., 2009) in  
570 Matlab (Mathworks. Inc). In short, each variable was first digitized using the equal space ('eqspace') binning  
571 method with 7 bins. The effect of different binning methods as well as the number of bins on MI values are  
572 also explored (Fig. S1). In the analysis based on the 'eqpop' binning method, the size of individual bins  
573 was modified so that a roughly equal number of observations was placed in each bin, instead of keeping  
574 the size of individual bins constant. Because in most trials only one spike was observed, only the first spike  
575 was considered when calculating the information in  $S$ . Thus, the spike latency  $S$  can be digitized to a  
576 single word, which has (number of bin + 1) possible outcomes, instead of a binary list which could have

577 2^(number of bin) possible values. Shuffle correction combined with Panzeri-Treves (Panzeri and Treves,  
578 1996) bias correction was used to perform all information calculations for neural recordings (note that  
579 shuffle corrections can introduce a small source of variability, which can be seen for instance in comparing  
580 I(S,PSP) in Figure 2A with and H(S) or with the joint MI in 2C, or that can lead to an error bar above the  
581 stimulus entropy or below 0). The performance of the algorithm was evaluated by randomly selecting a  
582 subset of trials to calculate the mutual information (I(S;PSP), I(PSP;Vt) and I(PSP;St)), and subsequently  
583 checking the number of trials (Ns) needed for the calculated information values to reach asymptote (Fig.  
584 S2). When the ‘eqspace’ binning method was used, all information values reached asymptote after Ns > 70,  
585 well below the average Ns in the present data set (124±33.2 (range: 78-220) stimulus repetitions per  
586 stimulus).

587 **Calculation of minimum observation size:**

588 An essential step in the information calculation method listed above is the estimation of the stimulus-  
589 response probability distributions from the experimental data. Following Panzeri and colleagues (Ince et  
590 al., 2010b) we calculated the number of experimental trials per stimulus condition, Ns, to be ~32 times  
591 larger than the number of possible response pattern, R, to get an accurate estimation ( $Ns/R \approx 32$ ). This also  
592 means that to accurately estimate information between the subthreshold responses (Am, S1, Ot, all binned  
593 to 7 bins) and the stimulus,  $32 \times 7 \times 7 \times 7 = 10976$  trials/ stimulus =91h continuous recordings will be needed.  
594 Given technical infeasibility of maintaining whole cell access for the designated period we performed bias  
595 corrections to account for the upward bias in information estimation with limited sample sizes (see (Ince et  
596 al., 2010b) and (Victor, 2009) for further discussion). Methods like quadratic extrapolation (QE), Panzeri-  
597 Treves (PT) correction (Panzeri and Treves, 1996) and Nemenman-Shafee-Bialek (NSB) t experiments and  
598 94±25.6 (range, 60-146) trials/stimulus for the *in vivo* whole-cell recordings.

599 **Mutual information analysis for multiple neurons**

600 For multi-neuron MI analysis we followed the approach to first decode and then estimate the MI between  
601 stimuli and the confusion matrix of the decode (Ince et al., 2010b; Panzeri and Diamond, 2010; Quian  
602 Quiroga and Panzeri, 2009) using support vector machine (SVM) in MATLAB with radial basis functions  
603 as the kernel transform. We utilized 90/10% cross validation during decoding to obtain an estimate of the  
604 generalized performance of the decoder. SVM decoding outperformed other decoders with an average  
605 performance of 94%, compared with some other decoders (diagonal linear (77%), linear (79%), quadratic  
606 (80%)). The use of an intermediate decoder ensured that the calculation was bias free (given that we observe  
607 the correct value of 0 bits for an uninformative set of stimuli, with otherwise very similar properties (see  
608 Fig. 3G)), but came at the expense of lower bound in MI estimates since a (potentially existing) better  
609 decoder would improve the MI.

610 For the in-vitro recordings we first had to generate bootstrapped populations of sufficient size to perform  
611 the population MI calculations. In order to preserve the within-cell variability of responses across stimulus  
612 and trials, we only drew bootstrap samples from the trials of each cell independently. As in the simulations  
613 we drew 100 samples of groups of each population size. Curves in Fig. 4 display averages over these  
614 samples.

615 **Network Simulations**

616 The reconstruction of information in the neural network was performed in an *in silico* model of the barrel  
617 cortex.

618 **Neural network**

619 The model included a realistic account of the number of (Izhikevich, 2004, 2003) neurons and connectivity  
620 (Supplemental Table 1) within a barrel column for Layers 2/3, with inputs arriving from the L4, mimicking  
621 the conditions in the *in vitro/in vivo* experiments. For more details on the network model, see (Huang et al.,  
622 2020).

623 Synaptic currents in this network were modeled by a double-exponential function. Parameters of those  
624 functions (peak amplitude, rise time, half width, and pair-pulse ratio) were adjusted to match experimentally  
625 measured PSPs in barrel cortex (Supplemental Table 1; see Thomson and Lamy (2007) for a review). The  
626 onset latency was calculated from the distance between cell pairs; the conduction velocity of action potential  
627 was set to 190 $\mu$ m/ms.

628 Differences in activation state of cortex were included in the model by setting the common initial voltage  
629 and the equilibrium potential  $vr$  of all cells to -80, -70, or -60mV in a third of the trials, thus accounting for  
630 potential up- and down-states as well as an intermediate state.

631 **Synaptic input from layer 4**

632 Layer 4 stimulation was provided in the model based on population PSTHs collected extracellularly in  
633 anesthetized animals *in vivo* (Celikel et al., 2004). We used PSTHs of principal and 1st order surround  
634 whisker stimulation, as well as two linear interpolations between the two, yielding 4 stimuli with 2 bits total  
635 entropy, matching the numbers in the *in vitro* experiments. The PSTHs only specified the population firing  
636 rate in L4. We further explored population coding properties, by modifying the variability of spike timing  
637 across trials. If response times and spike counts were conserved across multiple trials, spike timing and  
638 counts within and between neurons start to carry additional information.

639 In the ‘Rate + Poisson’ condition, we assumed no trial-to-trial reliability beyond that given by the  
640 PSTH. Spike times were drawn based on Poisson statistics for each time with the PSTH modulating the  
641 firing rate (see Fig. 4D left). This condition forms a lower bound on the transferred information between  
642 L4 and L2/3, under the experimental constraints on the model. On the other extreme, in the ‘Rate + Trial  
643 Reliability’ condition, the PSTHs varied as before, but in addition neurons emitted the same sequence of  
644 spikes for every trial, preserving timing and count perfectly. This condition forms an upper bound on the  
645 information transfer, since within the experimental constraints no additional variability is introduced, which  
646 would reduce the mutual information. Finally, we consider the ‘No Rate + Trial Reliability’ condition,  
647 where the population PSTHs are uninformative across stimuli, and stimulus information is only contained  
648 in the spike trains of individual neurons. This case is a reference for other stimulus scenarios, where the  
649 PSTH may not vary much (e.g. texture-type stimuli), and individual timing becomes more important.

650 We also explored conditions between these extremes (Fig. 5), where the information in population  
651 or single neuron response was systematically varied. For the case of the population response we varied the  
652 different in time and firing rate of the PSTHs for different stimuli (Fig. 5A). Time differences were  
653 implemented by simply shifting the entire PSTH in time (tested shifts: [0,1,2,3] ms per stimulus, i.e.  
654 maximum shift was 9 ms). Rate differences were implemented as the fraction between the maximal and the  
655 minimal stimulus (tested fractions were [1,2,3,4], where e.g. 4 corresponds to the weakest stimulus being  
656 25% of the strongest stimulus at the peak of the PSTH). The case of time shift 0 ms and rate fraction 1 is  
657 uninformative on the level of population rate. Single neuron reliability in this case was chosen as a medium  
658 level of single unit reliability ( $SD_T = 3$  ms,  $SD_C = 20\%$ ). Single neuron reliability in response was also  
659 explored in timing and rate (Fig. 5B). Starting from perfect timing and rate, we degraded the information  
660 extractable from single neurons, by introducing timing variability (spike times were shifted by Gaussian-  
661 distributed noise with standard deviation  $SD_T$ ) and rate/count variability (spikes were deleted or added, by

662 linearly mixing between Poisson and perfectly reliable spiking, with mixing parameter  $SD_C$ , denoted as %  
663 in the figure). For both procedures, the modifications were performed while keeping the population PSTH  
664 approximately unchanged, i.e. for timing the overall timing distribution was contracted to keep the original  
665 PSTH, and for rate, spikes were shifted between neurons, rather than only removed from individual neurons.

666 These independent variations of population and single unit responses allowed us to separate the  
667 contribution of these two information sources to the information available in groups of L2/3 excitatory and  
668 inhibitory neurons (see Results).

669 **Abbreviations**

670	AP	: Action potential / spike	PSP	: Postsynaptic potential
671	PW	: Principal whisker	SW	: Surround whisker
672	H	: Entropy	I	: (Mutual) Information
673	S	: Stimulus	PSTH	: Peristimulus time histogram
674	$\Delta_T$	: Variation in spike timing	$\Delta_C$	: Variation in spike rate
675	L	: (Cortical) layer	CSR	: Columnar Synchronous Response
676				

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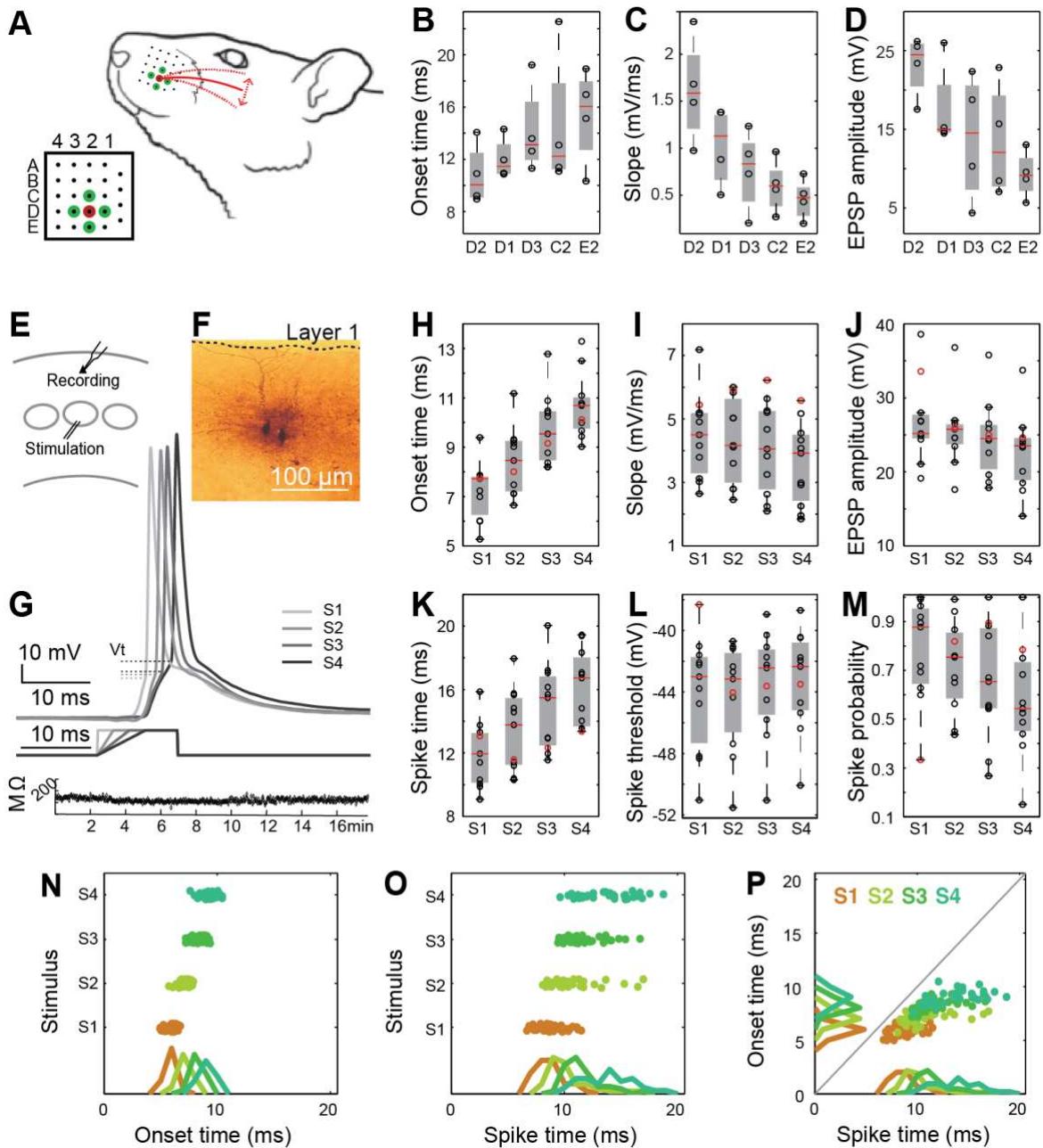
685 **Tables**

686 **Table 1: Summary of information recovery results depending on L4 encoding and L2/3 decoding schemes.**

L4 encoding→	population level		single neuron level	
L2/3 decoding↓	timing ( $\Delta_T > 0$ , $\Delta_C = 1$ )	rate ( $\Delta_T = 0$ , $\Delta_C > 1$ )	timing reliability ( $SD_T > 0$ , shift=0)	count reliability ( $SD_T = 0$ , shift>0)
<b>spike times</b>	inhibitory neurons more susceptible	inhibitory neurons more susceptible	excitatory neurons more susceptible	excitatory neurons more susceptible
<b>rate</b>	no effect	inhibitory neurons more susceptible	excitatory neurons more susceptible	excitatory neurons more susceptible

687

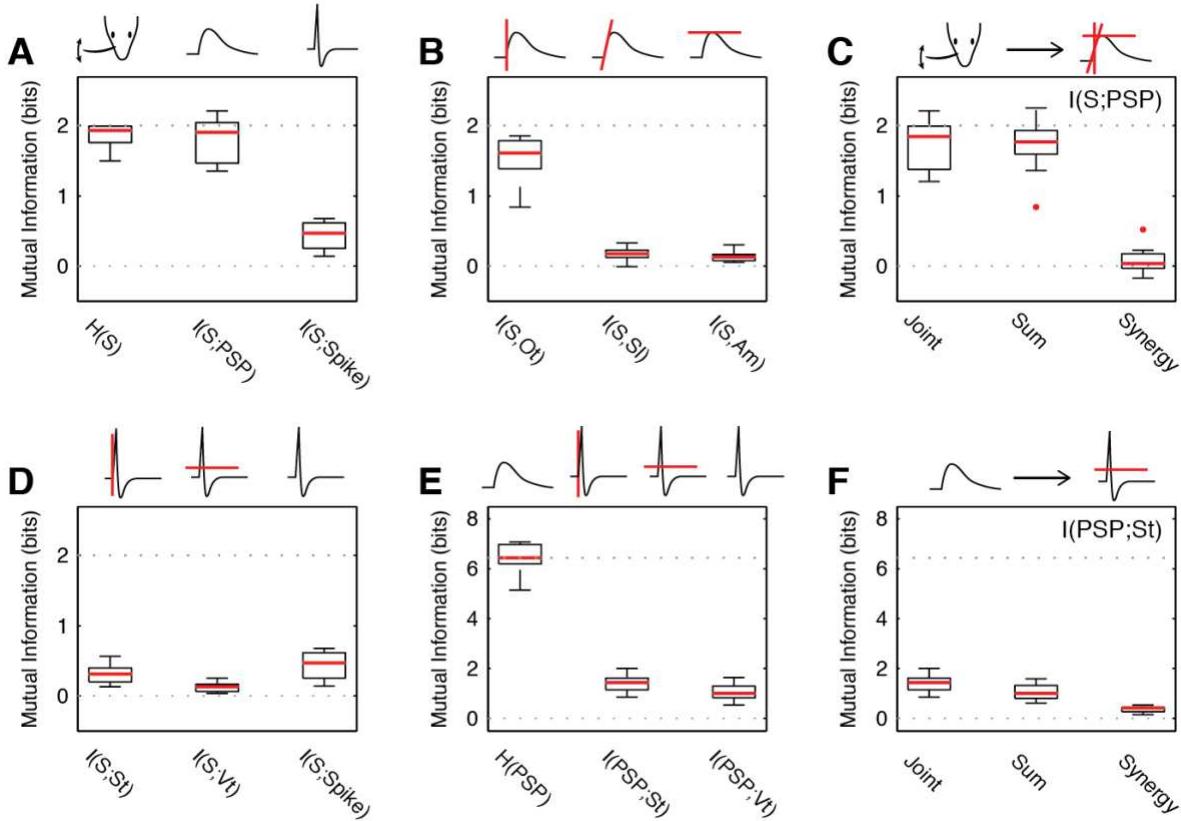
688 **Figures**



690 **Figure 1. *In vivo* and *in vitro* stimulus representation in single L2/3 somatosensory cortical neurons**

691 **(A)** We deflected whisker D2 and its first order neighbors (D1,D3,C2,E2) individually to determine the  
692 spatial encoding properties of cortical L2/3 pyramidal neurons in the D2 barrel under anesthesia using  
693 whole cell current clamp recordings. **(B-D)** EPSP response to *in vivo* stimulation. Analysis of the EPSP  
694 parameters showed that principal whisker stimulation was correlated with earlier onset times **(B)**, larger  
695 slopes **(C)** and larger amplitudes **(D)** compared to the surround whiskers. Onset time was described as the  
696 latency between stimulus onset and the time it takes for the membrane to reach 10% of the peak somatic  
697 EPSP amplitude. The EPSP slope was calculated to be between 10-90% of the somatic EPSP. The  
698 amplitude was measured at the peak. All measurements were performed on monosynaptic EPSPs.  
699 **(E-M)** Response to *in vitro* stimulation mimicking *in vivo* stimulation. Due to the sparse nature of action  
700 potentials *in vivo*, we developed a stimulation protocol to mimic the subthreshold stimulus encoding  
701 properties of L2/3 neurons *in vitro*. **(E)** Whole cell intracellular current clamp recordings were performed  
702 in L2/3 while L4 neurons were stimulated using a bipolar electrode. **(F)** Soma location of randomly selected  
703 neurons. **(G)** The stimuli were direct current injections with equal maximal amplitudes as the *in vivo*  
704 EPSCs, but the rising slope of the current was systematically reduced across the four stimulus conditions  
705 (see (Huang et al., 2016)). **(H-M)** L2/3 pyramidal neurons' responses to L4 stimulation. Each circle shows  
706 the average (over trials) response of one neuron (N=11). **(H-J)** EPSP response to *in vitro* stimulation. **H:**  
707 Onset time, **I:** Slope, **J:** EPSP amplitude; **(K-M)** Spike response to *in vitro* stimulation. **K:** Spike time, i.e.  
708 latency to spike after stimulus onset; **L:** Spike threshold, described as the membrane potential at which the  
709 second derivative reaches a global (positive) maximum; **M:** Action potential (i.e. spike) probability, across  
710 trials. **(N-P)** Spike versus EPSP response to *in vitro* stimulus. While both EPSP and spike parameters  
711 displayed an average dependence on the stimulus, EPSP parameters are more accurately determined by the  
712 stimulus than spike parameters on single trials.

713



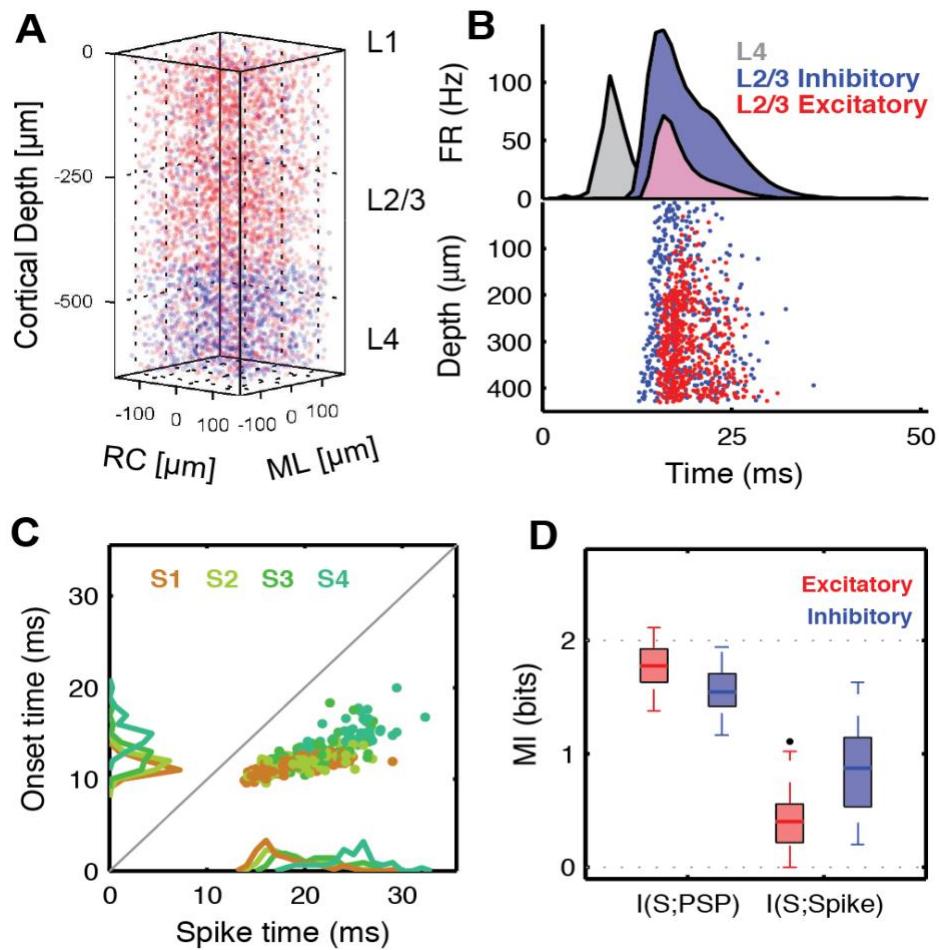
714

715 **Figure 2. Postsynaptic potentials encode substantially more stimulus information than spikes *in vitro***

716 (A) The information between PSPs and the stimulus is significantly higher than the information between  
 717 spikes and the stimulus. While the PSP contains a large fraction of the stimulus entropy (95%,  $I(S;PSP)$ ,  
 718  $1.81 \pm 0.31$  bit vs.  $H(S)$ ,  $1.86 \pm 0.17$  bit,  $p = 0.16$ ), most of this information is not transferred to the spike  
 719 ( $I(S;Spike)$ ,  $0.47 \pm 0.19$  bit, 24%). (B) The majority of the information in the PSP is carried by the onset  
 720 timing ( $Ot$ ,  $1.6 \pm 0.31$  bit, 85%), while slope ( $Sl$ ,  $0.17 \pm 0.09$  bit, 10%) amplitude ( $Am$ ,  $0.13 \pm 0.07$  bit, 5%)  
 721 carry only small amounts of information. (C)  $Ot$ ,  $Sl$ , and  $Am$  add their information independently, as the  
 722 synergy between them is close to 0 (Synergy :  $0.03 \pm 0.18$  bit,  $p = 0.15$ , t-test). (D) The information in the  
 723 spike is contributed by spike time ( $St$ ,  $0.31 \pm 0.13$  bit, 16%) and threshold ( $Vt$ ,  $0.12 \pm 0.08$  bit, 6.2%), and  
 724 jointly only reach 21% of the total information (repeated from A). (E) Substantial information transfer  
 725 occurs between the PSP and the spike, although this constitutes only 22% ( $St$ ) and 15% ( $Vt$ ) of the entropy

726 in the PSP. (F) The information in the properties of the PSP adds largely independently to the joint  
727 information, with a small but highly significant synergistic contribution of different PSP properties (0.41  
728  $\pm 0.13$  bit, 6.4%,  $p < 10^{-5}$ ). In all figures data is plotted as inter-quartile intervals and red lines denote the  
729 median of each distribution. Outliers are plotted as red dots. The dotted line denotes the maximal stimulus  
730 entropy.

731



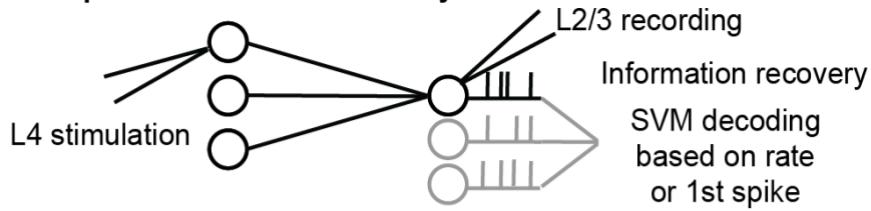
732

733 **Figure 3. Anatomically constrained barrel column *in silico* reproduces the relationships between sub-  
734 and supra-threshold information**

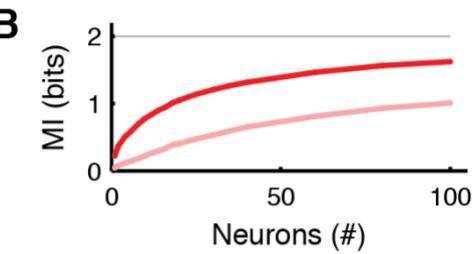
735 (A) An anatomically based model of a barrel column for L2/3/4 was generated to analyze the information  
736 transfer between L4 and L2/3 in analogy to the physiological recordings (see Huang et al. (2020) for  
737 details). (B) In response to stimulation in L4 with a whisker-like PSTH (grey), excitatory (red) and  
738 inhibitory (blue) cells respond in L2/3, with inhibitory activity eventually extinguishing the total activity in  
739 the network. (C) Corresponding to the *in vitro/in vivo* data, the timing of PSPs for a given stimulus is more

740 precise than the spikes they evoke (compare to Fig. 1L). **(D)** The relationship between PSPs and spikes in  
741 terms of timing and reliability leads to single cell mutual information very similar to the recorded data  
742 (excitatory cells, compare to Fig.2A). Inhibitory cells (not recorded), show less information in their PSP  
743 response, but more information in the spikes (all properties combined for both cell-types). Dotted line:  
744 stimulus entropy.

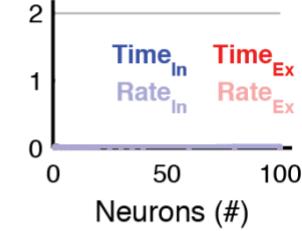
### A Population Information Analysis



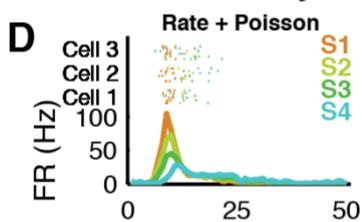
**B L2/3 in vitro**



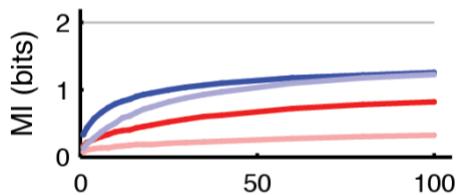
**C Validity**



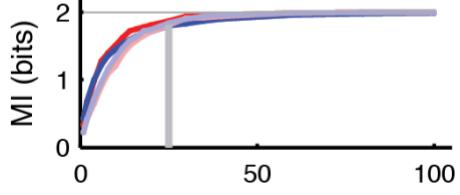
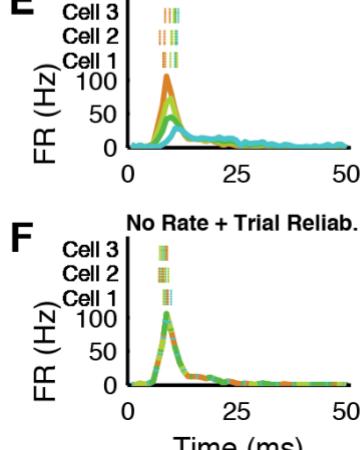
**D L4 Activity**



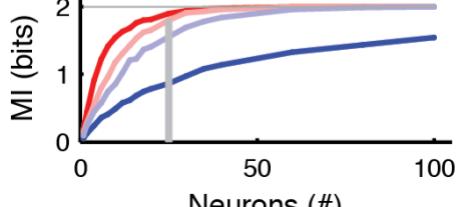
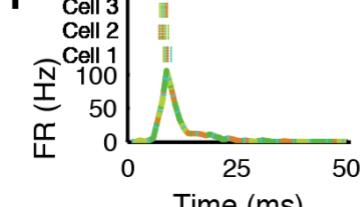
**E L2/3 Decoding**



**F Rate + Trial Reliab.**

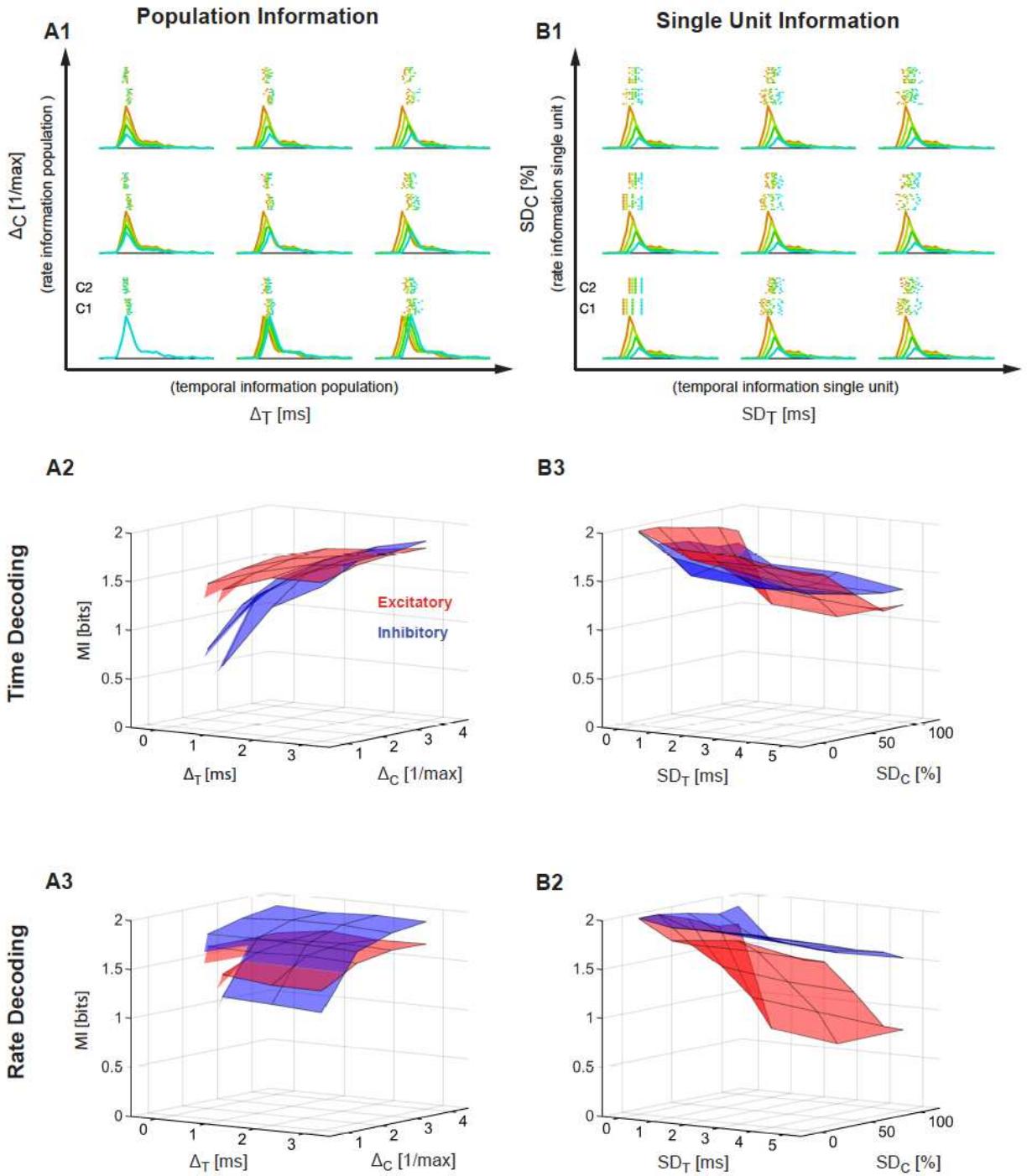


**G No Rate + Trial Reliab.**



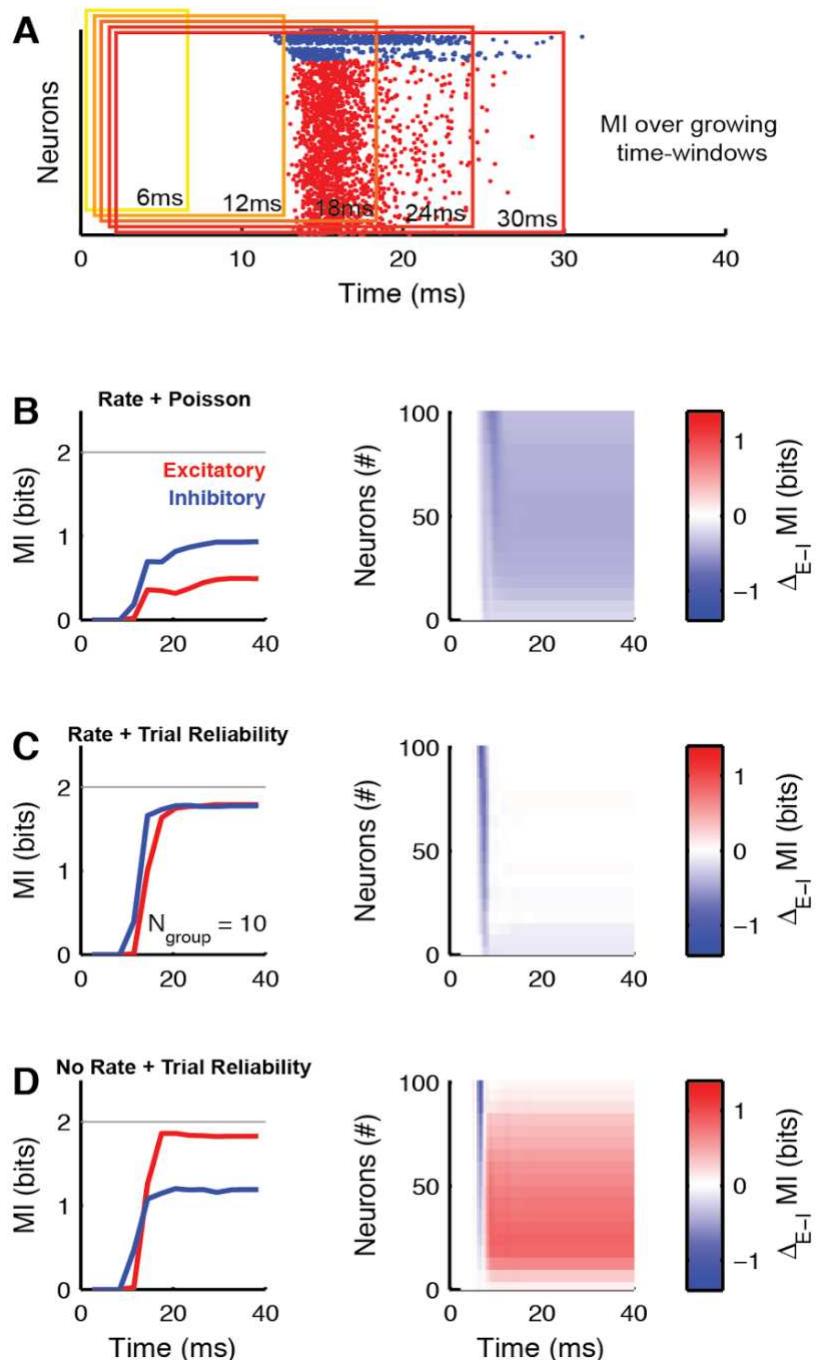
746 **Figure 4. Information recovery in neural populations recorded *in vitro*.**

747 If a postsynaptic EPSP carries near complete information about the stimulus in the periphery  
748 (Figure 2), how does the postsynaptic neuron reconstruct this information from poorly informative  
749 action potentials of the presynaptic neurons? (A) To address this question we evaluate the mutual  
750 information from population spike trains of groups of excitatory or inhibitory neurons. To prevent  
751 the sampling bias, MI is estimated between the stimulus and an SVM decoding from the population  
752 response. (B) Population information estimated from bootstrapped *in vitro* recordings show nearly  
753 complete recovery of stimulus information. Asymptote is reached above 81% for 100 neurons for  
754 temporal decoding (dark red), and remains systematically lower for the rate-based decoding (light  
755 red). (C) Estimating population information for non-informative stimuli (identical PSTH, Poisson-  
756 spiking) leads to vanishingly low MI values, demonstrating that the analysis does not introduce a  
757 positive bias. (D) If the population activity in L4 is only constrained by the PSTH and otherwise  
758 spikes are drawn according to Poisson-distributions (bottom left, different colors = different  
759 stimuli), then inhibitory neurons carry more information for both time (dark blue) and rate (light  
760 blue) decoding, than excitatory neurons (dark & light red respectively). The gray line denotes the  
761 entropy of the stimulus. (E) If PSTHs differ across stimuli but spike timing is stereotypical across  
762 trials ('Rate + Trial Reliability', top left, multiple trials per neuron above each other), coding  
763 becomes highly effective and independent of the cell-type and coding strategy (~25 cells). (F) If  
764 L4 PSTHs do not distinguish stimuli, but only the timing of individual neurons across trials is  
765 stereotypical (No Rate + Trial Reliability, top left), a remarkable shift occurs, with excitatory  
766 neurons reaching almost complete information for much smaller group sizes (~25 cells). In all  
767 plots the vertical grey line indicates where 90% of the information is represented.  
768



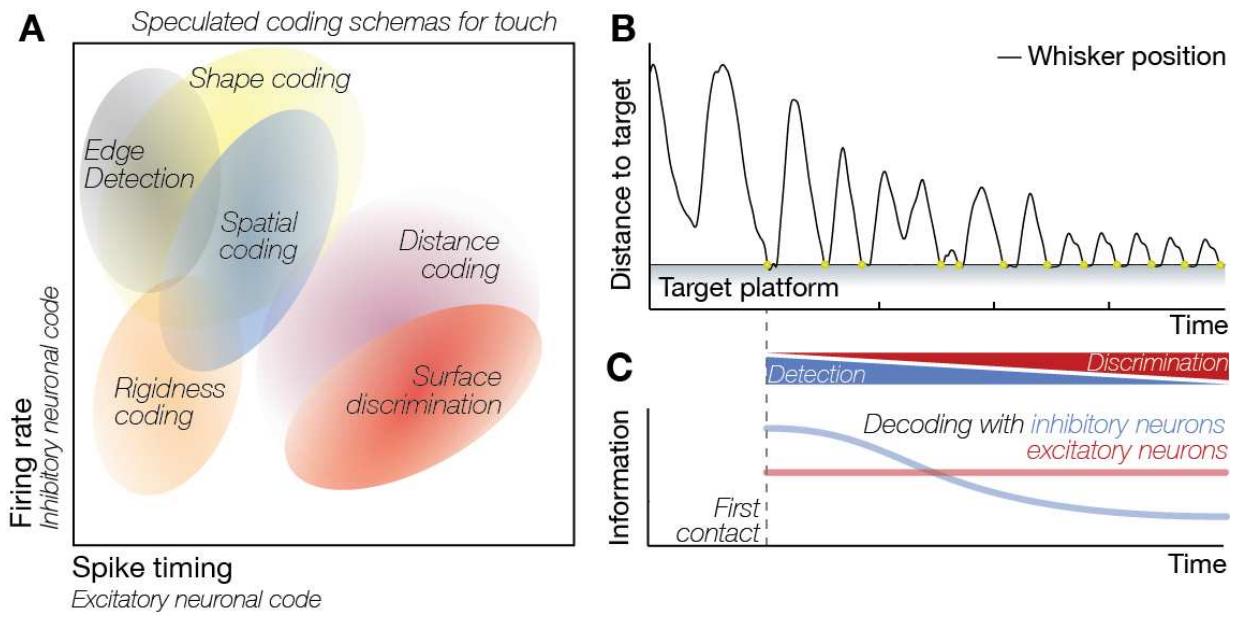
770 **Figure 5. Stimulus encoding by presynaptic single neurons and populations of neurons selectively**  
771 **influences the decoding performance of the postsynaptic excitatory or inhibitory neurons,**  
772 **respectively.**

773 (A1) Stimulus information can be encoded in differences in rate or timing on the level of the  
774 population PSTH. Different combinations of these two coding dimensions are varied, with  $\Delta_T$   
775 (abscissa) indicating different timing for different stimuli (different colors, see Fig. 4), and  $\Delta_C$   
776 (ordinate) indicating different rates for different stimuli. Maximal information is achieved for high  
777 values of  $\Delta_T$  and  $\Delta_C$ . For each condition the population PSTHs and two example cells are shown  
778 (raster plot for 10 trials, above). Spike-times of individual neurons are Poisson-distributed given  
779 the PSTH. NB C1 and C2 denote the responses of two different example cells. (A2) Decoding of  
780 first spike timing reveals a greater sensitivity of inhibitory neurons (blue) to the level of  
781 information in the L4 population response, both for time and rate information in L4. Conversely,  
782 excitatory neurons (red) are comparatively insensitive. (A3) Decoding of rate again reveals a  
783 greater sensitivity of inhibitory neurons to the level of information in the L4 response for different  
784 rates. Since we did not limit the time window of analysis, neither of the cell types is influenced by  
785 variation in time, while leaving the rate information unchanged. (B1) Stimulus information can  
786 also be encoded in the reliable discharge of single units. We modulated the reliability by  
787 introducing variability in timing ( $SD_T$ ) or variability in count ( $SD_C$ ) independent of each other.  
788 Maximal information is achieved for  $SD_T$  and  $SD_C$  both close to 0, i.e. perfectly reliable responses.  
789 Colors and raster plots as in A1. (B2) Decoding of first spike timing reveals a great sensitivity of  
790 excitatory neurons to the L4 information in single unit responses for both variability in time ( $SD_T$ )  
791 and rate ( $SD_C$ ). (B3) Decoding of rate shows a very strong sensitivity of excitatory neurons on the  
792 single unit information. Conversely, inhibitory neurons exhibit almost no sensitivity to single unit  
793 information in L4, and are thus dominated by L4 population information.



795 **Figure 6. Neuronal information recovery is completed in <20ms after stimulus onset.**

796 **(A)** Time-scales of information recovery computed by calculating the MI between stimulus (not shown)  
797 and spike trains (single trial example shown) over time-windows of increasing lengths (6-30ms at 3ms  
798 steps) **(B)** For the ‘Rate + Poisson’ encoding in L4, both excitatory (red) and inhibitory (blue) neurons in  
799 L2/3 reach their respective maximal information (left: group size of 10 cells, the gray line denotes the  
800 entropy of the stimulus), ~25-30ms after stimulus onset. L2/3 inhibitory neurons (blue) encode more  
801 information, independent from the peri-stimulus time and group size (right). The color code shows the  
802 difference in MI between the excitatory and inhibitory groups, with red for larger MI in the excitatory, and  
803 blue for greater MI for the inhibitory neurons. **(C)** For the ‘Rate + Trial Reliability’ condition in L4, the  
804 information content of the two populations is quite similar (left) with a slight advantage for the inhibitory  
805 neurons at early times, but no dependence on group size (right). **(D)** In the ‘No Rate - Trial Reliability’  
806 case, the divergence between information content only begins around 12ms after stimulus onset, after which  
807 excitatory neurons achieve a substantial coding advantage, especially for smaller group sizes.



808

809 **Figure 7. Multiplexed coding of touch.**

810 If intracellular information transfer, i.e. from EPSP-to-spike, results in a significant loss of near complete  
811 information, originally available in a single EPSP (Fig. 2), and if this information is recovered in local  
812 networks (Fig. 4 and 5) before the next sensory stimulus arrives (Fig. 6) using the rate and timing of spikes  
813 at the single cell and population levels (Fig. 4-6), selective decoding of stimulus properties by excitatory  
814 and inhibitory neural populations (Fig. 5) will result in a multiplexed code for sensory processing. **(A)** If  
815 excitatory and inhibitory neurons preferentially decode the stimulus information from the spike timing of  
816 individual neurons and the population rate of presynaptic neuronal activity (Fig. 5), respectively,  
817 information content of the activity in excitatory and inhibitory neurons should vary predictably – see the  
818 suggested coding schema for touch. **(B-C)** The information content across the neural populations will also  
819 vary depending on the complexity of the stimulus. **(B)** During tactile object localization in freely behaving  
820 animals (Celikel and Sakmann, 2007; Voigts et al., 2015, 2008), for example, as the animal approaches the  
821 tactile target and makes multiple contacts, the information content will change not only because the  
822 kinematics of touch varies, e.g. the amplitude whisker deflections is reduced to match the predicted position  
823 of the sensory target (Voigts et al., 2015), but also the neurons will represent different features of the sensory

824 target. (C) We speculate that information in the inhibitory neurons will better predict the stimulus location,  
825 although the information content of the excitatory neurons will eventually supersede as surface features are  
826 encoded with the subsequent contacts with the target.

827

828

829 **References**

830 Alenda A, Molano-Mazón M, Panzeri S, Maravall M. 2010. Sensory input drives multiple  
831 intracellular information streams in somatosensory cortex. *The Journal of Neuroscience*  
832 **30**:10872–84. doi:10.1523/JNEUROSCI.6174-09.2010

833 Allen CB, Celikel T, Feldman DE. 2003. Long-term depression induced by sensory deprivation  
834 during cortical map plasticity in vivo. *Nature neuroscience* **6**:291–9. doi:10.1038/nn1012

835 Amarasingham A, Chen T-L, Geman S, Harrison MT, Sheinberg DL. 2006. Spike Count  
836 Reliability and the Poisson Hypothesis. *J Neurosci* **26**:801–809.  
837 doi:10.1523/JNEUROSCI.2948-05.2006

838 Amarasingham A, Harrison MT, Hatsopoulos NG, Geman S. 2011. Conditional modeling and  
839 the jitter method of spike resampling. *Journal of Neurophysiology* **107**:517–531.  
840 doi:10.1152/jn.00633.2011

841 Arabzadeh E, Zorzin E, Diamond ME. 2005. Neuronal Encoding of Texture in the Whisker  
842 Sensory Pathway. *PLOS Biology* **3**:e17. doi:10.1371/journal.pbio.0030017

843 Avermann M, Tomm C, Mateo C, Gerstner W, Petersen CCH. 2012. Microcircuits of excitatory  
844 and inhibitory neurons in layer 2/3 of mouse barrel cortex. *Journal of Neurophysiology*  
845 **107**:3116–3134. doi:10.1152/jn.00917.2011

846 Azarfar A, Calcini N, Huang C, Zeldenrust F, Celikel T. 2018. Neural coding: A single neuron's  
847 perspective. *Neuroscience & Biobehavioral Reviews* **94**:238–247.  
848 doi:10.1016/j.neubiorev.2018.09.007

849 Barth AL, Poulet JFA. 2012. Experimental evidence for sparse firing in the neocortex. *Trends in  
850 Neurosciences* **35**:345–355. doi:10.1016/j.tins.2012.03.008

851 Bialek W, Rieke F. 1992. Reliability and information transmission in spiking neurons. *Trends in  
852 Neurosciences* **15**:428–434. doi:10.1016/0166-2236(92)90005-S

853 Bialek W, Rieke F, de Ruyter van Steveninck RR, Warland D. 1991. Reading a Neural Code.  
854 *Science* **252**:1854–1857.

855 Billings G, Piasini E, Lorincz A, Nusser Z, Silver RA. 2014. Network Structure within the  
856 Cerebellar Input Layer Enables Lossless Sparse Encoding. *Neuron* **83**:960–974.  
857 doi:10.1016/j.neuron.2014.07.020

858 Blatow M, Rozov A, Katona I, Hormuzdi SG, Meyer AH, Whittington MA, Caputi A, Monyer  
859 H. 2003. A Novel Network of Multipolar Bursting Interneurons Generates Theta  
860 Frequency Oscillations in Neocortex. *Neuron* **38**:805–817. doi:10.1016/S0896-  
861 6273(03)00300-3

862 Brecht M, Roth A, Sakmann B. 2003. Dynamic receptive fields of reconstructed pyramidal cells  
863 in layers 3 and 2 of rat somatosensory barrel cortex. *The Journal of physiology* **553**:243–  
864 65. doi:10.1113/jphysiol.2003.044222

865 Brecht M, Schneider M, Sakmann B, Margrie TW. 2004. Whisker movements evoked by  
866 stimulation of single pyramidal cells in rat motor cortex. *Nature* **427**:704–710.

867 doi:10.1038/nature02266  
868 Brette R. 2015. Philosophy of the spike: rate-based vs. spike-based theories of the brain.  
869 *Frontiers in Systems Neuroscience* **9**. doi:10.3389/fnsys.2015.00151  
870 Caputi A, Rozov A, Blatow M, Monyer H. 2009. Two calretinin-positive GABAergic cell types  
871 in layer 2/3 of the mouse neocortex provide different forms of inhibition. *Cereb Cortex*  
872 **19**:1345–1359. doi:10.1093/cercor/bhn175  
873 Celikel T, Sakmann B. 2007. Sensory integration across space and in time for decision making in  
874 the somatosensory system of rodents. *Proceedings of the National Academy of Sciences*  
875 **104**:1395–400. doi:10.1073/pnas.0610267104  
876 Celikel T, Szostak VA, Feldman DE. 2004. Modulation of spike timing by sensory deprivation  
877 during induction of cortical map plasticity. *Nature neuroscience* **7**:534–541.  
878 doi:10.1038/nn1222  
879 Chklovskii DB, Koulakov AA. 2004. MAPS IN THE BRAIN: What Can We Learn from Them?  
880 *Annual Review of Neuroscience* **27**:369–392.  
881 doi:10.1146/annurev.neuro.27.070203.144226  
882 Clem RL, Celikel T, Barth AL. 2008. Ongoing in vivo experience triggers synaptic  
883 metaplasticity in the neocortex. *Science* **319**:101–4. doi:10.1126/science.1143808  
884 Crochet S, Petersen CCH. 2006. Correlating whisker behavior with membrane potential in barrel  
885 cortex of awake mice. *Nature Neuroscience* **9**:608–610. doi:10.1038/nn1690  
886 Crochet S, Poulet JFA, Kremer Y, Petersen CCH. 2011. Synaptic Mechanisms Underlying  
887 Sparse Coding of Active Touch. *Neuron* **69**:1160–1175.  
888 doi:10.1016/j.neuron.2011.02.022  
889 Dalgleish HWP, Russell LE, Packer AM, Roth A, Gauld OM, Greenstreet F, Thompson EJ,  
890 Häusser M. 2020. How many neurons are sufficient for perception of cortical activity?  
891 *eLife* **9**:e58889. doi:10.7554/eLife.58889  
892 De Kock CPJ, Bruno RM, Spors H, Sakmann B. 2007. Layer- and cell-type-specific  
893 suprathreshold stimulus representation in rat primary somatosensory cortex. *The Journal  
894 of Physiology* **581**:139–154. doi:10.1113/jphysiol.2006.124321  
895 DeCharms RC, Zador AM. 2000. Neural representation and the cortical code. *Annual review of  
896 neuroscience* **23**:613–647.  
897 Denève S. 2008. Bayesian spiking neurons I: inference. *Neural Computation* **20**:91–117.  
898 doi:10.1162/neco.2008.20.1.91  
899 Denève S, Machens CK. 2016. Efficient codes and balanced networks. *Nature neuroscience*  
900 **19**:375–82. doi:10.1038/nn.4243  
901 Denève S, Pouget A. 2003. Basis functions for object-centered representations. *Neuron* **37**:347–  
902 59.  
903 Destexhe A, Rudolph M, Paré D. 2003. The high-conductance state of neocortical neurons in  
904 vivo. *Nature Reviews Neuroscience* **4**:739–51. doi:10.1038/nrn1198  
905 Diamond ME, Petersen RS, Harris JA. 1999. Learning through maps: functional significance of

906 topographic organization in primary sensory cortex. *J Neurobiol* **41**:64–68.

907 Doron G, von Heimendahl M, Schlattmann P, Houweling AR, Brecht M. 2014. Spiking

908 Irregularity and Frequency Modulate the Behavioral Report of Single-Neuron

909 Stimulation. *Neuron* **81**:653–663. doi:10.1016/j.neuron.2013.11.032

910 Eliasmith C, Anderson CH. 2002. Of neurons and engineersNeural Engineering. Computation,

911 Representation, and Dynamics in Neurobiological Systems. Cambridge, Massachusetts,

912 London, England: MIT Press. pp. 1–23.

913 Feldmeyer D, Lübke J, Sakmann B. 2006. Efficacy and connectivity of intracolumnar pairs of

914 layer 2/3 pyramidal cells in the barrel cortex of juvenile rats: Layer 2/3 pyramidal cell

915 synapses. *The Journal of Physiology* **575**:583–602. doi:10.1113/jphysiol.2006.105106

916 Feldmeyer D, Lübke J, Silver RA, Sakmann B. 2002. Synaptic connections between layer 4

917 spiny neurone- layer 2/3 pyramidal cell pairs in juvenile rat barrel cortex: physiology and

918 anatomy of interlaminar signalling within a cortical column. *The Journal of Physiology*

919 **538**:803. doi:10.1113/jphysiol.2001.012959

920 Finnerty GT, Connors BW. 2000. Sensory deprivation without competition yields modest

921 alterations of short-term synaptic dynamics. *PNAS* **97**:12864–12868.

922 doi:10.1073/pnas.230175697

923 Franke F, Fiscella M, Sevelev M, Roska B, Hierlemann A, Azaredo da Silveira R. 2016.

924 Structures of Neural Correlation and How They Favor Coding. *Neuron* **89**:409–422.

925 doi:10.1016/j.neuron.2015.12.037

926 Gao P, Ganguli S. 2015. On simplicity and complexity in the brave new world of large-scale

927 neuroscience. *Current Opinion in Neurobiology* **32**:148–155.

928 doi:10.1016/j.conb.2015.04.003

929 Gollisch T, Meister M. 2008. Rapid neural coding in the retina with relative spike latencies.

930 *Science* **319**:1108–1111. doi:10.1126/science.1149639

931 Harding-Forrester S, Feldman DE. 2018. Chapter 4 - Somatosensory maps In: Vallar G, Coslett

932 HB, editors. *Handbook of Clinical Neurology, The Parietal Lobe*. Elsevier. pp. 73–102.

933 doi:10.1016/B978-0-444-63622-5.00004-8

934 Harris KD, Mrsic-Flogel TD. 2013. Cortical connectivity and sensory coding. *Nature* **503**:51–8.

935 doi:10.1038/nature12654

936 Helmstaedter M, Staiger JF, Sakmann B, Feldmeyer D. 2008. Efficient Recruitment of Layer 2/3

937 Interneurons by Layer 4 Input in Single Columns of Rat Somatosensory Cortex. *Journal*

938 *of Neuroscience* **28**:8273–8284. doi:10.1523/JNEUROSCI.5701-07.2008

939 Holmgren C, Harkany T, Svensenfors B, Zilberter Y. 2003. Pyramidal cell communication

940 within local networks in layer 2/3 of rat neocortex. *The Journal of Physiology* **551**:139–

941 153. doi:10.1113/jphysiol.2003.044784

942 Houweling AR, Brecht M. 2008. Behavioural report of single neuron stimulation in

943 somatosensory cortex. *Nature* **451**:65–68. doi:10.1038/nature06447

944 Huang C, Resnik A, Celikel T, Englitz B. 2016. Adaptive Spike Threshold Enables Robust and

945 Temporally Precise Neuronal Encoding. *PLoS Computational Biology* **12**.  
946 doi:10.1371/journal.pcbi.1004984

947 Huang C, Zeldenrust F, Celikel T. 2020. Cortical representation of touch in silico. *bioRxiv*.  
948 doi:<https://doi.org/10.1101/371252>

949 Ince RAA, Mazzoni A, Petersen RS, Panzeri S. 2010a. Open source tools for the information  
950 theoretic analysis of neural data. *Front Neurosci* **3**. doi:10.3389/neuro.01.011.2010

951 Ince RAA, Panzeri S, Kayser C. 2013. Neural Codes Formed by Small and Temporally Precise  
952 Populations in Auditory Cortex. *J Neurosci* **33**:18277–18287.  
953 doi:10.1523/JNEUROSCI.2631-13.2013

954 Ince RAA, Senatore R, Arabzadeh E, Montani F, Diamond ME, Panzeri S. 2010b. Information-  
955 theoretic methods for studying population codes. *Neural Networks* **23**:713–727.  
956 doi:10.1016/j.neunet.2010.05.008

957 Izhikevich EM. 2004. Which model to use for cortical spiking neurons? *IEEE Transactions on*  
958 *Neural Networks* **15**:1063–1070.

959 Izhikevich EM. 2003. Simple Model of Spiking Neurons. *IEEE Transactions on Neural*  
960 *Networks* **14**:1572–1596. doi:10.1109/TNN.2003.820440

961 Johansson RS, Birznieks I. 2004. First spikes in ensembles of human tactile afferents code  
962 complex spatial fingertip events. *Nature Neuroscience* **7**:170–7. doi:10.1038/nn1177

963 Kaas JH. 1997. Topographic Maps are Fundamental to Sensory Processing. *Brain Research*  
964 *Bulletin* **44**:107–112. doi:10.1016/S0361-9230(97)00094-4

965 Kayser C, Ince RAA, Panzeri S. 2012. Analysis of Slow (Theta) Oscillations as a Potential  
966 Temporal Reference Frame for Information Coding in Sensory Cortices. *PLOS*  
967 *Computational Biology* **8**:e1002717. doi:10.1371/journal.pcbi.1002717

968 Kayser C, Logothetis NK, Panzeri S. 2010. Millisecond encoding precision of auditory cortex  
969 neurons. *PNAS* **107**:16976–16981. doi:10.1073/pnas.1012656107

970 Knudsen EI, Lac S, Esterly SD. 1987. Computational Maps in the Brain. *Annual Review of*  
971 *Neuroscience* **10**:41–65. doi:10.1146/annurev.ne.10.030187.000353

972 Ko H, Hofer SB, Pichler B, Buchanan K a, Sjöström PJ, Mrsic-Flogel TD. 2011. Functional  
973 specificity of local synaptic connections in neocortical networks. *Nature* **473**:87–91.  
974 doi:10.1038/nature09880

975 Kole K, Scheenen W, Tiesinga P, Celikel T. 2018. Cellular diversity of the somatosensory  
976 cortical map plasticity. *Neuroscience & Biobehavioral Reviews* **84**:100–115.  
977 doi:10.1016/j.neubiorev.2017.11.015

978 Li N, Chen T-W, Guo ZV, Gerfen CR, Svoboda K. 2015. A motor cortex circuit for motor  
979 planning and movement. *Nature* **519**:51–56. doi:10.1038/nature14178

980 Litwin-Kumar A, Doiron B. 2012. Slow dynamics and high variability in balanced cortical  
981 networks with clustered connections. *Nature neuroscience* **15**:1498–505.  
982 doi:10.1038/nn.3220

983 London M, Roth A, Beeren L, Häusser M, Latham PE. 2010. Sensitivity to perturbations in vivo

984 implies high noise and suggests rate coding in cortex. *Nature* **466**:123–127.  
985 doi:10.1038/nature09086

986 Lübke J, Roth A, Feldmeyer D, Sakmann B. 2003. Morphometric Analysis of the Columnar  
987 Innervation Domain of Neurons Connecting Layer 4 and Layer 2/3 of Juvenile Rat Barrel  
988 Cortex. *Cereb Cortex* **13**:1051–1063. doi:10.1093/cercor/13.10.1051

989 Magri C, Whittingstall K, Singh V, Logothetis NK, Panzeri S. 2009. A toolbox for the fast  
990 information analysis of multiple-site LFP, EEG and spike train recordings. *BMC*  
991 *neuroscience* **10**:81. doi:10.1186/1471-2202-10-81

992 Maheswaranathan N, Baccus SA, Ganguli S. 2018. Inferring hidden structure in multilayered  
993 neural circuits. *PLoS Computational Biology* **18**:e1006291. doi:doi.org/10.1101/120956

994 Margrie TW, Brecht M, Sakmann B. 2002. In vivo, low-resistance, whole-cell recordings from  
995 neurons in the anaesthetized and awake mammalian brain. *Pflugers Arch* **444**:491–498.  
996 doi:10.1007/s00424-002-0831-z

997 Marre O, Botella-Soler V, Simmons KD, Mora T, Tkačik G, Berry MJ. 2015. High Accuracy  
998 Decoding of Dynamical Motion from a Large Retinal Population. *PLoS Computational*  
999 *Biology* **11**:1–25. doi:10.1371/journal.pcbi.1004304

1000 Martens MB, Celikel T, Tiesinga PHE. 2015. A Developmental Switch for Hebbian Plasticity.  
1001 *PLOS Computational Biology* **11**:e1004386. doi:10.1371/journal.pcbi.1004386

1002 Mastrogiuseppe F, Ostojic S. 2017. Linking connectivity, dynamics and computations in  
1003 recurrent neural networks. *arXiv* 1711.09672. doi:10.1016/j.neuron.2018.07.003

1004 Narayanan RT, Egger R, Johnson AS, Mansvelder HD, Sakmann B, de Kock CPJ, Oberlaender  
1005 M. 2015. Beyond Columnar Organization: Cell Type- and Target Layer-Specific  
1006 Principles of Horizontal Axon Projection Patterns in Rat Vibrissal Cortex. *Cereb Cortex*  
1007 **25**:4450–4468. doi:10.1093/cercor/bhv053

1008 Nemenman I, Bialek W, de Ruyter van Steveninck R. 2004. Entropy and information in neural  
1009 spike trains: Progress on the sampling problem. *Phys Rev E* **69**:056111.  
1010 doi:10.1103/PhysRevE.69.056111

1011 Oberlaender M, de Kock CPJ, Bruno RM, Ramirez A, Meyer HS, Dercksen VJ, Helmstaedter M,  
1012 Sakmann B. 2012. Cell Type-Specific Three-Dimensional Structure of Thalamocortical  
1013 Circuits in a Column of Rat Vibrissal Cortex. *Cereb Cortex* **22**:2375–2391.  
1014 doi:10.1093/cercor/bhr317

1015 Panzeri S, Brunel N, Logothetis NK, Kayser C. 2010. Sensory neural codes using multiplexed  
1016 temporal scales. *Trends in Neurosciences* **33**:111–120. doi:10.1016/j.tins.2009.12.001

1017 Panzeri S, Diamond ME. 2010. Information carried by population spike times in the whisker  
1018 sensory cortex can be decoded without knowledge of stimulus time. *Front Synaptic*  
1019 *Neurosci* **2**. doi:10.3389/fnsyn.2010.00017

1020 Panzeri S, Petersen RS, Schultz SR, Lebedev M, Diamond ME. 2001. The role of spike timing in  
1021 the coding of stimulus location in rat somatosensory cortex. *Neuron* **29**:769–77.

1022 Panzeri S, Treves A. 1996. Analytical estimates of limited sampling biases in different

1023 information measures. *Network: Computation in Neural Systems* **7**:87–107.  
1024 doi:10.1080/0954898X.1996.11978656

1025 Peron SP, Freeman J, Iyer V, Guo C, Svoboda K. 2015. A Cellular Resolution Map of Barrel  
1026 Cortex Activity during Tactile Behavior. *Neuron* **86**:783–799.  
1027 doi:10.1016/j.neuron.2015.03.027

1028 Petersen CCH. 2019. Sensorimotor processing in the rodent barrel cortex. *Nature Reviews  
1029 Neuroscience* **20**:533–546. doi:10.1038/s41583-019-0200-y

1030 Petersen RS, Panzeri S, Diamond ME. 2002. Population coding in somatosensory cortex.  
1031 *Current Opinion in Neurobiology* **12**:441–447.

1032 Petersen RS, Panzeri S, Diamond ME. 2001. Population coding of stimulus location in rat  
1033 somatosensory cortex. *Neuron* **32**:503–14.

1034 Poulet JFA, Petersen CCH. 2008. Internal brain state regulates membrane potential synchrony in  
1035 barrel cortex of behaving mice. *Nature* **454**:881–885. doi:10.1038/nature07150

1036 Proville RD, Spolidoro M, Guyon N, Dugué GP, Selimi F, Isope P, Popa D, Léna C. 2014.  
1037 Cerebellum involvement in cortical sensorimotor circuits for the control of voluntary  
1038 movements. *Nature Neuroscience* **17**:1233–1239. doi:10.1038/nn.3773

1039 Quijan Quiroga R, Panzeri S. 2009. Extracting information from neuronal populations:  
1040 information theory and decoding approaches. *Nature reviews Neuroscience* **10**:173–85.  
1041 doi:10.1038/nrn2578

1042 Renart A, Rocha JD, Barthó P, Hollender L, Parga N, Reyes A, Harris KD. 2010. The  
1043 Asynchronous State in Cortical Circuits. *Science* **327**:587–590.  
1044 doi:10.1126/science.1179850.The

1045 Rosenblatt F. 1958. The perceptron: A probabilistic model for information storage and  
1046 organization in the brain. *Psychological Review* **65**:386–408.

1047 Seung HS, Yuste R. 2012. Neural Networks In: Kandel ER, Schwartz JH, Jessell TM,  
1048 Siegelbaum SA, Hudspeth AJ, editors. *Principles of Neural Science*. McGraw-Hill. pp.  
1049 1581–1600.

1050 Sriram B, Li L, Cruz-Martín A, Ghosh A. 2020. A Sparse Probabilistic Code Underlies the  
1051 Limits of Behavioral Discrimination. *Cereb Cortex* **30**:1040–1055.  
1052 doi:10.1093/cercor/bhz147

1053 Staiger JF, Bojak I, Miceli S, Schubert D. 2015. A gradual depth-dependent change in  
1054 connectivity features of supragranular pyramidal cells in rat barrel cortex. *Brain Struct  
1055 Funct* **220**:1317–1337. doi:10.1007/s00429-014-0726-8

1056 Stewart RS, Huang C, Arnett MT, Celikel T. 2013. Spontaneous oscillations in intrinsic signals  
1057 reveal the structure of cerebral vasculature. *Journal of Neurophysiology* **109**:3094–3104.  
1058 doi:10.1152/jn.01200.2011

1059 Stützgen MC, Schwarz C. 2010. Integration of Vibrotactile Signals for Whisker-Related  
1060 Perception in Rats Is Governed by Short Time Constants: Comparison of Neurometric  
1061 and Psychometric Detection Performance. *J Neurosci* **30**:2060–2069.

1062 doi:10.1523/JNEUROSCI.3943-09.2010  
1063 Thomson AM, Lamy C. 2007. Functional maps of neocortical local circuitry. *Front Neurosci* **1**.  
1064 doi:10.3389/neuro.01.1.1.002.2007  
1065 Ujfalussy BB, Makara JK, Branco T, Lengyel M. 2015. Dendritic nonlinearities are tuned for  
1066 efficient spike-based computations in cortical circuits. *eLife* **4**:e10056.  
1067 doi:10.7554/eLife.10056  
1068 Victor JD. 2009. Approaches to Information-Theoretic Analysis of Neural Activity. *Biological  
1069 Theory* **1**:302–316.  
1070 Voigts J, Herman DH, Celikel T. 2015. Tactile Object Localization by Anticipatory Whisker  
1071 Motion. *Journal of neurophysiology* **113**:620–632. doi:10.1152/jn.00241.2014  
1072 Voigts J, Sakmann B, Celikel T. 2008. Unsupervised whisker tracking in unrestrained behaving  
1073 animals. *Journal of neurophysiology* **100**:504–515. doi:10.1152/jn.00012.2008.  
1074 Weinberg RJ. 1997. Are Topographic Maps Fundamental to Sensory Processing ? *Brain  
1075 Research Bulletin* **44**:113–116. doi:10.1016/S0361-9230(97)00094-4  
1076 Wilent WB, Contreras D. 2004. Synaptic Responses to Whisker Deflections in Rat Barrel Cortex  
1077 as a Function of Cortical Layer and Stimulus Intensity. *Journal of Neuroscience*  
1078 **24**:3985–3998. doi:10.1523/JNEUROSCI.5782-03.2004  
1079 Zeldenrust F, Calcini N, Yan X, Bijlsma A, Celikel T. 2020. Cell type specific information  
1080 transfer for sparse coding. *bioRxiv* 2020.11.06.371658. doi:10.1101/2020.11.06.371658  
1081